

Carbon-13 NMR in Polymer Science

Carbon-13 NMR in Polymer Science

Wallace M. Pasika, EDITOR

Laurentian University

Based on a symposium

sponsored by the

Macromolecular Science

Division at the 61st

Conference of The Chemical

Institute of Canada in

Winnipeg, Manitoba,

June 4-7, 1979.

A C S S Y M P O S I U M S E R I E S **103**

AMERICAN CHEMICAL SOCIETY

WASHINGTON, D. C. 1979



Library of Congress \square Data

Carbon-13 NMR in polymer science.
(ASC symposium series; 103 ISSN 0097-6156)

Includes bibliographies and index.

1. Nuclear magnetic resonance spectroscopy—Congresses. 2. Carbon—Isotopes—Spectra—Congresses. 3. Polymers and polymerization—Spectra—Congresses.

I. Pasika, Wallace M. II. Chemical Institute of Canada. Macromolecular Science Division. III. Series: American Chemical Society. ACS symposium series; 103.

QD272.S6C37 547'.84 79-13384
ISBN 0-8412-0505-1 ACSMC8 103 1-344 1979

Copyright © 1979

American Chemical Society

All Rights Reserved. The appearance of the code at the bottom of the first page of each article in this volume indicates the copyright owner's consent that reprographic copies of the article may be made for personal or internal use or for the personal or internal use of specific clients. This consent is given on the condition, however, that the copier pay the stated per copy fee through the Copyright Clearance Center, Inc. for copying beyond that permitted by Sections 107 or 108 of the U.S. Copyright Law. This consent does not extend to copying or transmission by any means—graphic or electronic—for any other purpose, such as for general distribution, for advertising or promotional purposes, for creating new collective works, for resale, or for information storage and retrieval systems.

The citation of trade names and/or names of manufacturers in this publication is not to be construed as an endorsement or as approval by ACS of the commercial products or services referenced herein; nor should the mere reference herein to any drawing, specification, chemical process, or other data be regarded as a license or as a conveyance of any right or permission, to the holder, reader, or any other person or corporation, to manufacture, reproduce, use, or sell any patented invention or copyrighted work that may in any way be related thereto.

PRINTED IN THE UNITED STATES OF AMERICA

**American Chemical
Society Library**

1155 16th St. N. W.

Washington, D. C. 20036

In Carbon-13 NMR in Polymer Science; Pasika, W.;
ACS Symposium Series; American Chemical Society: Washington, DC, 1979.

ACS Symposium Series

Robert F. Gould, *Editor*

Advisory Board

Kenneth B. Bischoff

Donald G. Crosby

Robert E. Feeney

Jeremiah P. Freeman

E. Desmond Goddard

Jack Halpern

Robert A. Hofstader

James D. Idol, Jr.

James P. Lodge

John L. Margrave

Leon Petrakis

F. Sherwood Rowland

Alan C. Sartorelli

Raymond B. Seymour

Aaron Wold

Gunter Zweig

FOREWORD

The ACS SYMPOSIUM SERIES was founded in 1974 to provide a medium for publishing symposia quickly in book form. The format of the Series parallels that of the continuing ADVANCES IN CHEMISTRY SERIES except that in order to save time the papers are not typeset but are reproduced as they are submitted by the authors in camera-ready form. Papers are reviewed under the supervision of the Editors with the assistance of the Series Advisory Board and are selected to maintain the integrity of the symposia; however, verbatim reproductions of previously published papers are not accepted. Both reviews and reports of research are acceptable since symposia may embrace both types of presentation.

PREFACE

Carbon-13 NMR is opening up new vistas in polymer chemistry. Analytical information on polymeric systems, information regarding their structural, dynamic, and polymerization characteristics, can be provided by C-13 NMR. In many instances C-13 NMR complements already familiar characterization techniques or allows characterization to be carried out more readily. In other instances, only C-13 NMR can provide answers.

While organizing the symposium upon which this volume is based, the Macromolecular Science Division of the Chemical Institute of Canada attempted to include papers representing a wide range of applications for using Carbon-13 NMR to characterize polymers and to have both synthetic and biomacromolecular systems considered.

Many contributed to the success of the symposium. The executive and members of the Macromolecular Science Division thank the following firms for fiscal support: Polysar Limited; Abitibi Paper Company; Reichhold Limited; Xerox Research Centre of Canada Limited; Varian Associates of Canada; Glidden Company; Domtar Limited; DuPont of Canada; Shell Canada; Gulf Oil Canada; and the Dunlop Research Centre. A special vote of thanks is extended to the speakers at the symposium and to the authors for their excellent presentations and for their cooperation in "putting it together." J. Comstock of the American Chemical Society showed understanding and patience in bringing this volume to print.

Laurentian University
Sudbury, Ontario
Canada
March 26, 1979

WALLACE M. PASIKA

The Study of the Structure and Chain Dynamics of Polysulfones by Carbon-13 NMR

F. A. BOVEY and R. E. CAIS

Bell Laboratories, Murray Hill, NJ 07974

Sulfur dioxide does not homopolymerize but does participate in a rather wide variety of free radical copolymerizations with unsaturated monomers. The resulting polysulfones have been known for quite a long time. Solonina (1,2) obtained a white solid from the reaction of SO_2 with allyl ethers in 1898, but such products were not recognized as copolymers until the work of Marvel and Staudinger in the 1930's.

The class of vinyl monomers which will copolymerize with SO_2 is not clearly distinguishable in terms of fundamental structural characteristics from the class of those that will not. All terminal olefins, beginning with ethylene and continuing on up to the higher olefins, copolymerize and, with the exception of ethylene, give copolymers of strictly 1:1 alternating structure, since neither the chains ending in SO_2 nor those ending in olefin can add their own monomer; i.e. both reactivity ratios r_1 and r_2 are zero. Others include cyclopentene, cyclohexene, cycloheptene and some open-chain olefins with internal double bonds, provided the substituents at the double bond are not all larger than methyl.

Monomers which can add to their own radicals are capable of copolymerizing with SO_2 to give products of variable composition. These include styrene and ring-substituted styrenes (but not α -methylstyrene), vinyl acetate, vinyl bromide, vinyl chloride, and vinyl fluoride, acrylamide (but not N-substituted acrylamides) and allyl esters. Methyl methacrylate, acrylic acid, acrylates, and acrylonitrile do not copolymerize and in fact can be homopolymerized in SO_2 as solvent. Dienes such as butadiene and 2-chlorobutadiene do copolymerize, and we will be concerned with the latter compound in this discussion.

An important feature of olefin- SO_2 polymerizations is their relatively low ceiling temperatures: the reverse reaction becomes evident at quite low temperatures. In fact, it is while studying such copolymers that Dainton and Ivin in 1948 (3) first clearly recognized the existence of ceiling temperatures in vinyl polymers. The reversibility of the propagation has important effects on the composition of the chains, which, as we shall see, exhibits a very

0-8412-0505-1/79/47-103-001\$06.25/0
© 1979 American Chemical Society

strong dependence on temperature even at constant monomer feed ratio. This dependence invariably takes the form of tending to exclude SO_2 as the temperature is increased. This reversibility accounts also for some other unusual features of the chain structure, including deviation from first-order Markov statistics and, in the case of chloroprene, a seemingly anomalous and rather interesting tendency to form a more regular chain structure as the polymerization temperature is increased.

The use of nmr, --carbon-13 nmr in particular--, has given a much deeper insight into the structure of SO_2 copolymers than was possible by the older, traditional method of analytically determining monomer ratios in the polymer as a function of the monomer feed. In fact, it can be safely said that the use of nmr has completely revolutionized the study of copolymers. (The impact of nmr on copolymer studies is studiously ignored in all polymer textbooks, which tend to reflect the status of the field twenty years ago.)

Styrene- SO_2 Copolymers. I would now like to discuss two systems which illustrate the power of C-13 nmr in structural studies. The first is the styrene- SO_2 system. As already indicated, this is of the type in which the chain composition varies with monomer feed ratio and also with temperature at a constant feed ratio (and probably with pressure as well.) The deviation of the system from simple, first-order Markov statistics, --i.e. the Lewis-Mayo copolymerization equation--, was first noted by Barb in 1952 (4) who proposed that the mechanism involved complex formation between the monomers. This proposal was reiterated about a decade later by Matsuda and his coworkers (5,6). Such charge transfer complexes do in fact exist, with the olefin acting as the donor, but we shall see that it is not necessary to invoke complex formation to explain the observed kinetics. It is also unnecessary to invoke penultimate effects in the reaction of the propagating radicals.

The traditional compositional and kinetic measurements cannot distinguish effectively between the various models proposed to account for deviations from the simple copolymerization model. To do this requires monomer sequence data, and for this nmr is the method par excellence. But in order to make use of the potentially rich information provided by nmr, one must be able to make assignments of the resonances in spectra which are often quite complex. This is usually done by (1) isotopic labelling; (2) observing carbon multiplicity in the absence of decoupling; (3) relaxation measurements; (4) logical deduction from the spectra of a series of copolymers of varied composition; and (5) invoking chemical shift rules. In order to interpret the spectra of SO_2 copolymers, we must recognize certain features of the monomer sequences, the principal one of which is that they have a sense of direction. To avoid confusion, we must remember that for any particular sequence we are always looking at the central vinyl monomer unit in the β -to- α direction from left to right. A

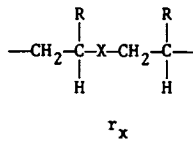
second, minor point is that since SO_2 units are not chiral, compositional triads are actually configurational dyads and compositional tetrads are configurational triads. These points are illustrated in Figures 1 and 2, where the chain sequences are represented as planar zigzags (viewed edge on). Figure 1 shows compositional triads involving the unit X, which in our case is SO_2 . As configurational dyads they may be racemic or meso of two types, depending on whether the chiral centers straddle an X unit or not. Also, we must note that XMM is not the same as MMX. In Fig. 2 the same representation is extended to compositional tetrads, i.e. configurational triads.

An important first step in interpreting the C-13 spectra is to distinguish α -carbons from β -carbons, i.e. methine from methylene. Observation of multiplicity when the proton decoupler is off is one way, but this is not always easy if the lines are broadened by chemical shift multiplicity. Measurement of T_1 has been used for this purpose since the β -carbon with two bonded protons relaxes about twice as fast as the α -carbon with only one. A very positive way is by deuterium labelling. In Fig. 3 is shown the main-chain 25 MHz carbon spectrum of two styrene- SO_2 copolymers containing 58 mol% styrene, or a ratio of styrene to SO_2 of 1.38 (7). In the bottom one, $\beta, \beta\text{-d}_2$ -styrene has been used, and all the β -carbon resonances are distinguishable from the α -carbon resonances since the presence of deuterium has eliminated their nuclear Overhauser effect; because of this and the deuterium J coupling (~ 20 Hz), they are markedly smaller and broader than the α -carbon resonances.

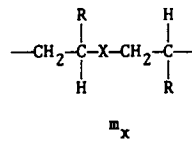
The assignments of resonances to particular monomer sequences are based primarily on their relative intensities as a function of overall composition. We define R as the overall ratio of styrene to SO_2 in the polymer. In Fig. 4 are shown the complete 25 MHz C-13 spectra, including at the left the aromatic carbons, for four copolymers of varied R (7). The spectrum of atactic polystyrene is also shown. The number of resonances shows that compositional triads are being distinguished: SMS, SMM, MMS, and MMM. Here, M stands for styrene and S for SO_2 . As the ratio R increases, MMM sequences become evident and can be assigned on the basis of the polystyrene spectrum. In the latter, the β -carbon is highly sensitive to configuration whereas the α -carbon is entirely insensitive, appearing as a narrow spike.

In Table 1 are shown the chemical shifts of the resonances in polystyrene sulfones. They are based on relative resonance intensities as a function of monomer ratio and on a consistent set of rules. We observe that the least shielded α - and β -carbons are those bonded directly to sulfone sulfur atoms. One can recognize very clearly a sulfone oxygen shielding effect when the carbon concerned is in the γ -position with relation to the oxygens. It appears to be analogous to the well known carbon γ -effect. Comparing α -MMS with α -SMS we see that for α -carbons it causes a shielding of about 5 ppm. Comparing β -SMM with β -SMS, we see a

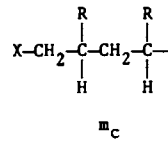
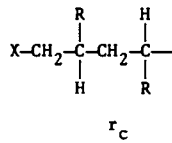
TRIADS:



MCM



XMM



MMX

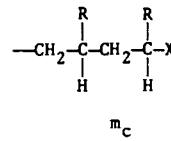
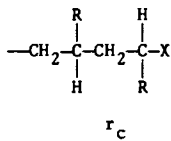


Figure 1. Compositional triads (configurational dyads) in chains of vinyl (M) copolymers with a comonomer which places a single atom X in the main chain

TETRADES:

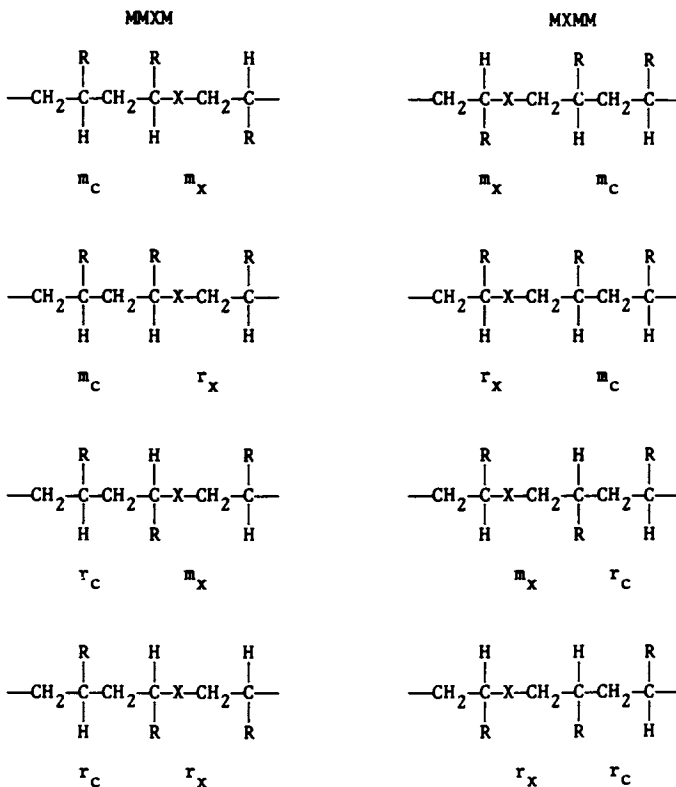


Figure 2. Compositional tetrads (configurational triads) in chains of vinyl (M) copolymers with a comonomer which places a single atom X in the main chain.

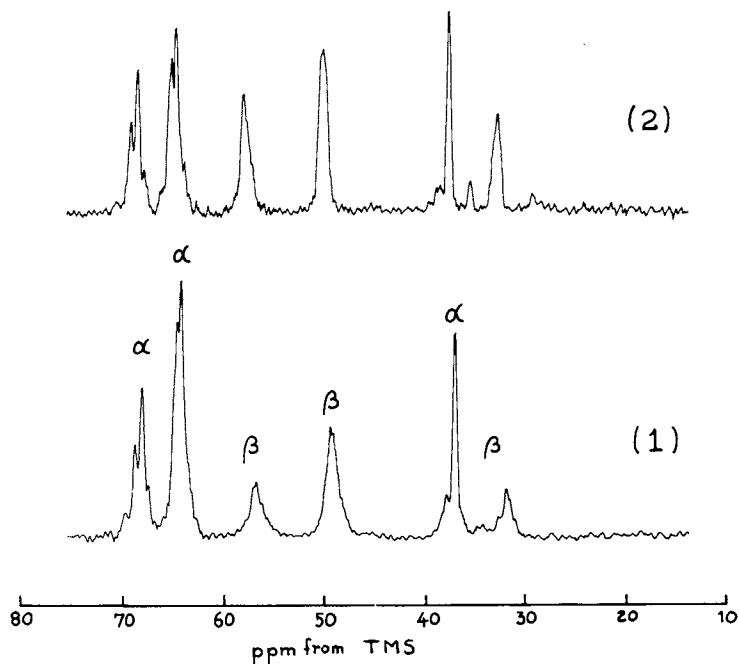


Figure 3. $C-13$ spectra (25 MHz) of the main chain of styrene- SO_2 copolymers containing 58 mol % styrene ($R = 1.38$). The bottom spectrum is of a copolymer with $\beta,\beta-d_2$ -styrene (25% solution in $CDCl_3$ at $55^\circ C$).

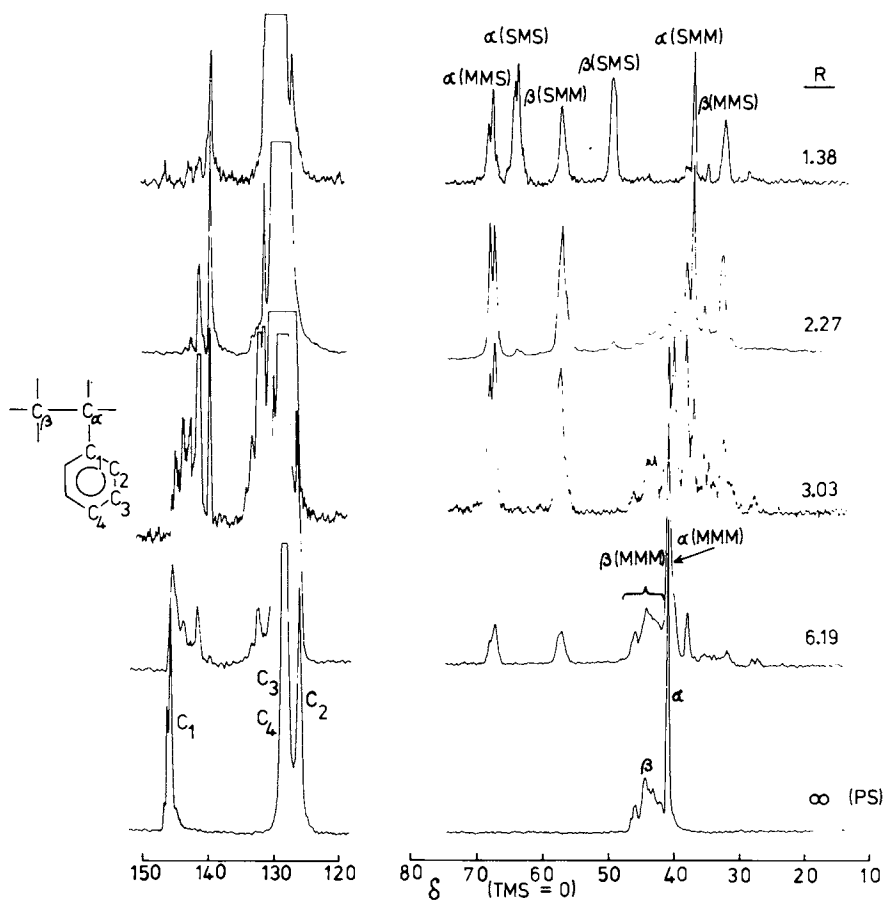
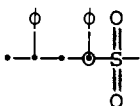
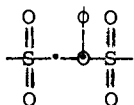
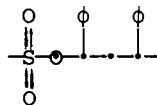
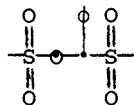
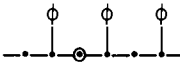
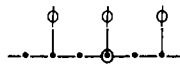
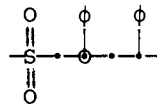
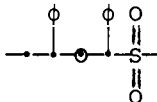


Figure 4. $C-13$ spectra (25 MHz) of styrene- SO_2 copolymers of four different compositions. The aromatic carbons are on the left and the main chain carbons on the right. R values are the mole ratios of styrene to SO_2 in the copolymers. The spectrum at the bottom is that of atactic PS. (All observed as 25% solutions in $CDCl_3$ at $55^\circ C$ except PS, which was observed as 20% solution in cyclohexane- d_{12} at $77^\circ C$.)

shielding of about 9 ppm for β -carbons. The same is true for β -MMM vs. β -MMS. We note that α -SMM and β -MMS are actually more shielded than polystyrene itself.

Table 1

Main Chain Carbon Chemical Shifts in Styrene-SO₂ Copolymers

		<u>Shift, ppm from TMS</u>
α -MMS		68
α -SMS		63
β -SMM		57
β -SMS		48
β -MMM		44
α -MMM		40
α -SMM		38
β -MMS		33

An interesting feature of the styrene-SO₂ system, --which indeed is true of all SO₂ copolymerizations with comonomers capable of homopolymerizing--, is the existence of a ceiling temperature above which the formation of alternating units, SMS, is forbidden. The number fraction of M sequences of length n is

given by;

$$N_M(n) = \frac{p(SM^nS)}{\sum_{n=1}^{\infty} p(SM^nS)}$$

i.e. the probability of occurrence of a length of n styrene units divided by the sum of the probabilities of all styrene sequences of all lengths. Since it can be shown that:

$$\sum_{n=1}^{\infty} p(SM^nS) = p(MS),$$

and since in the present case:

$$p(MS) = p(S),$$

because all sulfone units S have M as neighbors, we have

$$N_M(n) = \frac{p(SM^nS)}{p(S)}$$

For the number fraction of SMS sequences:

$$N_M(1) = \frac{p(SMS)}{p(S)}$$

A plot of $N_M(1)$ versus polymerization temperature is shown in Fig. 5. It will be seen that at low temperature it approaches unity, which would correspond to a strictly alternating structure, and that it declines very rapidly, reaching zero at about 40° . Therefore, above 40° alternating sequences are not generated. At 40° , it turns out that a chain ending in $-SM^\bullet$ will add another M and then add S , and so on, yielding a polymer with predominantly the regular structure $SMMMSMM$ etc. As the polymerization temperature is increased further, the SO_2 units are in effect squeezed out progressively until around 100° a second ceiling temperature occurs and no appreciable amount of SO_2 is incorporated; only polystyrene is produced. There is probably a moral here with regard to those monomers which, like methyl methacrylate, acrylonitrile, and acrylate esters, merely seem to homopolymerize in SO_2 as solvent. If polymerization were conducted at a sufficiently low temperature, SO_2 probably would be incorporated.

The copolymerization scheme we favor is as follows:



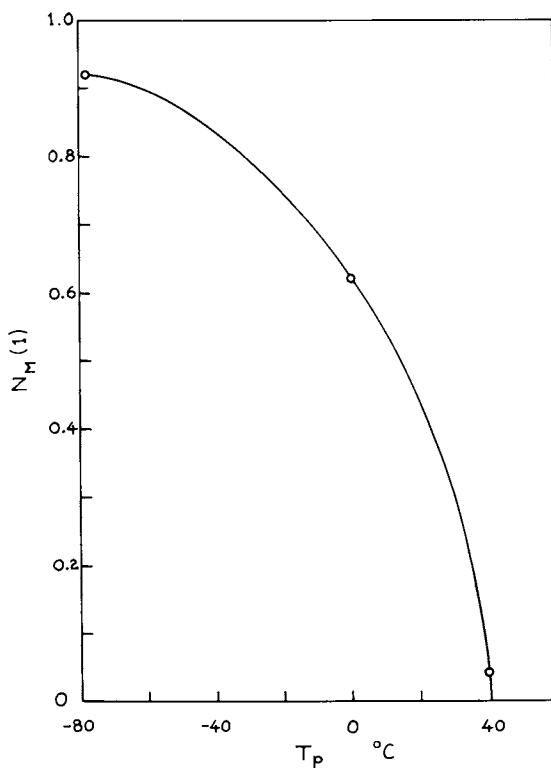


Figure 5. Plot of the number fraction of SMS sequences, $N_M(1)$ as a function of polymerization temperature for styrene- SO_2 copolymers



The K values are the equilibrium constants which describe the position of the propagation-depropagation equilibrium and are equal to the ratio of the propagation rate constant to the depropagation rate constant:

$$K = k_{\text{propag.}} / k_{\text{depropag.}}$$

$$K_1 = \frac{[\sim\text{SMS}\cdot]}{[\sim\text{SM}\cdot][\text{S}]}$$

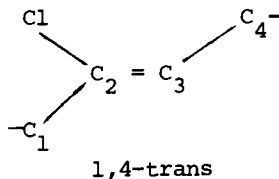
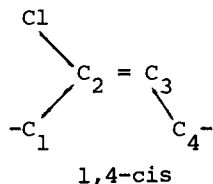
$$K_2 = \frac{[\sim\text{SM}\cdot]}{[\sim\text{SMS}\cdot][\text{M}]}$$

The formation of alternating copolymer is represented by the overall equilibrium constant

$$K_{\text{alt}} = K_1 K_2 = \frac{1}{[\text{S}][\text{M}]}$$

As K_{alt} approaches unity we approach the ceiling temperature for alternating copolymerization. At very low temperature, --for example -78° --, the alternating structure predominates because only S is reactive enough to add to $\sim\text{M}\cdot$ and $\sim\text{S}\cdot$, of course, can only add M. Above 40° , alternating sequences cannot survive.

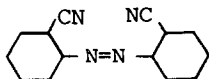
Chloroprene-SO₂ Copolymers. The second copolymer system to be discussed is the chloroprene-SO₂ system. This presents potential complications beyond that of the styrene-SO₂ system because the chloroprene may enter the chain in 1,4-cis, 1,4-trans, 1,2 or 3,4 fashion:





If it enters in 1,2 or 3,4 fashion, chiral centers are introduced and relative stereoisomerism must be considered. Actually, as it turns out, things are not so complicated as they might be because of a strong preference for one mode of addition. Complications are quite sufficient, however, as they include a head-to-head:head-to-tail option not normally open to vinyl monomers but available to unsymmetrically substituted dienes adding in 1,4-fashion. In fact, little progress can be made in structural determination except by use of C-13 nmr at superconducting frequencies.

We have prepared a series of copolymers under the conditions shown in Table 2. The monomer feed was always a 50:50 ratio of chloroprene to sulfur dioxide. Copolymerizations were carried out in bulk at temperatures from -78° to 100° . Initiators were tertiary butyl hydroperoxide at low temperatures, where it forms a redox system with the SO_2 and is more effective than one might otherwise expect. Silver nitrate was used at 0° and 25° , azoiso-butyrionitrile at 40° and 60° , and azodicyclohexanecarbonitrile



at 100° .

R values were calculated from elemental analysis for carbon, hydrogen, and chlorine. It can be seen again that temperature has a very marked effect on composition. Even at 100° , however, about 16 mol% sulfur dioxide is present. There was also produced a small quantity (1 to 10% of the amount of copolymer) of the cyclic addition product, 3-chloro-2,5-dihydrothiophene-1,1-dioxide, m.p. $99-100^{\circ}$.

We show in Scheme I the representation of the four chloroprene-centered sequences MMS, MMM, SMS, SMM (M as before represents monomer and S represents sulfur dioxide). The chloroprene carbons are numbered in the conventional manner. The first two carbons of chloroprene units which are neighbors of the central chloroprene unit or of the sequence are also represented. We will be concerned with all four carbons in all four sequences.

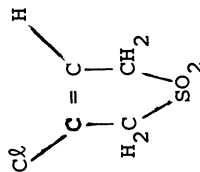
The spectrum of the polymer prepared at 40° , with an R value of 1.64, is shown in Fig. 6. The spectrum divides naturally into two parts: that corresponding to the olefinic carbons, C_2 and C_3 ,

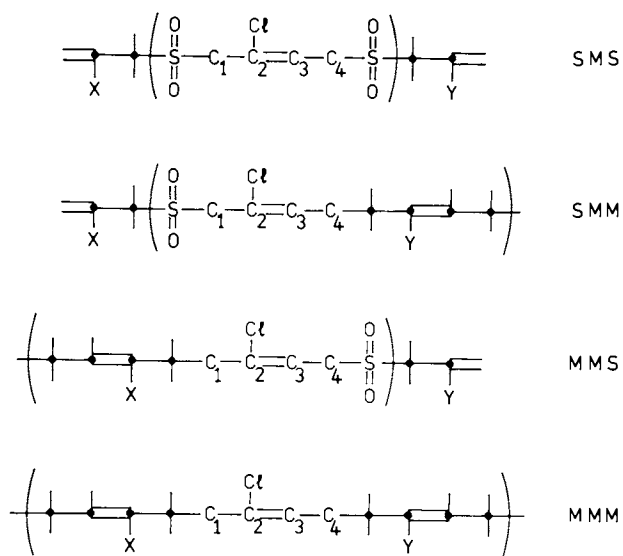
Table 2
Preparation of Chloroprene-SO₂ Copolymers; All in Bulk at 50:50 Mol Ratio

Temp. (°C)	Polym. Time (h)	Initiator	[Init.] (mol%)	Yield (wt%)	R	Notes
-78	17.25	t-BuOOH	0.46	9.8	1.15	sol. DMSO, DMF
-45	1.67	"	"	14.0	1.20	"
-17	6.25	"	"	15.4	1.20	2% sol. hot CHCl ₃
0	14.80	AgNO ₃	0.02	22.6	1.20	sol. DMSO, DMF
25	4.67	"	"	22.3	1.30	"
40	6.32	AIBN	0.05	22.1	1.64	"
60	3.30	"	"	32.1	2.44	49% gel, 0.9% CDTD ²
100	1.03	ACHCN ¹	"	15.4	5.08	52% gel, 10.1% CDTD

¹1,1-Azodicyclohexanecarbonitrile.

²3-Chloro-2,5-dihydro-thiophene-1,1-dioxide.





Scheme I. $X = Cl$ or H ; $Y = Cl$ or H .

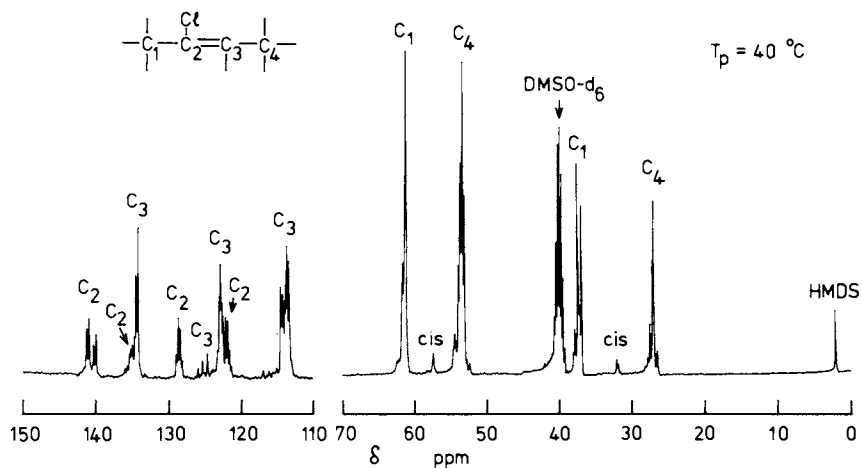


Figure 6. A 90-MHz C-13 spectrum of chloroprene-SO₂ copolymer prepared at 40°C and having an R value of 1.64

appears at the left between 110 and 150 ppm downfield from the hexamethyldisiloxane reference, and the part corresponding to the methylene carbons, C_1 and C_4 , appears at the right in the more shielded region of 25-65 ppm. We shall first assign the carbons, then the compositional sequences, and finally concern ourselves with head-to-head:head-to-tail isomerism.

The olefinic carbons C_2 and C_3 were assigned by (a) a partially relaxed dynamic experiment in which by proper choice of pulse interval the C_2 carbon resonances, having longer T_1 , remained as inverted signals while the C_3 carbons recovered to positive values; and (b) from the proton-coupled spectrum, in which the C_3 resonances were split to doublets.

In the methylene region the C_1 and C_4 resonances at higher field occur in the same position as for polychloroprene and the assignments were taken from the work of Coleman *et al.* (8). The methylene carbons at lower field have no precedent in the polychloroprene spectrum and were assigned by coherent irradiation of each of the corresponding protons and noting which carbon gave the larger Overhauser effect.

We must now consider the nature of the chloroprene units. We have represented them as 1,4 and we further deduce that they are trans. If 1,2 or 3,4 units were present the pendant vinyl groups, having two protons on C_1 and C_4 , would yield triplets in the olefinic region when the decoupler is off. No such triplets are observed, as so these structures can be eliminated. This also means the elimination of asymmetric carbons and the complications to which they can give rise.

The answer to the cis-trans question is to be found in the methylene carbon spectrum of Fig. 7. If we look at the C_1 (61 ppm) and C_4 (53 ppm) peaks for the -78° polymers, --which we recall has an almost exclusively alternating structure--, we see that they are clearly split, but by less than 1 ppm. We might at first think this represents cis and trans structures. However, experience with diene polymer spectra shows that when methylene carbons are involved in a cis structure they shield each other by 8 to 10 ppm. This is due to the operation of the γ steric effect, particularly strong when the carbon bonds actually eclipse each other rather than being merely gauche. In chloroprene units one expects the C_4 carbon to shift little between a cis and trans structure because it always sees a bulky substituent across the double bond. The C_1 carbon, however, sees such a substituent only in the cis structure, in which it should accordingly be upfield from the trans carbon. We believe the small peaks at 32 and 57 ppm represent the cis structure, as reflected in the C_1 resonances. It can be clearly seen that trans strongly predominates. This assignment is also confirmed by the trans C-C double bond stretching frequency at 1660 cm^{-1} ; cis would be expected at 1652 cm^{-1} .

In Fig. 8 are shown the olefin carbon spectra of four copolymers as a function of temperature, including that shown in Fig. 6. As we have seen, eight groups of resonances are recognizable.

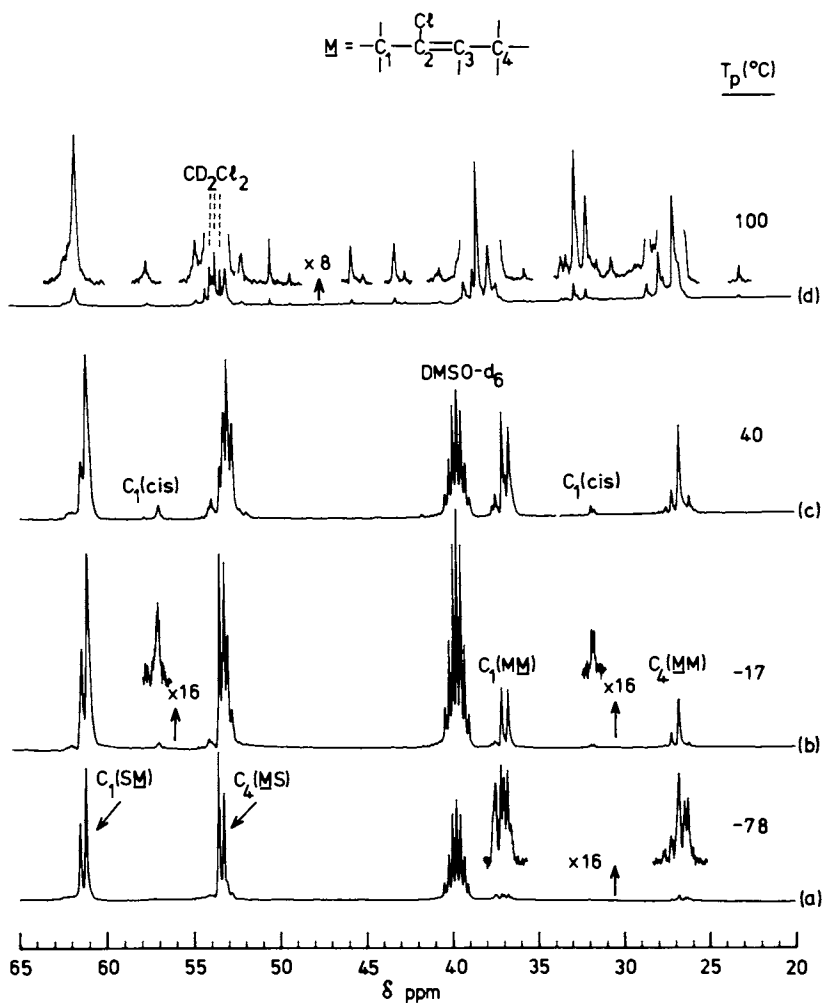


Figure 7. A 90-MHz spectra of methylene carbons in four copolymers prepared at different temperatures. R Values are (top to bottom) 5.08, 1.64, 1.20, and 1.15. The top spectrum was run in CD_2Cl_2 and the others in d_6 -DMSO.

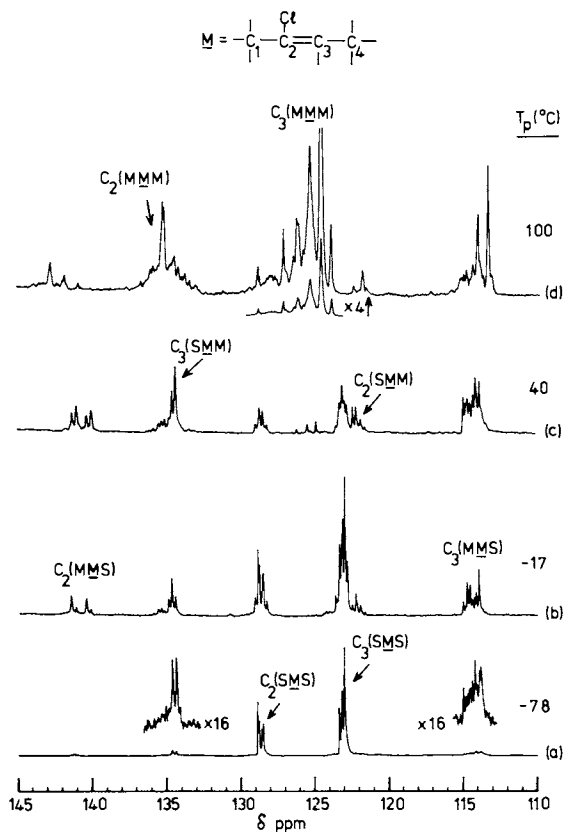


Figure 8. A 90-MHz spectra of olefin carbons in four copolymers prepared at different temperatures. R values are (top to bottom) 5.08, 1.64, 1.20, and 1.15. The top spectrum was run in CD_2Cl_2 and the others in d_6 -DMSO.

This means we are dealing with four types of compositional triads (two types of carbons in each, C_2 and C_3): MMS, MMM, SMS, and SMM. The olefinic carbons look both ways along the chain and can tell what the neighboring monomer units are on both sides.

In the polymer prepared at -78° , the R value is nearly one and so almost the only sequence present is SMS. This enables this assignment to be made with confidence. The homopolymer type sequence, --MMM--, can be assigned by reference to the paper of Coleman, *et al.* (8) on polychloroprene, to which we have already made reference. The sequences MMS and SMM can be assigned as shown by appealing to chemical shift rules similar to those we have already discussed for polystyrene sulfone:

(a) A sulfone in the β position to a carbon exerts a shielding effect of -11 ppm, owing to the operation of the oxygen " γ -effect".

(b) A sulfone in the γ -position to a carbon exerts a deshielding effect of $+8$ ppm, arising from what may be termed an oxygen " δ -effect".

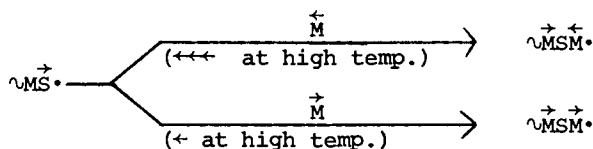
We then expect C_2 in MMS to be 11 ppm downfield from C_2 in SMS. Similarly C_3 in MMS should be 8 ppm upfield from C_3 in SMS. There are peaks in the expected positions. Assignments for SMM follow by default.

Sequence assignments in the methylene region (Fig. 7) are made considerably simpler by the fact that only compositional dyads rather than triads can be discriminated here. The C_1 carbons of the central chloroprene unit can distinguish units to the left but cannot distinguish those to the right, which are too far away. Similarly, C_4 carbons can distinguish units to the right but not to the left. The upfield pair of C_1 and C_4 resonances are in the positions corresponding to chloroprene homopolymers, whereas the downfield pair of resonances represent C_1 and C_4 carbons next to sulfone groups. Again, assignments are aided by recalling that alternating sequences predominate in the low temperature copolymers.

We note minor but complex resonances in both the olefinic and methylene spectra of the -78° temperature polymer. These arise in part from residual MM sequences but mainly from admixture of polymer formed at higher temperatures in the course of working up the product. We may note also in Fig. 7 that the proportion of *cis* 1,4 structures, as seen in the C_1 resonances, increases with polymerization temperature.

Let us finally consider the last form of isomerism with which we must deal. Geometrical isomerism and monomer sequence structure appear to be understood. However, there are clearly still splittings in both the olefinic and methylene spectra that have not been accounted for. This is evident even in the otherwise quite regular alternating structure of the -78° polymer, and must be due to head-to-tail head-to-head isomerism of the chloroprene units in the manner illustrated earlier. It is known that substantial proportions of head-to-head tail-to-tail units occur in

polychloroprene. The C_1 methylene carbons in SM dyads seem to reflect only this form of isomerism, being unresponsive to others. As we look at the behavior of this resonance as a function of temperature, we note a surprising thing: as the polymerization temperature increases it goes from a doublet to a singlet (Fig. 7). Thus, in this respect, but in no other, the chain becomes more regular with increasing temperature rather than less. It has so far been universally observed that polymer microstructure behaves the opposite way, becoming more irregular with increasing temperature. Why the difference in this case? We believe that the possibility of depropagation reactions explains this behavior. At low temperatures, forward propagation is favored; both head-to-tail and head-to-head propagation are nearly equally favored, head-to-tail being slightly preferred. This is not really the thermodynamically preferred ratio of structures, which is evidently almost exclusively head-to-tail, but nothing can be done to correct this as the structure is, so to speak, trapped. As the temperature is increased, however, the depropagation reaction comes more and more into play. A monomer which comes in the "wrong" way, i.e. head-to-head, gets a further chance to go out and come back in again.



The depropagation reaction at the top is favored over that at the bottom. Note that this explanation applies only to MSM sequences. Such a corrective device is not open to MM sequences, in which we may expect the fraction of head-to-head to increase with temperature in the "normal" manner.

I have discussed two cases of chain microstructure determination in SO_2 copolymers. First, the styrene- SO_2 system, which exhibits the general kinetic and compositional behavior of such systems, particularly as a function of polymerization temperature. Second, and considerably more complicated, is the chloroprene- SO_2 system. This one represents the limit of what can be handled and more or less completely solved at the present time. To do so requires about all our resources: ^{13}C at superconducting frequencies and a variety of strategies for carbon type assignment, compositional assignment, and determination of the mode of addition of the chloroprene units.

Polysulfone Chain Dynamics. Carbon-13 nmr has been used recently to resolve a puzzling problem in the dynamics of polysulfone chains (9,10,11). Some years ago, Bates, Ivin and Williams (12) reported that measurements of the dielectric dispersion in solutions of alternating 1:1 copolymers of sulfur dioxide with hexene-1 and 2-methylpentene-1 show no loss in the high frequency region, i.e.

beyond 1 MHz. A high frequency loss region is to be expected for flexible polymers having electric dipole components perpendicular to the main chain direction. It was further found that the frequency of maximum loss was inversely proportional to the degree of polymerization raised to the power 1.5-2.0, being for example about 24 kHz in benzene at 25° for polyhexene-1 sulfone having a number average molecular weight of 210,000. From these observations it was concluded that the presence of sulfone groups so stiffens the chain that overall tumbling is the only motion effective in relaxing the molecule in an oscillating electric field. This conclusion is quite surprising as molecular models and comparison to other polymers of comparable glass temperature (polystyrene and polymethyl methacrylate) do not suggest such a very high degree of steric hindrance. Carbon-13 relaxation measurements enable one to obtain at least an approximate measure of the chain flexibility in such polymers. If we assume that a single correlation time τ_c describes the motion of the backbone chain and that this motion is isotropic, then the following equations apply (13):

$$\frac{1}{NT_1} = \frac{1}{10} \frac{\gamma_H^2 \gamma_C^2 \hbar^2}{r_{C-H}} \cdot \chi(\tau_c)$$

For the nuclear Overhauser enhancement:

$$\eta = \frac{\gamma_H}{\gamma_C} \frac{6\tau_c / \left[1 + (\omega_H + \omega_C)^2 \tau_c^2 \right]^{-\tau_c} \left[1 + (\omega_H - \omega_C)^2 \tau_c^2 \right]}{\chi(\tau_c)}$$

where

$$\chi(\tau_c) = \frac{\tau_c}{1 + (\omega_H - \omega_C)^2 \tau_c^2} + \frac{3\tau_c}{1 + \omega_C^2 \tau_c^2} + \frac{6\tau_c}{1 + (\omega_H + \omega_C)^2 \tau_c^2}$$

The symbols have their usual meanings (13). From measured values of NT_1 and η on a poly(butene-1 sulfone) of degree of polymerization 700 the values of τ_c (in nanosec.) shown in Table III are obtained. The discrepancy between the values of τ_c from NT_1 and from η , particularly marked for the side-chain motions, indicates the inadequacy of the single- τ_c model. Nevertheless it is evident that the backbone motions are relatively rapid. (Comparison to polybutene-1 (14) shows that SO_2 groups retard the motion of the copolymer chains by a factor of about 50.) The question now becomes: why are these rapid motions NMR-active but dielectrically inactive? One possible type of motion which would account for this is shown in Fig. 9. Five backbone bonds and six main-chain atoms are involved, i.e. the sequence C-S-C-C-S-C, with concerted segmental transitions about two C-S bond, allowing interconversion of the $t t t$, $g^+ t g^-$, and $g^- t g^+$ conformers. The backbone C-C bond remains trans; the sulfone dipoles remain antiparallel and cancel each other.

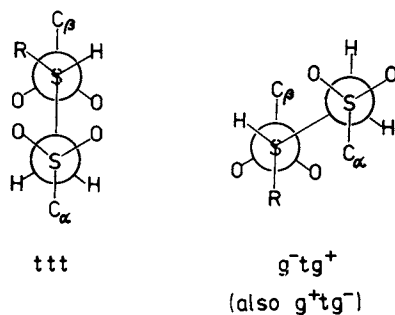


Figure 9. Proposed allowed equilibrium conformational states for poly (α -olefin sulfones) in solution. Note that the sulfone dipoles cancel and that during the transitions $ttt \rightleftharpoons g^-tg^+ \rightleftharpoons g^+tg^-$ there is no net reorientation of these dipoles (dielectrically inactive motions), but there is a reorientation of backbone C-H vectors (C-13 NMR active motions).

Table 3

Correlation Times for Backbone and Side-Chain Motions
in Poly(but-1-ene sulfone) of P_n = 700 as a 25% w/v
Solution in Chloroform-d, Deduced from the Simple
Isotropic Single- τ_c Motional Model

Temp, K	τ_c , ns					
	Backbone from		Ethyl Branch			
	NT ₁	η	CH ₂ from		CH ₃ from	
			NT ₁	η	NT ₁	η
328	22	2.6	0.29	1.9	0.027	1.3
313	23	3.1	0.33	2.2	0.030	1.3
298	27	3.4	0.38	2.7	0.039	1.3

Somewhat surprisingly, a 2:1 styrene:SO₂ copolymer is slightly more flexible than polystyrene itself (9). The styrene-styrene (SMMS) units evidently constitute flexible joints which permit an overall freedom of motion despite local constraints at SMS units. The greater length of the C-S bond compared to that of the C-C bond (0.180 vs. 0.154 nm) may also contribute.

LITERATURE CITED

- Solonina, W., Zh. russk. fiz.-khim. Obshch., (1898), 30, 826.
- Solonina, W., Chem. Zent., (1899), 1, 249.
- Dainton, F. S. and Ivin, K. J., Nature, (1948), 162, 705.
- Barb, W. G., Proc. Roy. Soc., London, Ser. A, (1952), 212, 177.
- Matsuda, M., Iino, M., and Tokura, N., Makromol. Chem., (1962), 52, 98.
- Tokura, N. and Matsuda, M., Kogyo Kagaku Zasshi, (1961), 64, 50.
- Cais, R. E., O'Donnell, J. H., and Bovey, F. A., Macromolecules, (1977), 10, 254.
- Coleman, M. M., Tabb, D. L. and Brame, E. G., Rubber Chem. Tech., (1977), 50, 49.
- Cais, R. E. and Bovey, F. A., Macromolecules, (1977), 10, 757.
- Stockmayer, W. H., Jones, A. A., and Treadwell, T. L., Macromolecules, (1977), 10, 762.
- Fawcett, A. H., Heatley, F., Ivin, K. J., Stewart, C. D. and Watt, P., Macromolecules, (1977), 10, 765.
- Bates, T. W., Ivin, K. J. and Williams, G., Trans. Faraday Soc., (1967), 63, 1964.
- See, for example, Shaefer, J., "Structural Studies of Macromolecules by Spectroscopic Methods", K. J. Ivin, Ed., Wiley-Interscience, New York, NY, 1976, pp. 20.-226.
- Schilling, F. C., Cais, R. E., and Bovey, F. A., Macromolecules, (1978), 11, 325.

Discussion

D.J. Worsfold, NRC, Ont. Problems often arise in carbon-13 NMR work with the integration of the peaks. In the analysis of copolymer systems it would be useful to have very good peak integration. Can you in fact succeed in this?

F.A. Bovey. That is a question I wish Dr. Cais were here to answer. The assumption generally in such problems is that the Overhauser enhancement is at least the same for all peaks. I am not certain this is a justifiable assumption. In fact, if you study Table II carefully you will see Overhauser enhancements which are in fact not necessarily the same. That this can lead to problems is quite clear. If work is done at a sufficiently high temperature I think the assumption of equal Overhauser enhancements is probably reasonably valid. In this case the peaks can be intercompared. We usually do this by old fashioned planimeter methods rather than by using the integrator on the instrument. It seems to be more reliable. This matter of integration is a definite problem which is becoming increasingly recognized.

A.A. Jones, Clark Univ., Mass. I would tend to agree with the dynamic picture you presented. The idea that the high frequency motion in the sulfone hexene type polymers doesn't cause a net relaxation of dipoles but moves the magnetic dipole-dipole interaction around to cause nuclear relaxation is, I think, the crux of the matter.

G. Babbitt, Allied Chemical Corp., N.J. You showed quite large shielding and deshielding effects of SO₂ units in the styrene-SO₂ copolymer chains. They are very dramatic. Have you made any efforts to explain this?

F.A. Bovey. No, I think the first thing we would like to understand is the carbon carbon gamma or gauche effect from which we might proceed to understand hetero atom gauche effects. I do not think anyone understands either one of them right now -- at least I don't. Most of the theoretical efforts which I have seen are not very convincing. The original explanation of Cheney and Grant (B.V. Cheney and D.M. Grant, J. Am. Chem. Soc., 89, 5319 (1967) required the carbons to have protons on them in order for this effect to be evident. This is certainly not true. There are gamma effects for all kinds of atoms other than carbon which do not have attached protons. These systems exhibit strong effects. Even with the protons attached I never quite understood what their explanation was. In fact it cannot be limited to that explanation because it can be seen in many situations. I think every atom can have a gamma shielding effect. Even fluorines have a gamma effect, as we

found in interpreting the C-13 spectrum of polyvinylidene fluoride. It will take a very potent theoretical person to work out the explanation of these effects and this has not happened yet to my knowledge. Perhaps someone else would care to comment on this question.

C.J. Carman, B.F. Goodrich, Ohio. I want to substantiate a point with regard to the hetero atom effect. We have tried to extract the gamma effect of chlorine atoms in PVC and chlorinated polymers and we have found that it's not quite as large as with oxygen and not necessarily predictable as to its direction. I think that sometime in the future the hetero atom effects on gamma may be extractable and of interest. They are not very predictable as far as transferring from one system to another. They can be very large as with oxygen.

F.A. Bovey. Alan Tonelli in our laboratory has been working on the prediction of carbon chemical shifts in polymer chains and he finds what you found for the polyvinyl chloride. I think you first pointed out that the gamma effect can be used to explain such chemical shifts. Polyvinyl chloride can be explained if enough variable parameters are used. PVC is not the simplest case. Polypropylene on the other hand can be explained with a single constant but PVC apparently cannot. Dr. Tonelli hasn't published any of this yet, he is still working it out.

T.K. Wu, E.I. duPont de Nemours, Delaware. Were your spectra obtained at a field higher than 22.6 MHz?

F.A. Bovey. I'm glad you asked that question. I should have pointed out very clearly that all of the spectra I showed for the chloroprene system were done at 90 MHz while those of the styrene system were obtained at 25 MHz.

T.K. Wu, E.I. duPont de Nemours, Delaware. A couple of years ago Dr. Obiano, Dr. Haine and I studied a simpler alternating copolymer, that is, theylene CO, which we called a semi-alternating copolymer because the ethylene content was variable. From the C-13 and proton NMR spectra we felt we could treat this copolymer as a hypothetical copolymer of ethylene and a fictitious comonomer ethylene-CO. Using that picture we found the comonomer distribution was very close to a Bernoullian distribution.

RECEIVED March 13, 1979.

Polysaccharide Branching and Carbon-13 NMR

F. R. SEYMOUR

Baylor College of Medicine, Texas Medical Center, Houston, TX 77030

A fortunate relationship exists between current C-13 n.m.r. spectroscopy and polysaccharide structure. The structures of many polysaccharides are complex, providing a variety of resonances resulting from different structural features. At 25 MHz, a typical currently available field-strength, many resonances are well resolved and this encourages attempts to assign these resonances to specific carbon atom positions. Unfortunately, C-13 n.m.r. is not a fundamental structural technique -- that is, at our present state of knowledge it is not possible to directly interpret a saccharide C-13 n.m.r. spectrum without recourse to comparison spectra of known compounds. However, the above disadvantage is redressed in that C-13 n.m.r. spectra allow an intimate view of molecular structure, especially with regard to the nature of the anomeric linkages for specific types of residues. This very precise anomeric linkage data complements our basic structural analysis technique, permethylation g.l.c.-m.s., a technique that provides no information about the anomeric configuration of the various linkages. The general structural analytical approach which has been developed for polysaccharide structural analysis has previously been discussed in this Symposium Series (1). In general, data has been taken from fragmentation g.l.c.-m.s., permethylation fragmentation g.l.c.-m.s. (2-5), high-pressure liquid-chromatography of acetolysis products (3), C-13 n.m.r. (6,7), and P-31 n.m.r. (8), with these data then being inter-related. The following discussion will primarily deal with C-13 n.m.r., but the additional data is important. For example, permethylation fragmentation g.l.c.-m.s. analysis, which has been performed for all polysaccharides discussed, provides definitive evidence for the type and degree of saccharide branching. All of the following C-13 n.m.r. spectra-to-structure correlations are in agreement with this additional data.

As the basic spectral analysis approach consists of comparing the C-13 n.m.r. spectra of a large number of polysaccharides of known, or partially known, structure, the availability of a

0-8412-0505-1/79/47-103-027\$06.25/0
© 1979 American Chemical Society

comprehensive polymer collection is important. Furthermore, in comparison to other structural techniques (e.g. g.l.c.--m.s. analysis) Fourier transform C-13 n.m.r. is relatively insensitive, and requires large amounts of material (~ 200 mg) for complete analysis at high signal-to-noise ratio in reasonable spectrometer acquisition times. When considering the total spectrometer time required for obtaining a large reference collection of spectra, the available sample size becomes critical. Extracellular microbial polysaccharides provide an excellent source of reference materials, available in relatively large amounts, and are known to contain a variety of structural features. Many polysaccharides consist of hexose residues linked in the pyranoside ring form. In addition, many members of this polymer class are exclusively composed of D-glucopyranosyl residues (the D-glucans). Dextran is an extensively studied sub-group of D-glucans, and are polymers which contain a large percentage of (1 \rightarrow 6)-linked α -D-glucopyranosyl residues. Our original C-13 n.m.r. spectral studies were carried out on dextrans for four reasons: a) the compounds were available in suitable amounts, b) an abundant literature on the characterization of these compounds existed (9, 10), c) we had just completed accurate g.l.c.--m.s. structural analysis for many of these compounds (which indicated the different polymers had varied and well-defined structures), and d) it was hoped that the relatively small structural differences in various dextran series would allow a precise correlation between changes in structure to changes in spectra. Once the general spectral relationships were established for the various dextrans, it would then be possible to apply these relationships to polysaccharides of more diverse character.

It should be emphasized that although many structural features of the various polysaccharides are known, in general these polysaccharides are by no means completely characterized. However, with extensive C-13 n.m.r. chemical shift and intensity data, it is possible to employ these reference spectra to further characterize the compounds under consideration.

An important contribution to the analysis of 25 MHz spectra was the realization that elevated sample temperatures greatly aid spectral acquisition and interpretation (6). On raising the sample temperature the C-13 resonances narrow, providing both enhanced resolution and decreasing (by about 1/4) the number of acquisitions necessary to obtain a comparable signal-to-noise ratio for a corresponding ambient spectrum. It is for this reason that we normally record and reference our polysaccharide spectra at the highest temperature (90 $^{\circ}$) readily compatible with an aqueous solution. However, such high-temperature spectra must be interpreted with care due to the chemical shift dependence on temperature ($\Delta\delta/\Delta T$) for each resonance involved. In general, all 90 $^{\circ}$ resonances are displaced down-field (relative to ambient, 34 $^{\circ}$, resonances) by ~ 1 p.p.m. -- when referenced against the deuterium lock. The $\Delta\delta/\Delta T$ for different resonances vary as much as two

fold. Within spectrometer error $\Delta\delta/\Delta T$ is constant for dextrans through out the 30--90° range. Though the individual resonances narrow at elevated temperatures, the general "profile" of the spectrum is constant for most polysaccharides that have been studied (8, 12) -- the few exceptions will be discussed below.

Our original approach to polysaccharide C-13 n.m.r. spectral analysis consisted of making a minimum number of hypotheses about expected structure-to-spectra relationships (8). By then comparing spectra to known structure for a series of D-glucans, we attempted to establish the validity of these hypotheses and to establish how diverse a structural difference could be accommodated. The hypotheses were as follows. Firstly, that each polymer could be considered as an assembly of independent saccharide monomers. Secondly, that these hypothetical saccharide monomers would be O-alkylated (O-methylated) in the same positions as the actual saccharide linked residues (it had previously been established that O-methylation of any α -D-glucopyranosyl carbon atom position resulted in a down-field displacement of ~ 10 p.p.m. for the associated resonance). Thirdly, that each differently substituted residue would have a completely different set of chemical shift values for each carbon atom position (different from the unsubstituted saccharide) but that only the carbon atom positions involved in inter-saccharide linkages would have $\Delta\delta$ greater than 1 p.p.m. And, fourthly, that the hypothetical O-alkylated residues would contribute resonances to the total spectrum proportional to their mole ratio in the polymers.

At this point it is convenient to reiterate the major features and relationships normally encountered for polysaccharide structures. Each saccharide residue is normally linked through the reducing (hemi-acetyl) functional group (the anomeric position) to the hydroxy position of another residue. Each residue contains a number of hydroxy positions, but only one anomeric position. When a residue is linked only through the anomeric position, then that residue is a non-reducing terminal residue. When a residue is linked through both a hydroxy position and an anomeric position, then a linear, chain extending residue results. In like manner, should a residue be linked through both the anomeric position and two hydroxy positions, then a branch-point residue results. In general, any polysaccharide contains a single reducing end group and $b+1$ terminal residues, when b represents the number of branch-point residues. Therefore, large polysaccharides effectively contain no reducing end groups, and an equal number of branch-point residues to terminal non-reducing residues.

Dextrans

As previously stated, our C-13 n.m.r. spectral analysis was first applied to dextrans. In the spectral analysis hypotheses described above, it is an inherent assumption that each residue

will be considered independently, without regard to the nature of the adjacent linked residues. This procedure was employed for simplicity of spectral analysis, and on the assumption that such adjacent saccharide effects would be undetectably small. However, we knew that such adjacent saccharide effects would ultimately need to be considered. Our own P-31 n.m.r. spectra (8) had shown such effects-at-a-distance, and a previously published pullulan spectrum had shown resonance splitting which had convincingly been explained as due to changes in (1→4)-linked α -D-glucopyranosyl residues which come before and after a (1→6)-linked α -D-glucopyranosyl residue in the linear chain (12). However, a large amount of data accumulated for the dextrans over the past three decades indicate that these compounds contain a backbone composed exclusively of (1→6)-linked α -D-glucopyranosyl residues, with branching occasionally occurring from this backbone chain via the 2, 3, or 4-hydroxy positions (9). Such branching effects need not introduce as profound a conformational change into the individual residues as does the introduction of a different linkage type into the polysaccharide backbone. In practical terms the hypothesis of isolated residues has served very well for the analysis of the dextran spectra. Unsubstituted α -D-glucopyranoside (and apparently any hexapyranoside) exhibits six resonances which can be observed in three regions: the 90--95-p.p.m. region, containing the anomeric (C-1) carbon resonance; the \sim 70--75-p.p.m. region, containing the C-2, C-3, C-4, and C-5 resonances; and the 60--70-p.p.m. region containing the C-6 resonance. On O-substitution the resonance of the anomeric carbon is displaced down-field by \sim 10 p.p.m. to the 97--102-p.p.m. region. Similar O-substitution of carbon atoms with resonances normally in the 70--75-p.p.m. region results in displacements into the 80--85-p.p.m. region. The free C-6 resonance, at \sim 68 p.p.m., on O-substitution is displaced to \sim 62 p.p.m., so that both free and linked carbon resonances are in the 60--70-p.p.m. region. Therefore, as the percentage of free (reducing) D-glucose residues contained in the dextrans is insignificant, only four spectral regions are of interest: the 60--70-p.p.m. region, containing both free and linked C-6 resonances, the 70--75-p.p.m. region containing the free C-2, C-3, and C-4 resonances and the pyranoside ring C-5 resonance; the 75--85-p.p.m. region containing linked C-2, C-3, and C-4 carbon resonances, and the 95--103-p.p.m. region containing linked anomeric (C-1) resonances (see Figure 1, upper spectrum).

The two spectral regions most amenable to analysis are the 95--103-p.p.m. region (C-1 resonances) and the 75--85-p.p.m. region (the linked C-2, C-3, and C-4 resonance position). This phenomenon could easily be expected, as these two regions contain the resonances of the carbon atoms which are directly participating in the inter-saccharide linkages. The 75--85-p.p.m. region presents a rather straight-forward situation, as each branch-type results in a specific branching residue (e.g. 2-O-branching from

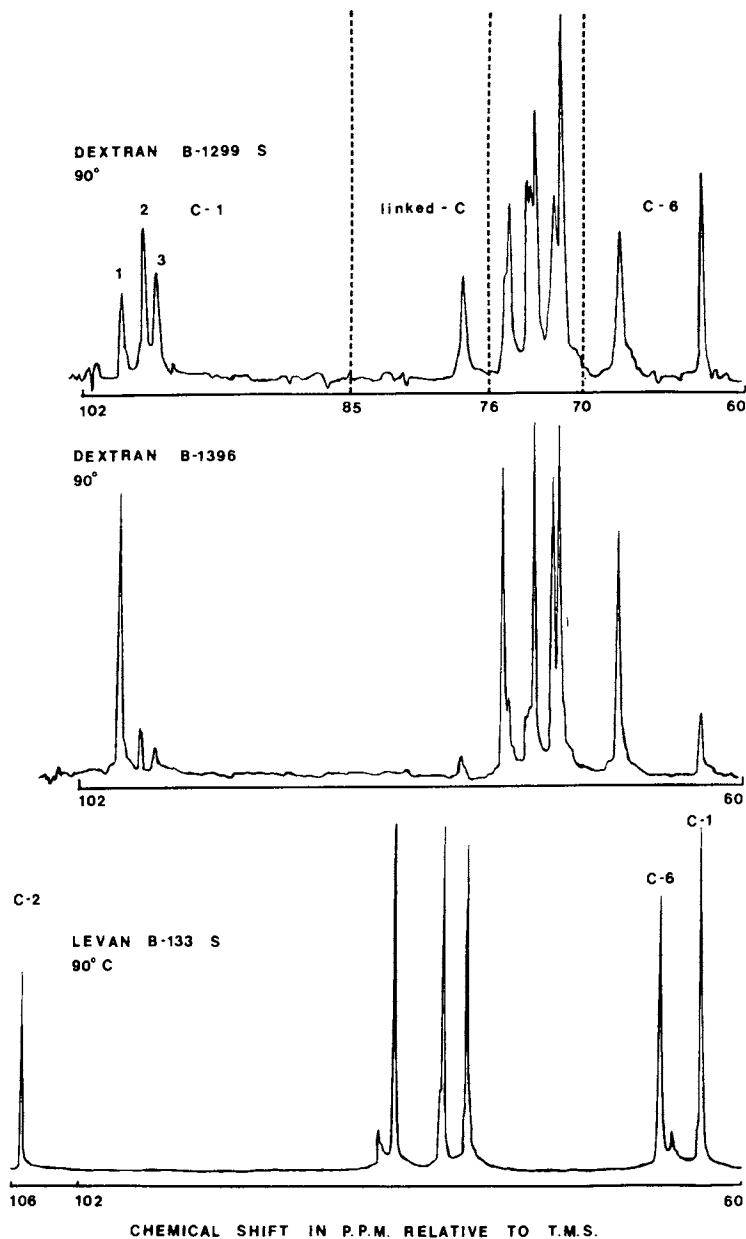
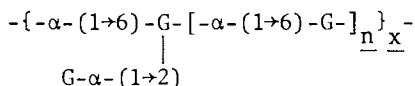


Figure 1. The 90° ^{13}C NMR spectra for: (top) dextran B-1299 Fraction S; (middle) dextran B-1396; and (bottom) levan B-133 Fraction S.

a backbone composed of 6-O-substituted α -D-glucopyranosyl residues yields a 2,6-di-O-substituted residue) and the linked carbon resonance is now displaced ~ 10 p.p.m. down-field from the 70--75-p.p.m. region. At 90° α -(1 \rightarrow 2)-, α -(1 \rightarrow 3)-, and α -(1 \rightarrow 4)-linkages result in diagnostic resonances, respectively, at approximately 78, 80, and 82 p.p.m. The term "approximately" is used advisedly because some differences in chemical shifts have been noted for the same type of inter-sugar linkages. However, if the spectra are analyzed by considering the type of residue present, rather than the linkage type present, then the diagnostic resonances in the 75--85-p.p.m. region are consistent. Two examples will be presented to demonstrate this residue type specificity effect for the diagnostic resonances.

Several series of graded (in terms of degree of branching) dextrans have been studied. The example presented here deals with dextrans branching through 2,6-di-O-substituted α -D-glucopyranosyl residues (12). The average repeating unit for these dextrans is shown in Structure 1, where \bar{G} represents the D-glucopyranosyl residue and \bar{n} represents the average number of 6-O-substituted α -D-glucopyranosyl residues between branch points.



Structure 1

Assuming that all α -(1 \rightarrow 6)-linkages are in the dextran backbone, our permethylation g.l.c.-m.s. data (which indicate that only the residues shown above are present in this series of dextrans) are consistent with a polymeric structure that is comb-like with side branches a single residue long.

Figure 2 shows a specific example of the effect on a C-13 n.m.r. spectrum when the sample temperature is changed. With increasing temperature the resonances are displaced down-field and become more narrow. The upper plot of Figure 2 shows the linear $\Delta\delta/\Delta T$ for the resonances designated in the spectra. The dextran anomeric ΔT effects are even greater than the selected example shown in Figure 2.

The dextran series B-1299 fraction S, B-1402, B-1424, B-1422, and B-1396 have been shown to have structure 1 with \bar{n} respectively equal to 1, 2, 3, 6, and 8. These dextran (and the following mannan and levan) designation numbers refer to the Northern Regional Research Center (NRRL) designation for the producing microorganism strain. A single sharp resonance in the 75--85-p.p.m. region was observed for each of the dextrans (see Figure 1, upper and middle spectra), respectively at 77.79, 77.80, 77.79, 77.74, and 77.76 p.p.m. These chemical shift differences essentially represent the limits of our spectrometer error.

Comparison of two series of dextran data shows the importance of considering the spectra in terms of specific residue type. One

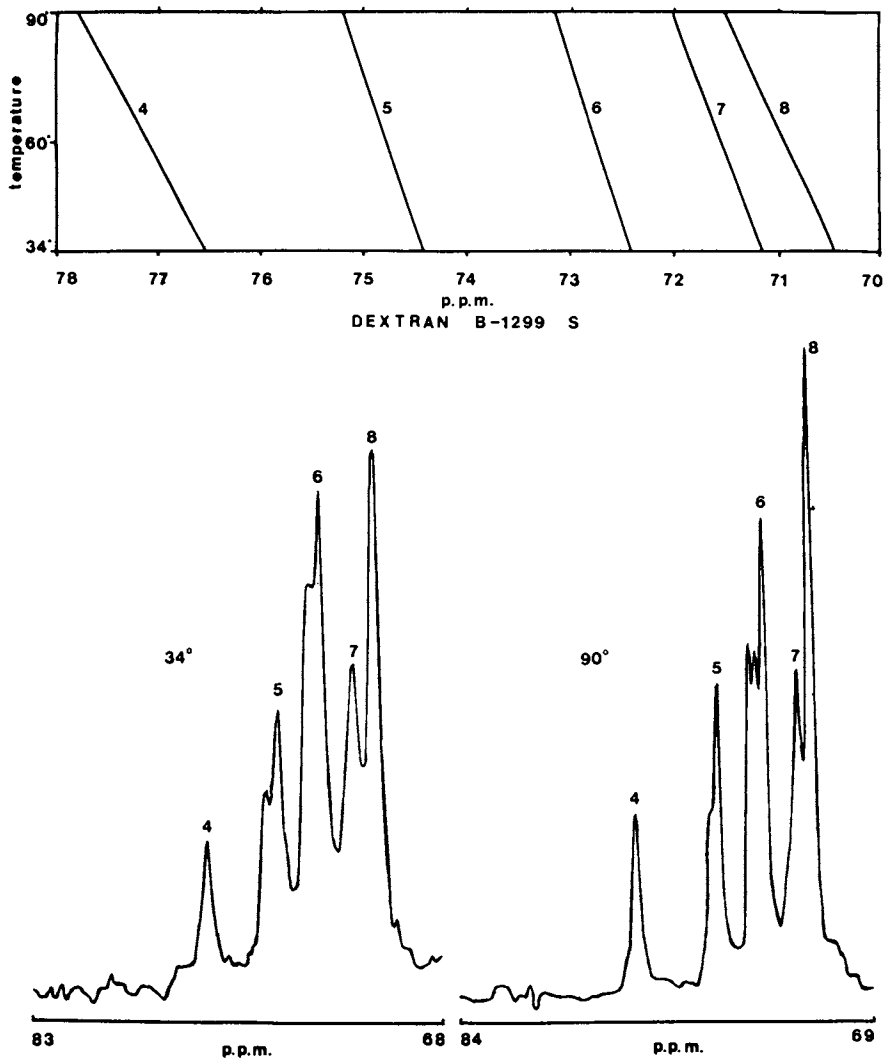
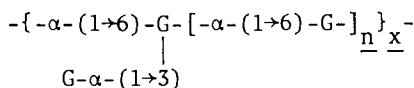


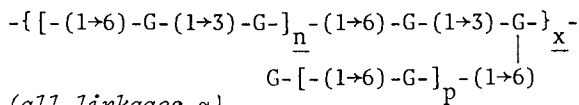
Figure 2. $C-13$ NMR spectra of the 69–84 ppm region for dextran B-1299 Fraction S (total 90° spectrum in Figure 1) at 34° and 90° . Top plot shows the $\Delta\delta/\Delta T$ for five resonances identified in the bottom spectra.

dextran series is essentially the same as the above 2-O-substituted series, except the branching is through the 3,6-di-O-glucopyranosyl residue (see Structure 2). Representative of this



Structure 2

series are dextrans B-742 fraction S, B-1142, B-1191, and B-1351 fraction S, dextrans which have respective values of \underline{n} to be 0, 1, 2, and ~ 10 . The C-13 spectrum of each of these dextrans contains a single sharp resonance in the 75--85-p.p.m. region with diagnostic chemical shifts respectively of 82.81, 82.89, 82.84, and 82.86; again the limits of our spectrometer accuracy. However, permethylation fragmentation g.l.c.-m.s. analysis has indicated that several of the (1 \rightarrow 3)-linkage containing "dextrans" differ dramatically from other dextrans. In fact, though these polysaccharides have been traditionally classed with the dextrans due to their origin, they quite possibly fail to fulfill the definition of "dextran" by not having backbones exclusively composed of 6-O-substituted α -D-glucopyranosyl residues. This class of polysaccharides is represented by the S fractions of dextrans B-1355, B-1492, and B-1501, and a general structure is proposed for this class (structure 3) which is based on C-13 n.m.r.



Structure 3 (all linkages α)

and g.l.c.-m.s. data (13).

The feature of immediate interest in the above general structure is that relatively large (and for dextran B-1355 fraction S, very large) percentages of 3-O-substituted α -D-glucopyranosyl residues are present with relatively small proportions of the 3,6-di-O-substituted α -D-glucopyranosyl residues. Careful examination of the 75--85-p.p.m. region for the above polymers shows that the major resonance is now respectively at 83.31, 83.31, and 83.32 p.p.m. These values are again indistinguishable, and are clearly displaced from the diagnostic chemical shifts observed for the dextran B-742 fraction S series, which exclusively contain (1 \rightarrow 3)-linkages associated with 3,6-di-O-substituted α -D-glucopyranosyl residues. In addition, dextran B-1501 fraction S contains a relatively modest excess of 3-O-substituted α -D-glucopyranosyl residues when compared with the 3,6-di-O-substituted α -D-glucopyranosyl residues, and careful examination of the 75--85-p.p.m. region of this dextran displays a major resonance at 83.32 p.p.m. (indicative of the former residue and designated \underline{d} in Figure 3, upper spectrum)

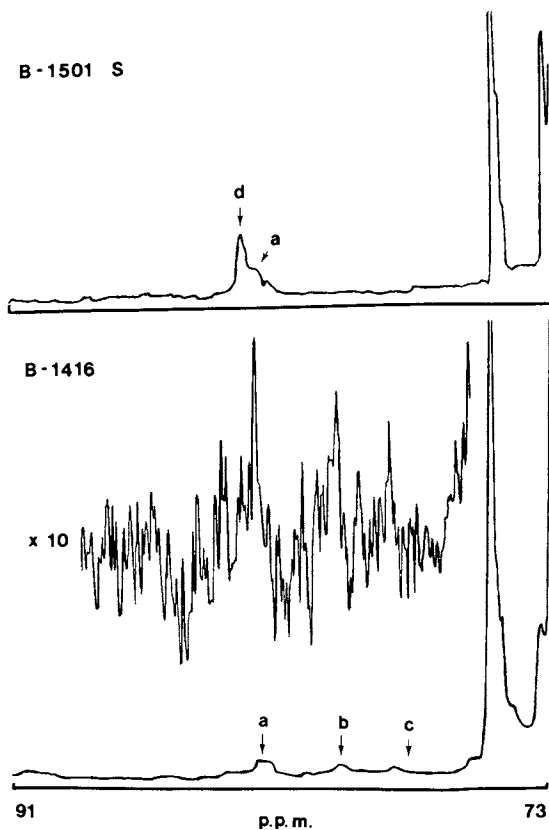


Figure 3. The 73–91 ppm 90° $C-13$ NMR region for the dextran B-1501 Fraction S and the dextran B-1416 spectra. The letters identify: Resonance a (from the 3,6-di- O -substituted α -D-glucopyranosyl residue); Resonance b (from the 4,6-di- O -substituted α -D-glucopyranosyl residue); Position c (for the 2,6-di- O - α -D-glucopyranosyl residue); and Resonance d (from the 3- O -substituted α -D-glucopyranosyl residue). A tenfold scale expansion plot of dextran B-1416 is also shown.

and a shoulder at 82.9 p.p.m. (indicative of the latter residue and designated a in Figure 3, upper spectrum). In conjunction with similar studies for (1→4)-linkage containing dextrans, it is concluded that resonances have been observed which are specific for the 3-O-substituted, 4-O-substituted, 6-O-substituted, 2,6-di-O-substituted, 3,6-di-O-substituted, and 4,6-di-O-substituted α -D-glucopyranosyl residues, and these diagnostic resonances are specific for each residue type.

The separation of these 75--85-p.p.m. diagnostic branching resonances is indicated in the lower part of Figure 3. No completely linear microbial dextran is known, but dextran B-1416 is a dextran example with low degree of branching. It is one of the few dextrans which have been identified as containing a mixture of branch-type residues. Both the 3,6-di-O-substituted and the 4,6-di-O-substituted residue resonances are weakly present. A third, and unexplained, very weak resonance (78.3 p.p.m.) is near, but clearly displaced from, the position c (77.8 p.p.m.) which is diagnostic for the 2,6-di-O-substituted α -D-glucopyranosyl residue.

In general, the spectra of any dextran is a composite consisting of the six major resonances associated with linear dextran (the 6-O-substituted α -D-glucopyranosyl residue) and minor resonances associated with the branching and corresponding terminal residues. Due to the corresponding terminal α -D-glucopyranosyl residue associated with each branch-point residue, it could be expected that each branching type could contribute twelve (2x6) minor resonances to the dextran spectra, and that the "minor" resonances would steadily become more intense (compared to the linear dextran resonances) as the polymer becomes more highly branched. We have not observed all of these expected additional resonances, and assume that in each case certain of these expected resonances are not resolved from the closely packed 70--75-p.p.m. region.

The anomeric, 95--110-p.p.m. region presents a second complementary spectral region for structural analysis. In contrast to the 75--85-p.p.m. region, the anomeric region presents two new resonances for each branching type -- one resonance from the branch-point residue and a second from the corresponding terminal residue. Based on a number of polymer analyses, the C-1 resonances are the most sensitive to structural change, and at this point there is failure for the hypothesis that neighboring residues have no effect on a given residue type's resonances (this is the only failure of this hypothesis which has been noted for the dextrans). The anomeric spectral region of the dextran series branching through the 2,6-di-O-substituted α -D-glucopyranosyl residue (see Structure 1 and Figure 1) displays two branching resonances (97.37 and 98.22 p.p.m.), in addition to the linear dextran anomeric resonance. One new additional resonance belongs to the C-1 of the branch-point residue and the other additional resonance corresponds to the C-1 terminal residue.

If the hypothesis stating that the resonances of each residue type were independent of adjacent residue structures, then the C-1 resonance of the terminal residue would be easy to identify by comparing the additional branching anomeric resonances of 2-branching, 3-branching, and 4-branching, thereby identifying the consistent minor resonance. This is not the case, as a comparison of the series of dextrans of different branching type demonstrates. The spectra of each branching type (see Figure 4, top spectra) display two additional anomeric branching resonances, but for each type of branching both minor resonances differ from the minor resonances of the other branching types of dextran. The actual assignment of these anomeric resonances will be discussed below. In addition, a small but distinct chemical shift is noted for the anomeric resonance of the 3,6-di-O-substituted residue (100.88 p.p.m.) compared to the 3-O-substituted (101.07 p.p.m.) residue.

Most importantly, in comparing the C-13 n.m.r. spectra of the dextran series branching through the 2,6-di-O-substituted α -D-glucopyranosyl residue to the permethylation g.l.c.-m.s. data, it was concluded that the intensity of all resonances present is, to a first approximation, proportional to the number of atoms of that carbon position present; and also, in general the peak heights of the minor resonances (compared to the major resonances of linear dextran) are proportional to the degree of branching. This effect was specifically confirmed by comparing the ratio of peak heights of a branching anomeric resonance (99.2 p.p.m.) and the linear dextran resonance (98.7 p.p.m.) to permethylation data (12). However, this effect can be shown for any well resolved minor resonance of any of the different branching type dextran series. The establishment of such relationships is of importance as C-13 resonance intensities do not necessarily bear any direct relationship to the percentage of specific position of carbon atoms present.

The general anomeric resonance C-13 n.m.r. region of D-glucans is of considerable interest as it has been concluded, on the basis of monomers, that β -configuration anomeric carbons have chemical shifts down-field from α -configuration anomeric carbons. All evidence in the literature at present confirms this general observation, but until a wide variety of monomers and polymer residues have been examined, it will be difficult to establish exactly what the range of the α - and β -anomeric resonance regions are, and whether these two regions can overlap. Currently it is suggested that these regions (corrected to 90°) would be about 95--103-p.p.m. for the α -configuration, and about 103--108-p.p.m. for the β -configuration. Due to large positive specific rotations, the dextrans were known to be predominantly, if not exclusively, α -linked. In none of our dextran spectra have we observed any anomeric resonance down-field from 101.6 p.p.m., nor up-field from 97.2 p.p.m. This suggests that the dextrans contain no (observable) minor β -linked residues, and that the general spectral

region for α -D-glucopyranosyl residues lies in the 97--102-p.p.m. region.

Relatively little spectral analysis has been done on the 60--70-p.p.m. or the 70--75-p.p.m. regions of the dextran spectra. The 70--75-p.p.m. region is closely packed with a number of resonances, and therefore the identification of specifically displaced carbon position resonances is difficult at our current spectral resolution. The 60--70-p.p.m. region provides a good indication of the degree of branching, by comparing the free C-6 resonance intensity (~ 63 p.p.m.) to the linked C-6 resonance intensity (~ 68 p.p.m.). However, little additional information is forthcoming as the C-6 position of the α -D-glucopyranosyl residue is relatively insensitive to the type of branching. In fact, the general insensitivity at the C-6 position can be observed in the relatively small $\Delta\delta$ of substitution (~ 5 p.p.m.).

This C-13 n.m.r. analytical technique has been referenced against 27 dextrans which had been studied by periodate-oxidation and permethylation fragmentation g.l.c.-m.s. techniques (14). These three independent structural methods give data for all dextrans which are in accord with each other. The 27 dextrans studied represent 25 structures which differ in type or degree of branching. Such extensive cross-referencing of data give us increased confidence with regard to spectra-to-structure relationships.

The nature and relationships of the resonances in dextran spectra have been dealt with at length due to the wide variety of branching types available, and to the relatively subtle changes in structure available as one progresses through a specific dextran branching type series. The C-13 n.m.r. spectra of the following polysaccharides can now be dealt with in a more succinct manner by describing how they differ from the above observations.

Synthetically branched amylose

This series of compounds, produced by adding D-glucopyranosyl monomers to amylose [the linear polymer composed of (1 \rightarrow 4)-linked α -D-glucopyranosyl residues], was of interest as a number of different highly-branched comb-like amylose products had been produced by employing different types of condensation reaction conditions. In general, such D-glucopyranosyl additions result in the formation of β -linkages, but certain reaction conditions (specifically the Helferich reaction conditions) were anticipated to yield the addition of α -linked D-glucopyranosyl terminal residues to the linear amylose chain. Various techniques, including permethylation g.l.c. analysis, were able to demonstrate the degree of monomer incorporation and the position of attachment (at the C-6 position of the amylose residues). However, establishment of the anomeric configuration proved much more difficult, as most methods of anomeric linkage determination depend on bulk properties of the polymer (e.g. specific rotation) and also on the

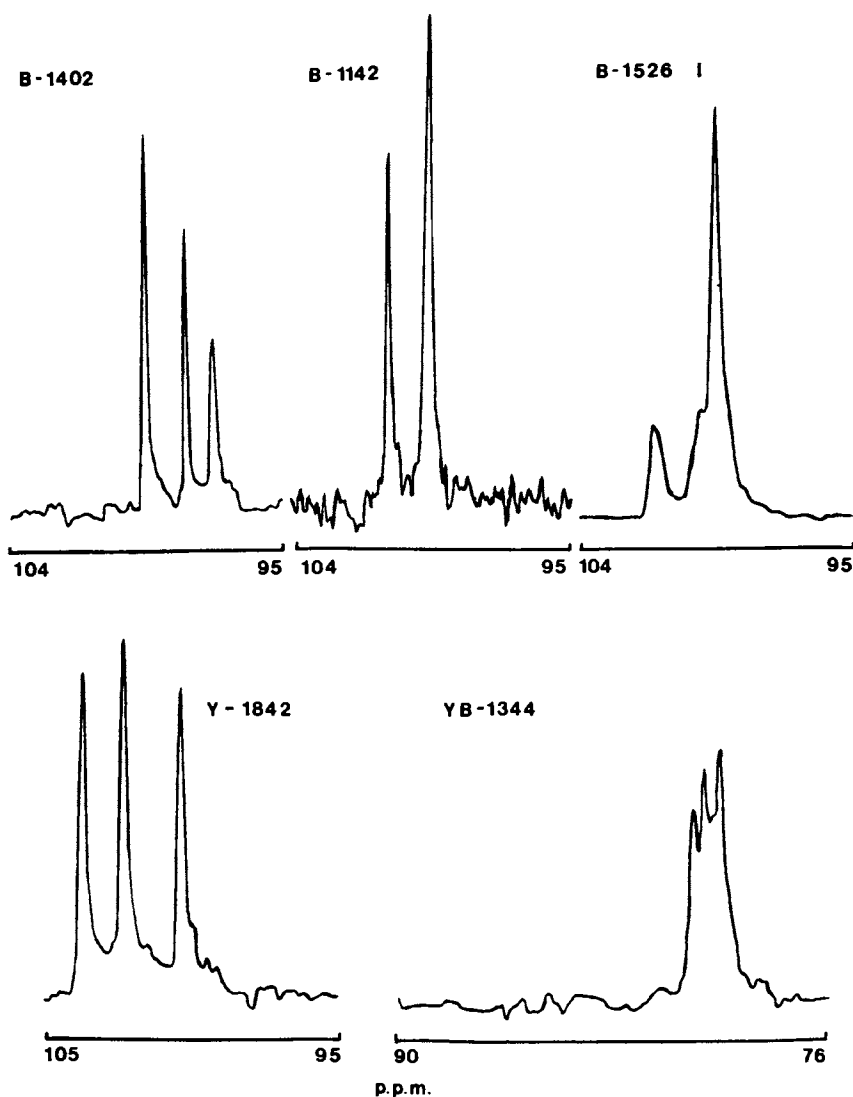
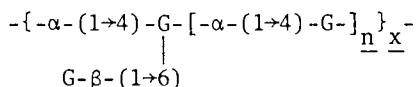


Figure 4. (Upper) Anomeric region of the C-13 NMR spectra at 90° for: dextran B-1402 (2,6-di-O-substituted branching); dextran B-1142 (3,6-di-O-substituted branching); dextran B-1142 (3,6-di-O-substituted branching); and dextran B-1526 Fraction I (4,6-di-O-substituted branching). For each dextran the most intense resonance (99.6 ppm) represents the 6-O-substituted α -D-glucopyranosyl residue. (Lower) Anomeric region of the 90° C-13 NMR spectra for mannan Y-1842 and the 76-90 ppm region for mannan YB-1344

fact that the amylose backbone, now incorporated into the new polymer, contributed large amounts of α -linkages (see Structure 4).



Structure 4

To answer this linkage type question we first recorded the spectra of glycogen (a polymer similar to amylose, but more soluble), of both methyl α -D-glucopyranoside and methyl β -D-glucopyranoside, and of the unknown synthetically branched amylose products (see Figure 5). All examples of synthetically branched amylose proved to have a strong branching anomeric resonance at 104.4 p.p.m. (15) (well into the β -anomeric linkage spectral region) and no anomeric resonance at 100.1 or 101.6 p.p.m. (resonances observed for 4,6-di-O-substituted α -D-glucopyranosyl branching in dextrans). On this basis it was concluded that all condensation conditions yielded essentially exclusively β -D-linked branching. Furthermore, the total spectrum of the synthetically branched amyloses was essentially a composite of the amylose resonances plus the methyl β -D-glucopyranoside resonances. Two points were of interest in this study. Firstly, the glycogen diagnostic resonances in the 75--85-p.p.m. region (that of the 4-O-substituted α -D-glucopyranosyl residue) was at 79.4 p.p.m. which was somewhat different from the diagnostic branching resonance of 4-O-branched dextran at 80.4 p.p.m. (the 4,6-di-O-substituted α -D-glucopyranosyl residue). As was previously shown for α -(1 \rightarrow 3)-linkages for dextran, this again indicates that the diagnostic resonances are dependent on the residue type present, and not on the linkage type. Secondly, resonances similar to both the above (1 \rightarrow 4)-linked glycogen and dextran resonances are observed in the synthetically branched amylose. The presence of these diagnostic resonances in the synthetically branched amylose is not surprising as this compound contains both 4-O-substituted and 4,6-di-O-substituted α -D-glucopyranosyl residues. However, the latter residue is now linked to a β -D-glucopyranosyl residue at the C-6 position. The branched amylose spectrum (Figure 5) clearly shows these effects -- resonance a (80.3 p.p.m.) is essentially identical to the diagnostic resonance for dextrans branching through the 4,6-di-O-substituted α -D-glucopyranosyl residue; resonance b (79.1 p.p.m.) is identical to that for glycogen (and therefore linear amylose). and resonance c (77.9 p.p.m.) corresponds to the closely spaced doublet observed for methyl β -D-glucopyranoside. The corresponding methyl α -D-glucopyranoside has no resonances in the 70--75-p.p.m. region.

Extracellular mannans

In contrast to the above data, which for dextrans and amyloses

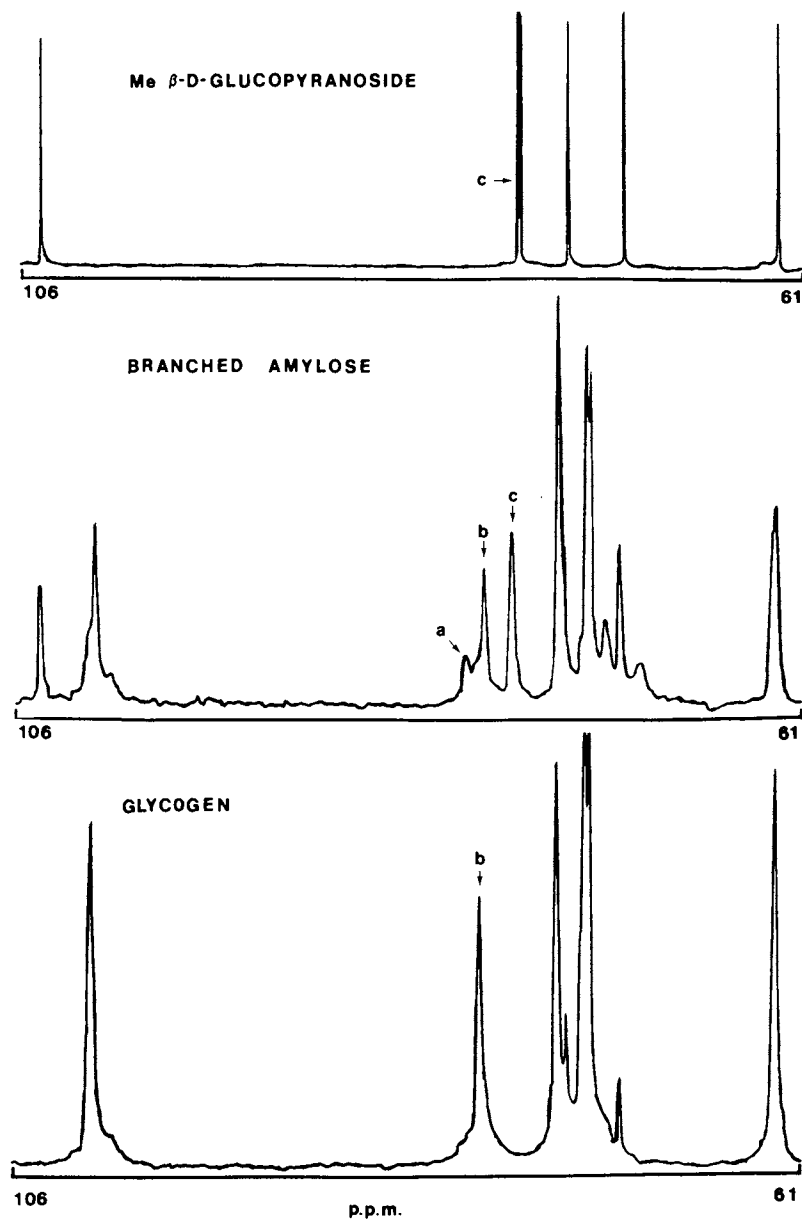
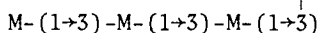
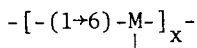


Figure 5. The 90° $\text{C-}^{13}\text{NMR}$ spectra of methyl α -D-glucopyranoside (the aglycon resonance at 55 ppm is not shown), a branched amylose (sample OAG-II-3), and rabbit liver glycogen

show a relatively simple structure-to-spectra relationship, the mannan spectra present greater difficulties in interpretation. This is quite possibly due to two different reasons. On the basis of permethylation g.l.c.-m.s. and h.p.l.c. acetolysis data the various mannan structures appear to display greater structural diversity than the dextrans (3). In addition, fewer minor structural variants (e.g. differences in degree of branching) are available for these mannans. In general, the mannans contain greater percentages of non-(1+6)-linkages than do the dextrans. Furthermore, permethylation g.l.c.-m.s. indicates that mannans, in contrast to the dextrans, often have these non-(1+6)-linkages associated with linear chain extending residues, rather than as branch-points associated with branched residues also containing (1+6)-linkages. Acetolysis data also indicates that many of these non-(1+6)-linked residues are sequentially linked. In accord with the dextran structures, we have previously proposed mannan structures which contain all (1+6)-linkages in the backbone -- though this may well not be the case for all or any of these polysaccharides. Two typical mannan spectra will now be examined to illustrate the differences between mannans and dextrans.

Mannan YB-1344 -- By combining the permethylation g.l.c.-m.s. analysis data with the postulate that all (1+6)-linkages are in a linear backbone, the following average repeating unit can be constructed (see Structure 5), where M represents the D-manno-

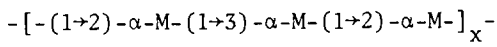


Structure 5 (all α -linkages)

pyranosyl residue. Such a structure differs radically from the essentially linear dextrans' structure which contain single residue branches. However, comparison of the mannan 75--85-p.p.m. diagnostic region to that of a typical (1+3)-linked dextran shows a profound difference. Where the dextrans display a single sharp resonance, mannan YB-1344 displays a series of overlapping resonances (see Figure 4). With the exception of the anomeric resonances, the total YB-1344 spectrum displays many more resonances than a comparable (similar degree of branching) dextran spectrum. For some reason the individual 3-O-linked α -D-mannopyranosyl residues are in a greater variety of environments than the dextran residues, and the logical inference from these data is that such environmental differences result from side-chains of greater length than a single saccharide residue.

Mannan Y-1842 -- Compared to previous dextran spectra-to-structure relationships, this polymer displays an unexpected spectrum. Careful permethylation g.l.c.-m.s. data indicate a 2:1 ratio of 2-O-substituted D-mannopyranosyl residues to 3-O-substituted D-mannopyranosyl residues, with very few branching and terminal residues. Such an assembly of residues yield the

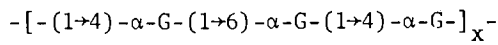
following average repeating unit (Structure 6). The (1→2)- and (1→3)-linkages have been shown in an ordered manner, but per-



Structure 6

methylation g.l.c.-m.s. data actually yield no information on the ordering of such residues. The permethylation g.l.c.-m.s. data also provide no information about the anomeric linkages of the residues. The spectra-to-structure relationship problem for this polymer is obvious, and is described as follows. When the hypotheses which were successfully applied to dextrans are applied in this case, the residues present, known from permethylation analysis, simply cannot give the observed spectrum. Two 2-O-substituted α -D-mannopyranosyl residues and one 3-O-substituted α -D-mannopyranosyl residue are expected to yield only two anomeric resonances, with the intensity of the former residue's resonances twice the intensity of the latter residue's resonances. Such an effect is not observed for the anomeric region of the mannan Y-1842 spectrum -- in fact three well spaced resonances of approximately equal intensity are observed (see Figure 4). Either the limiting situation for our working hypothesis has been reached, or mannan Y-1842 does not have the general structure we have assigned it. However, the combined data from permethylation fragmentation g.l.c.-m.s., specific rotations, and saccharide surveys by g.l.c.-m.s. and paper chromatography all support the presence of the specific type and ratio of saccharide residues indicated in Structure 6.

If the above mannan structures are properly understood, then the spectra-to-structure relationships must be reexamined. The permethylation g.l.c.-m.s. data show that the structure of mannan Y-1842 profoundly differs from the above dextran structures as this mannan has non-(1→6)-linkages incorporated into the backbone. Previous examination of pullulan, a D-glucan containing a 2:1 ratio of (1→4)- and (1→6)-linkages in the backbone, showed a resonance splitting of both the C-1 and C-4 resonances originating from the 4-O-substituted α -D-glucopyranosyl residues (but the C-1 resonance was split by only ~ 0.5 p.p.m.). This spectrum was convincingly interpreted by postulating pullulan to be a highly ordered polysaccharide of the following structure, and assuming



Structure 7

that those 4-O-substituted α -D-glucopyranosyl residues are in different chemical environments and will have different chemical shifts for various carbon atom positions.

Structures 6 and 7 are obviously very similar, both containing a 2:1 ratio of differently linked residues in a uniform pattern. This then may provide the explanation for the three

equal intensity anomeric resonances of mannan Y-1842 via the "before" and "after" 2-O-substituted α -D-glucopyranosyl residues. It is inherent in this assumption that mannan Y-1842 is a highly ordered linear polymer. Such a postulated structure needs to explain why the similar residue anomeric resonances of mannan Y-1842 are so much more separated than those of pullulan, and there are two possible explanations. Mannan Y-1842 contains linkages through the C-2 and C-3 positions, which are much closer to the anomeric carbon than the C-4 and C-6 linkages of pullulan, therefore, the effect observed for pullulan could be magnified for mannan Y-1842. In addition, the difference between α -D-glucopyranosyl and α -D-mannopyranosyl residues lies in the configuration of the C-2 position, with the α -D-mannopyranosyl containing the C-2 linkage in a position axial to the ring and closer to the C-1 position. For the above reasons, the C-1 resonance of the 2-O-substituted α -D-mannopyranosyl position could be quite dependent on the geometry of the adjacent linked saccharide.

The general thrust of the above arguments can be summed as follows. The chemical shifts of a given residue type are minimally affected by the nature of adjacent linked residues as long as those adjacent residues are not part of the polymer backbone or side-chain. However, once such different linkages are incorporated into a backbone chain, or short side-chain, then the relative positions of the residues can result in a mutual interaction displacing the chemical shifts associated with specific carbon positions. If these arguments are correct, they then explain why mannan spectra are more complex (and making the corresponding assignments is more complicated) than for the dextrans and amylose derivatives.

Levans and β -D-fructofuranosyl compounds

Though the above polymers have shown a wide variety of resonances and spectra-to-structure relationships, they have all had in common structures composed exclusively of hexapyranosyl residues. The levans, composed of β -D-fructofuranosyl residues, are of interest in that these polymers can be produced by the same microorganisms which produce dextrans, and also of interest for examining the structure-to-spectra relationships for this new ring system. The C-13 n.m.r. spectra of the α -D-glucopyranosyl and β -D-fructofuranosyl residues have an unusual relationship, for when the two spectra are compared there is essentially no overlap of resonances (see Figure 1). The resonances of dextran and levan are remarkably evenly spaced across the 60--110-p.p.m. region of the C-13 n.m.r. spectrum. The lack of development of g.l.c.-m.s. ketose techniques, comparable to those for aldose residues, make the general approach to establishing the branch-type data less feasible than for the dextrans and mannans. Furthermore, as will be seen below, the currently isolated β -D-fructofuranosyl polymers display much less variety of

structural features than do the D-glucans and D-mannans. However, the above resonance spacing of the β -D-fructofuranosyl residues allows an alternative approach to establishment of spectra-to-structure relationships (16).

The fortunate existence of a group of oligomers, which contain the β -D-fructofuranosyl residue as a central moiety being O-substituted at various positions by the α -D-glucopyranoside group, allows the study of the displacement of specific resonances due to this substitution. For example, 2-O-substitution of the β -D-fructofuranoside residue gives sucrose; 2,3-di-O-substitution gives melezitose. Employing $\Delta\delta$ of substitution data, plus chemical shift data from the 1,2-di-O-substituted β -D-fructofuranosyl residues of inulin, allow resonance assignment to the specific carbon positions of the β -D-fructofuranosyl residue. In general, it is concluded that O-substitution at the C-1 and C-6 positions of the β -D-fructofuranosyl residue results in $\Delta\delta$ displacements of only ~ 3 p.p.m., which are much smaller than the average $\Delta\delta$ of O-substitution for the α -D-glucopyranosyl residue. However, the C-1 and C-6 positions are primary alcohol positions out of the ring system, and a smaller than average $\Delta\delta$ of O-substitution was also observed for the α -D-glucopyranosyl C-6 resonances (which also is a primary alcohol position out of the ring). In addition, the resonance of the C-2 (anomeric) position of the β -D-fructofuranosyl residue gives evidence of being much less sensitive to structural change than the corresponding C-1 (anomeric) position of the α -D-glucopyranosyl residue.

The immediate application of this levan C-13 n.m.r. approach has been to analyze the levan fractions resulting from acid hydrolysis of native levan, and to survey a group of levans from the NRRL collection which were produced by diverse bacterial strains. We have concluded that acid hydrolysis has little effect on modifying the average degree of branching for resulting levan fragments. The group of NRRL levans exhibited a very close similarity of structure (especially when compared to the wide diversity of structure encountered in the corresponding NRRL dextran collection), but do show certain distinct differences in minor resonances which suggest that these levans differ in type of branch-point residues. The distinct differences in resonance position between dextran and levan C-13 n.m.r. spectra indicate that C-13 n.m.r. could be a very usable technique for quantitation of dextran-levan mixtures, and indeed it would appear that a large amount of structural evidence could be obtained for one class of these polymers in the presence of the other.

Resonance relaxation measurements

The above data have indicated that C-13 n.m.r. spectra can distinguish between subtle structural effects in polysaccharides, and that in many cases resonances can be assigned to specific

carbon atom positions. The information arising from the assignment of resonance to carbon atom position can be very important in establishing the effect of structural changes on resonance displacements. If such effects were clearly understood, then a C-13 n.m.r. spectrum could be accurately predicted for a given polymer, or conversely, an intimate knowledge of a specific polymer could be established solely on the basis of a C-13 n.m.r. spectrum. Furthermore, neither our g.l.c.-m.s. data, nor a simple C-13 n.m.r. spectrum, yield any evidence about the relative position of individual residues within a polymer. Fortunately, relaxation data provide certain evidence for both of these areas of uncertainty.

Analysis of the anomeric resonances produced due to branching can be very frustrating when based on the simple C-13 n.m.r. spectra. For the dextrans these anomeric branching resonances are well resolved one from another, and from the linear polymer resonance, but the introduction of a branch-point residue into a linear polysaccharide results in the creation of an additional terminal residue with its associated anomeric resonance. Therefore, each type of branching introduces two sets of new resonances into the spectrum -- one set from the branch-point residue, and one set from the terminal residue. This duplicate set of residue resonances causes confusion in resonance assignment as there is no way to separate and assign these sets of new resonances. As was previously noted, not all carbon atom positions can be expected to yield resonances of equal intensity in a given spectrum. The actual resonance intensity, and half-height width, is dependent on relaxation times (T_1), and on nuclear Overhauser effects (n.o.e.). The T_1 values for a specific resonance is essentially a measure of the relative degree of mobility within the polymer at the specific carbon atom position related to that resonance. The n.o.e. for a given resonance is a measure of carbon-proton dipole reorientation in competition with solvent interaction with a carbon atom associated with that resonance. It is possible to also calculate T_{1DD} , a function of T_1 and n.o.e., but for the polymers discussed here the relative sets of values for T_1 and T_{1DD} measurements parallel each other. The application of these concepts will be demonstrated by the following examples.

Dextran B-1299 fraction S yields a simple spectrum (17) (see Figure 1, upper spectrum). Permethylated g.l.c.-m.s. data indicate this polymer to have an average repeating unit represented by Structure 1, when $n=1$ -- the repeating unit then containing three saccharide residues. Again, as for all dextrans, the basic assumption that all (1 \rightarrow 6)-linkages are in the backbone is employed. The anomeric region shows three resonances of roughly equal intensity, and progressively more linear analogues of Structure 1 yield spectra (see Figure 4, dextran B-1402) demonstrating that resonance 1 corresponds to the chain extending 6-O-substituted α -D-glucopyranosyl residue. The

question now is, does resonance 2 correspond to the branching or terminal residue? As the terminal residue is not incorporated into the backbone, this residue should exhibit both the general motion of the chain, plus additional independent motion. As the T_1 value of each resonance is proportional to the relative freedom of the carbon atom associated with that resonance, it can be expected that the anomeric resonance associated with the terminal residue will have the largest T_1 value. Comparison of resonance 2 vs. resonance 3 of the dextran B-1299 fraction S spectrum clearly shows that the T_1 (241 millisecon) is larger than the T_1 (140 millisecon) of peak 3. Therefore, resonance 2 represents the anomeric resonance of the 2,6-di-O-substituted α -D-glucopyranosyl residue (17).

A second example (17) is that of the anomeric spectral region of dextran B-742 fraction S, a polysaccharide for which permethylation data indicate Structure 2, when $n=0$. This is an unusual polymer, as every backbone residue is 3-O-substituted. It is fortunate that this polymer exists, as the dextrans branching through 3,6-di-O-substituted residues present a problem in the anomeric spectral region, displaying only a single branching anomeric resonance in addition to the linear dextran resonance. It was therefore postulated that one of the two expected minor resonances was unresolved from the linear dextran anomeric resonance. For dextran B-742 fraction S there are essentially no normal linear dextran 6-O-substituted α -D-glucopyranosyl residues, only branch-point and terminal residues -- and one of the two approximately equal resonances appears at the same chemical shift as the linear dextran anomeric resonance. Resonance number 2 of dextran B-742 fraction S has a smaller T_1 (150 millisecon) than the corresponding resonance 1 ($T_1 = 250$ millisecon), and therefore resonance 2 represents the anomeric carbon of the 3,6-di-O-substituted α -D-glucopyranosyl residue. The extreme structural simplicity of dextran B-742 fraction S suggests another spectra-to-structure relationship. The two residue repeating unit of this dextran indicates that total resolution of the spectrum will result in twelve (2×6) resonances and the spectrum in Figure 6 comes close with eleven distinct resonances resolved. Due to a large intensity and a small T_1 value, resonance 8 is believed to be a composite resonance -- leaving 10 remaining resolved resonances. In terms of intensity, these ten resonances break into two sets, a weak intensity set and a strong intensity set. The strong intensity resonance set generally have larger T_1 values than the corresponding values for the weak intensity resonance set, indication that the weak intensity set of resonances correspond to the less mobile backbone residue carbon atom positions. This general assignment of resonances is supported by the independent knowledge that peak 3, representing a linked C-3, and peak 10, representing a linked C-6, must be associated with the 3,6-di-O-substituted α -D-glucopyranosyl backbone residue. These observations then

**American Chemical
Society Library**

1155 16th St. N. W.

In Carbon-13 NMR in Polymer Science; Pasika, W.;
ACS Symposium Series; American Chemical Society: Washington, DC, 1979.

Washington, D.C. 20036

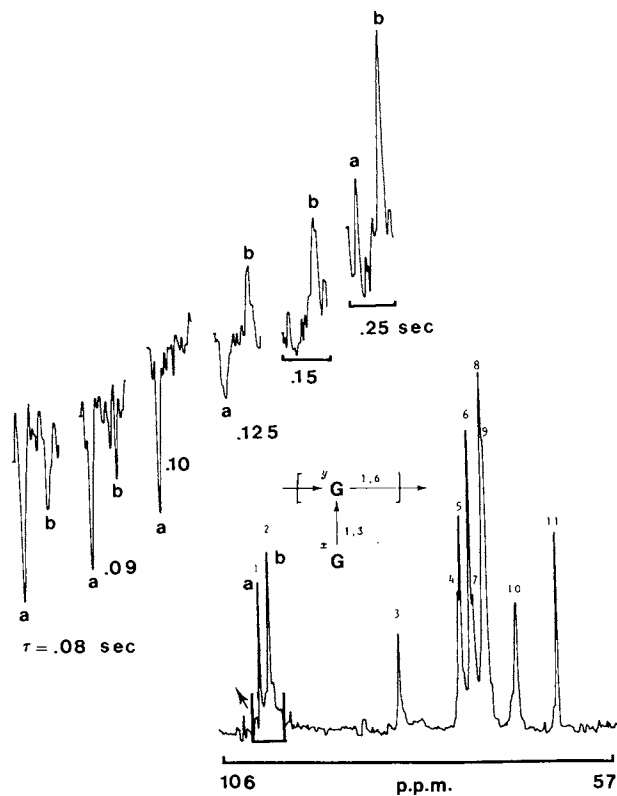


Figure 6. The 90° $\text{C-}^{13}\text{NMR}$ spectrum of dextran B-742 Fraction S, showing the relaxation spectra for Resonance a ($T_1 = 250$ msec) and for Resonance b ($T_2 = 150$ msec): a = 101.01 ppm; b = 99.76 ppm.

allow the introduction of another spectra-to-structure relationship, when resonances known to be associated with a specific residue are more intense than alternative structural data would indicate, then that residue has increased mobility and for a polymer is probably in a terminal or side chain position. For example, returning to the anomeric spectral region of dextran B-1299 fraction S, it can be noted that resonance 2 is more intense than resonance 3. Permethylation data indicate that all residue types are present in approximately equal amounts, therefore resonance 2 represents the C-1 of the terminal residue (in agreement with T_1 data). Such an approach can allow the tentative assignment of resonances when the more laborious determination of relaxation data is not feasible.

Such T_1 measurements have also been employed to differentiate between terminal and branch-point residue resonances for dextrans branching through the 4,6-di-O-substituted α -D-glucopyranosyl residue and for establishing that the excess 6-O-substituted α -D-glucopyranosyl residues of the class of compounds represented by structure 3 are in side chains relative to an alternating linkage type backbone (13).

It is known that relatively subtle solvent properties (e.g. the presence of trace metal ions or dissolved oxygen) can have a pronounced effect on T_1 values (18). For this reason, we have emphasized studies based on comparing relative T_1 values of resonances taken from the same spectrum of a given compound, rather than comparing absolute T_1 values taken from different spectra. To insure reproducibility, duplicate T_1 determinations were made in all cases. Monomer T_1 values (e.g. methyl α -D-glucopyranoside) can be obtained in less than an hour. However, we have experienced difficulty in obtaining consistent absolute T_1 values for successive samples of the same monosaccharide. Such reproducibility problems have not been observed for the polysaccharides, and we have observed no successive T_1 value differences which can be attributed to solvent or sample preparation.

The n.O.e. values have, in general, paralleled the T_1 values and provide little additional information. Increasing n.O.e. values reflect decreasing solvent interaction for the specific carbon atom position associated with the resonance studied. Interestingly, the largest n.O.e. values observed, ≈ 1.97 (near the theoretical maximum of 1.99 for this value) is associated with resonance 3 in the spectrum of dextran B-742 fraction S. This resonance 3 corresponds to the linked C-3 position of the branching backbone residue, and model building, or even a casual inspection of the drawn structure, indicates that this is an extremely hindered position.

General conclusions

The above discussion has indicated that C-13 n.m.r. data for a single polysaccharide can yield a large amount of structural

information. However, the correlation of C-13 n.m.r. data from a wide variety of polysaccharide structures can provide an even clearer insight into spectra-to-structure relationships, and also into the nature of the specific carbohydrate -- in fact few of the polysaccharides we have studied have had structures so well defined that C-13 n.m.r. could not add new information. The C-13 n.m.r. data have been especially welcome as they are extraordinarily complementary to data obtained from permethylation g.l.c.-m.s. As both C-13 n.m.r. and g.l.c.-m.s. data provide structural analysis in terms of residue types, rather than considering an assembly of isolated linkage types, an intimate view of the polymer structure has been obtained.

At present, C-13 n.m.r. studies are hampered by the relatively large sample sizes required for good signal-to-noise ratios, and for recording the multiple spectra required to obtain T_1 and n.O.e. data. Furthermore, the extremely large number of saccharide residues known to exist in polymers require further development in spectral analysis of other relatively unstudied residues -- and it is anticipated that increased spectral resolution will be required to differentiate between these new residue types. However, the recent history of n.m.r. has shown rapid advances in the introduction of spectrometers with dramatically increased sensitivity and resolution. There is no reason to believe that this pace of development will slacken in the near future. Therefore, the general tenor of this presentation has been to indicate that a reasonable entry into the area of C-13 n.m.r. spectra-to-structure relationships has been made, and that the future of these studies holds great promise for general polysaccharide structural elucidation.

Acknowledgements

This work was supported, in part, by a National Institutes of Health Grant HL-17372.

Literature cited

1. Seymour, F.R., in "Extracellular Microbial Polysaccharides", ACS Symposium Series No. 45, Sandford, P.A., and Laskin, A., Eds., American Chemical Society, Washington, D.C., 1977, pp 114-127.
2. Seymour, F.R., Plattner, R.D., and Slodki, M.E., Carbohydr. Res., (1975) 44, 181-198.
3. Seymour, F.R., Slodki, M.E., Plattner, R.D., and Stodola, R.M. Carbohydr. Res., (1976) 48, 225-237.
4. Seymour, F.R., Slodki, M.E., Plattner, R.D., and Jeanes, A., Carbohydr. Res., (1977) 53, 153-166.
5. Seymour, F.R., Chen, E.C.M., and Bishop, S.H., Carbohydr. Res., (Unusual dextrans III), (1979) 68, 113-121.
6. Seymour, F.R., Knapp, R.D., and Bishop, S.H., Carbohydr. Res.,

- (1976) 51, 179-194.
7. Seymour, F.R., Knapp, R.D., Bishop, S.H., and A. Jeanes, *Carbohydr. Res.*, (Unusual dextrans IV), 68 (1979) 23-140.
 8. Costello, A.J.R., Glonek, J., Slodki, M.E., and Seymour, F.R. *Carbohydr. Res.*, (1975) 42, 23-37.
 9. Sidebotham, R.L., *Adv. Carbohydr. Chem. Biochem.*, (1974) 30, 371-444.
 10. Jeanes, A., "Dextran Bibliography: 1861-1976" Miscellaneous Publication No. 1355, Agriculture Research Service, U.S. Department of Agriculture, 1978, 370 pp.
 11. Seymour, F.R., Knapp, R.D., Chen, E.C.M., Jeanes, A., and Bishop, S.H., *Carbohydr. Res.*, (Unusual dextrans V) in press.
 12. Jennings, H., and Smith, I.C.P., *J. Am. Chem. Soc.*, (1973) 95, 606-608.
 13. Seymour, F.R., Knapp, R.D., Chen, E.C.M., Bishop, S.H., and Jeanes, A., *Carbohydr. Res.*, (Unusual dextrans VIII) in press.
 14. Jeanes, A., and Seymour, F.R., *Carbohydr. Res.*, (Unusual dextrans VII) in press.
 15. Seymour, F.R., Knapp, R.D., Nelson, T.E., and Pfannemüller, B. *Carbohydr. Res.*, in press.
 16. Seymour, F.R., Knapp, R.D., Zweig, J., and Bishop, S.H., *Carbohydr. Res.*, in press.
 17. Seymour, F.R., Knapp, R.D., and Bishop, S.H., *Carbohydr. Res.*, (Unusual dextrans VI) in press.
 18. Stothers, J.B., "Carbon-13 Spectroscopy", Academic Press, New York, 1972, p 38.

RECEIVED March 13, 1979.

A Reinvestigation of the Structure of Poly(dichlorophenylene oxides) Using Carbon-13 NMR Spectroscopy (1)

JOHN F. HARROD and PATRICK VAN GHELUWE

Chemistry Department, McGill University, 801 Sherbrooke St. West, Montreal, P.Q., H3A 2K6 Canada

Since their original synthesis by Hunter in 1917 (2) the poly(halophenyleneoxides) have been the subject of sporadic interest in a number of laboratories (3, 4, 5, 6, 7). In spite of their structural similarity to the useful and commercially successful poly(alkylphenyleneoxides) (8), the known halogensubstituted poly(phenyleneoxides) have poor mechanical properties and have found no commercial application. At the outset of the present study it was generally believed that the physical properties of the known polymers were impaired by a high degree of branching.

The branching hypothesis was originally based on what were thought to be anomalously low intrinsic viscosities (3) and was later reinforced by the observation of complex ^1H -nmr spectra (4), examples of which can be seen in Figure 1. Our first suspicion that the branching hypothesis might not be correct arose from the observation that a Mark-Houwink plot of a considerable amount of viscosity/ \bar{M}_n data yielded parameters that were not unreasonable for a linear polymer. More importantly, some data points for polymers prepared from 4-bromo-2,6-dichlorophenoxide, known to be linear, fell on the same line as the points for supposedly branched polymers, prepared from trichlorophenoxide. (Figure 2). We therefore decided to reinvestigate the structure of poly(dichlorophenyleneoxide) using ^{13}C -nmr spectroscopy.

Results

Molecular weight effects on ^1H and ^{13}C spectra. The ^1H -nmr spectra of three samples of poly(dichlorophenyleneoxide) prepared from a copper(II)trichlorophenoxide complex and covering a molecular weight range of 5,000 to 30,000 Daltons are shown in Figure 1. By themselves it is not possible to decide whether the differences in these spectra are due to molecular weight effects or to structural differences between the various polymers. The ^{13}C -spectra of a low and a high \bar{M}_n polymer, shown in Fig.3c & d exhibit the same resonances, albeit with different linewidths and relative intensities, thereby indicating that the differences in the proton spectra are due to dynamic rather than structural effects.

Comparison of spectra of "linear" and "branched" polymers.

0-8412-0505-1/79/47-103-053\$05.00/0

© 1979 American Chemical Society

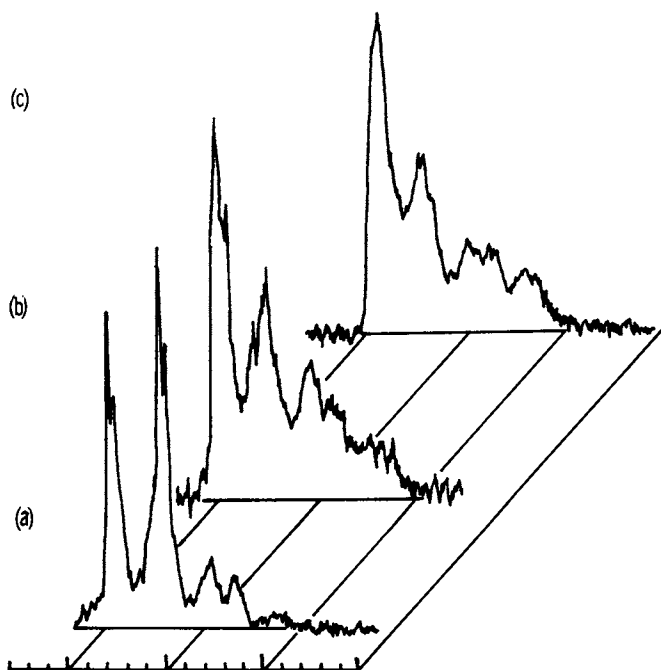


Figure 1. Effect of \bar{M}_n on $^1\text{H-NMR}$ spectra of polymers derived from trichlorophenoxide. $\bar{M}_n = 6,000$ (a), 13,300 (b), and 33,000 (c).

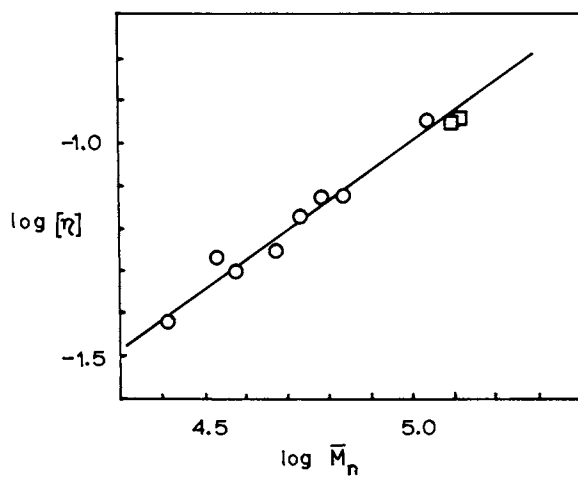


Figure 2. Mark-Houwink plot for polymers obtained from trichlorophenoxide (○) and 4-bromo-2,6-dichlorophenoxide (□)

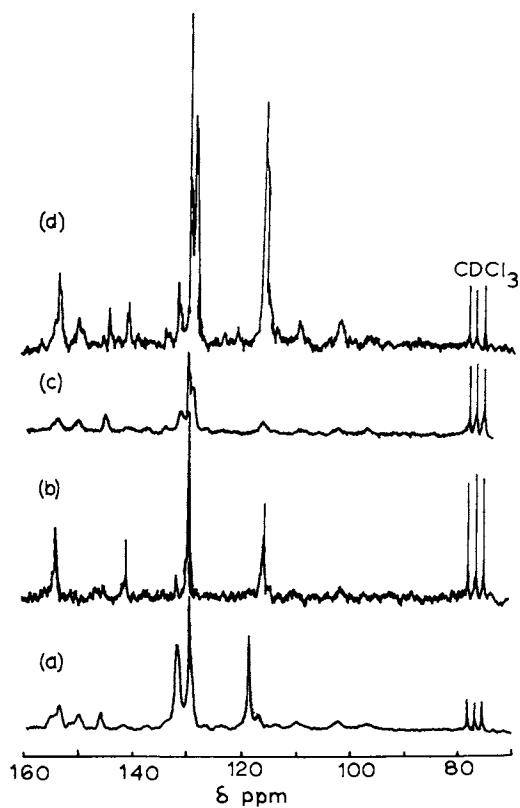
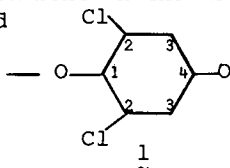
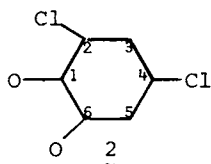


Figure 3. $C-13$ NMR spectra of polymers derived from (a) 2-bromo-4,6-dichlorophenoxide, (b) 4-bromo-2,6-dichlorophenoxide, (c) 2,4,6-trichlorophenoxide ($\bar{M}_n \sim 30,000$), and (d) 2,4,6-trichlorophenoxide ($\bar{M}_n = 5,000$)

As previously reported (4) linear poly(2,6-dichloro-1,4-phenylene oxide) exhibits the expected sharp singlet in its ^1H -nmr spectrum, together with some minor peaks due to chemical asperities in the polymer. The ^{13}C spectrum of this polymer (Figure 3) is also very simple, consisting of the expected four sharp lines of the four inequivalent carbons in the mer unit 1. These same four lines may be identified



spectra of the "branched" polymers, especially that of the low molecular weight material. Subtraction of the spectrum of 1 from that of the low \bar{M}_n "branched" polymer leaves a much simpler spectrum than would be expected for an irregular, branched polymer. In fact, six of the residual lines are exactly what one would expect for the 1,2-coupled mer unit 2. It was therefore concluded



that the so called "branched" polymer was in fact largely a copolymer of units 1 and 2 and that any unusual behaviour was a result of the properties of unit 2 rather than branching.

The polymer derived from 2-bromo-4,6-dichlorophenoxide and some copolymers. Polymerization of 4-bromo-2,6-dichlorophenoxide yields a polymer with a high proportion of units 1, indicating a high selectivity for displacement of bromine rather than chlorine (4,5). It was therefore anticipated that polymer derived from 2-bromo-4,6-dichlorophenoxide would contain a high proportion of units 2. This was not the case however since elemental analysis indicated that roughly half of the mer units in such polymers still carried bromine. The ^1H -nmr spectra of such polymers were indistinguishable from those of high \bar{M}_n polymers derived from 2,4,6-trichlorophenoxide. The ^{13}C spectra were very similar (Figure 3) except for the absence (or shifting) of a resonance at ca. 130 ppm. and for the presence of a strong resonance at 118.6 ppm in the bromine containing polymer.

A copolymer from equimolar 2,4,6-trichlorophenoxide and 4-bromo-2,6-dichlorophenoxide with $\bar{M}_n > 50,000$ gave both ^1H and ^{13}C spectra essentially identical to those of homopolymer from 2,4,6-trichlorophenoxide with $\bar{M}_n < 6000$. This copolymer contained only a trace of bromine by chemical analysis.

A copolymer from equimolar 2-bromo-4,6-dichlorophenoxide and 4-bromo-2,6-dichlorophenoxide of $\bar{M}_n > 50,000$ gave an ^1H spectrum

very similar to that of the homopolymer from trichlorophenoxide with $M_n < 6000$ and a ^{13}C spectrum very similar to that of the 2-bromophenoxide homopolymer. The copolymer spectrum was characterized by sharper resonances and a considerable enhancement in intensity of the line at 116 ppm. Both chemical analysis and the ^{13}C spectrum showed a level of bromine retention per 2-bromo unit comparable to that of the 2-bromophenoxide homopolymer. The spectra of the two copolymers are shown in Figure 4.

Effects of temperature and paramagnetic additives on the ^1H spectrum of poly(dichlorophenyleneoxide). The spectra of the polymers referred to in Figure 1 were all measured up to 120° in tetrachloroethylene. In no case did any significant change occur in the spectrum on heating.

The spectrum of a high molecular weight polymer was measured in the presence of tris(acetylacetonato)chromium(III). The results for several different concentrations of reagent are shown in Figure 5.

Discussion

The structure of poly(dichlorophenyleneoxide). Contrary to earlier conclusions, the present evidence shows that branching is not a major structural feature of poly(dichlorophenyleneoxide). On the other hand it seems reasonably clear that polymer prepared from 2,4,6-trichlorophenoxide contains substantial amounts of both 1,2- and 1,4-linked phenylene units. On the basis of this assumption all of the peaks in the ^{13}C spectrum can be assigned by analogy with chemically similar compounds and by the application of additivity rules modified to take account of the shielding effects of ethereal oxygen (9, 10, 11, 12). Such a tentative assignment is shown in Figure 6.

Following the above conclusion it is clear that the rather bizarre ^1H spectra of these polymers derive from the special features of 1,2-enchainment. Examination of molecular models reveals that runs of 1,2-enchained segments are considerably more restricted in their degrees of motional freedom than are runs of 1,4-enchained segments. The restriction arises partly from the absence of a "crankshaft" mode with 1,2-enchainment and partly from the steric interference of substituents on adjacent phenylene rings. It is probable that the 1,2-coupled units are essentially locked in position and can only move as the whole polymer molecule moves. The great complexity of the ^1H nmr spectra of polymers containing 1,2-coupled units could be due to the locking of chain units in a limited number of different conformations, each with a characteristic chemical shift.

The insensitivity of the ^1H nmr spectra to large changes in temperature is also indicative that the frozen backbone motions responsible for the spectral complexity are not easily thermally activated. The presence of chemically different triads composed of 1,2 and 1,4 units, each with a particular chemical shift, could

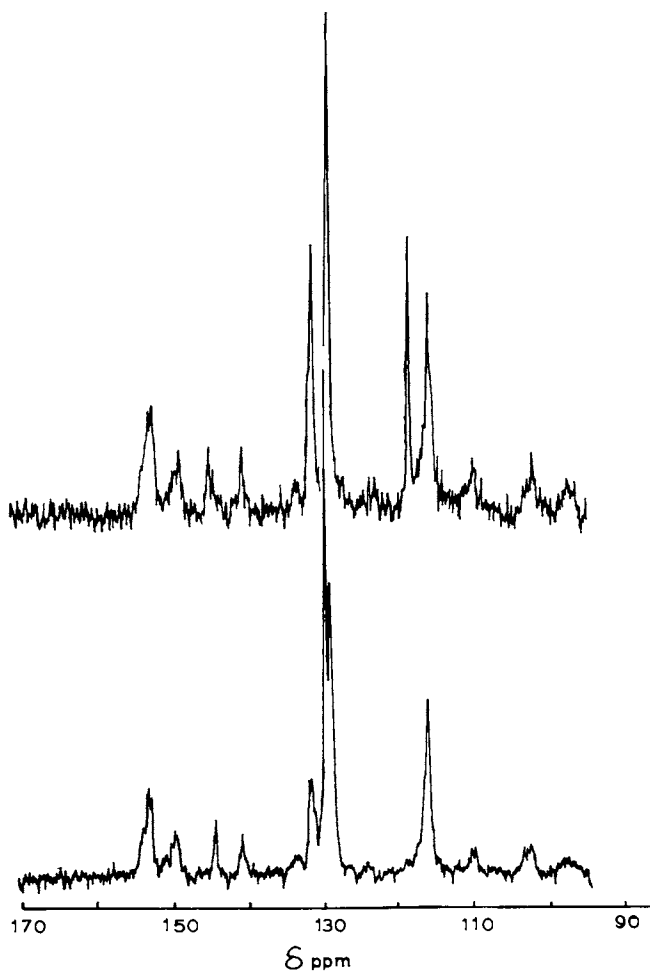


Figure 4. $C-13$ NMR spectra of copolymers derived from: (upper) 2-bromo-4,6-dichlorophenoxide and 4-bromo-2,6-dichlorophenoxide (1:1) and (lower) 2,4,6-Trichlorophenoxide and 4-bromo-2,6-dichlorophenoxide (1:1).

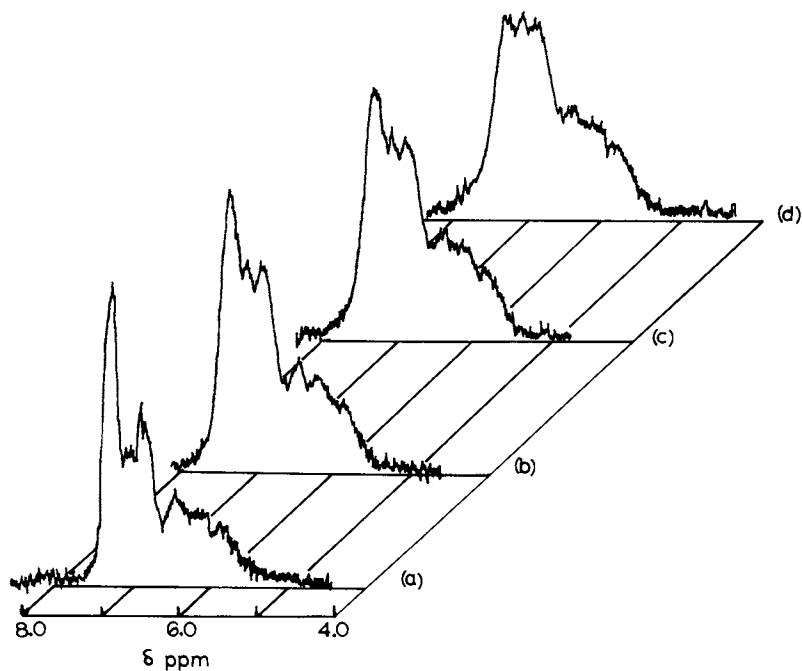


Figure 5. Effect of added $Cr(acac)_3$ on the 1H -NMR spectrum of polymer derived from 2,4,6-trichlorophenoxide. $[Cr(acac)_3] = 0M$ (a); $5.7 \times 10^{-3}M$ (b); $1.2 \times 10^{-2}M$ (c); $1.7 \times 10^{-1}M$ (d). Polymer concentration, 17% in sym-tetrachloroethylene.

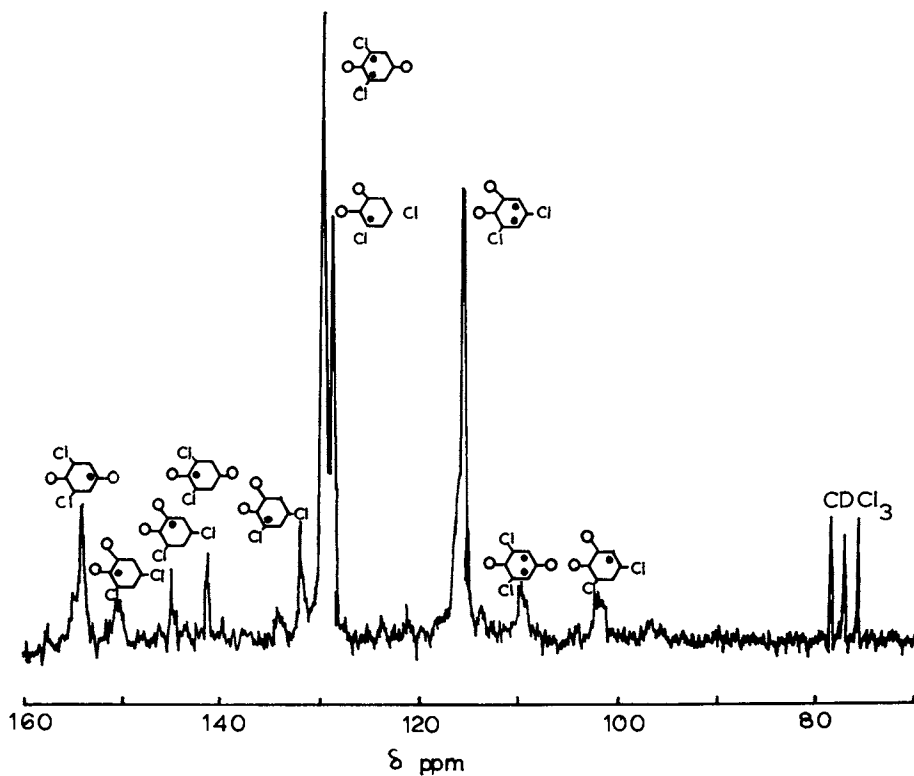


Figure 6. Assignment of resonances in ^{13}C NMR spectrum of low-molecular-weight polymer from 2,4,6-trichlorophenoxide

also complicate the spectrum.

All of the polymers have resonances at unusually high field ($>3.0 \tau$) suggesting that some of the protons are held in close proximity to adjacent aromatic rings. This possibility is further supported by the observation that relaxation reagents selectively collapse the lowest field peak, which is expected to be the resonance of protons least encumbered by neighboring aromatic rings and therefore more accessible to the relaxation reagent.

Unfortunately, we have found no way to estimate the relative amounts of 1,4 and 1,2-coupled units in these polymers. There are however some curious features of the results obtained with copolymers which suggest the operation of unusual selectivities in the polymerization reactions. For example, the copolymer resulting from polymerization of equimolar 4-bromo-2,6-dichlorophenoxide and 2,4,6-trichlorophenoxide ($M_n > 50,000$) gave both an ^1H and a ^{13}C spectrum very similar to that of a polymer from trichlorophenoxide with a much lower molecular weight. In the case of the ^1H spectrum, the copolymer was quite different from a mixture of polymers derived separately from the two monomers, particularly with respect to the great reduction in the sharp singlet characteristic of polymer with a high proportion of 1,4-units (4). The ^{13}C spectrum of the copolymer is a superposition of the spectra of the separately prepared polymers with respect to number and position of peaks, but the resonances are much sharper than those of polymer derived from 2,4,6-trichlorophenoxide and broader than those derived from 4-bromo-2,6-dichlorophenoxide (for an equivalent molecular weight). It thus appears that the deliberate introduction of more 1,4-units into the polymer chain, although leading to greater internal mobility, does not change the chemical composition in such a way as to seriously affect the chemical shifts and relative intensities of the ^{13}C resonances. This evidence suggests that the deliberate introduction of a considerable number of 1,4 units does not significantly increase the concentration of 1,4-triads (the entity which presumably is primarily responsible for the ^1H and ^{13}C spectral features of the 1,4-polymer). If this interpretation is correct it means that the copolymer sequence is far from random.

The behaviour of the copolymer derived from 2-bromo- and 4-bromophenoxides was similar to that described in the previous paragraph.

Regioselectivity in the polymerization reactions. The currently accepted mechanism for the reactions used to synthesise the polymers described in the present paper involves three key steps (13):

i) a ligand transfer of phenoxyl from Cu^{II} to an attacking radical.

ii) homolytic dissociation of a quinol ether.

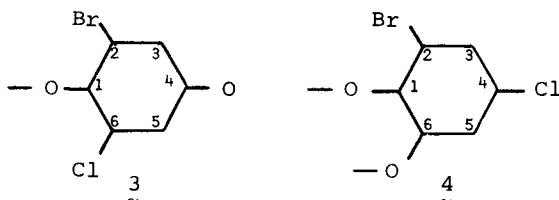
iii) halogen atom transfer from quinol ether to Cu^{I} .

It is expected that all three of these steps will be sensitive to

the nature of the halogen and the location of substituents in the various reacting species. Only for step (iii) can we say with any confidence what the effect will be, namely that bromine will transfer more rapidly than chlorine (14).

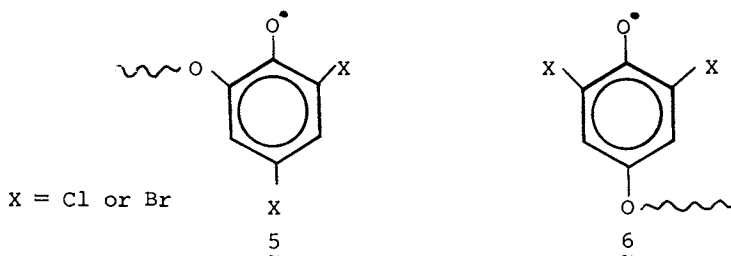
The high 1,4 content of polymers produced from 4-bromo-2,6-dichlorophenoxide is in accord with the expectation of easier access to the 4- relative to the 2-position in step (i) and greater reactivity of bromine in step (iii). The failure to achieve selectivity in the 2-bromo-4,6-dichlorophenoxide reaction could be attributed to the steric inaccessibility of the 2-position for step (i) overriding the greater reactivity of bromine in step (iii). This argument is not very convincing since it would lead to the conclusion that the polymer from 2,4,6-trichlorophenoxide be largely 1,4-coupled, contrary to the experimental evidence.

Chemical analysis of the polymer from 2-bromo-4,6-dichlorophenoxide revealed that somewhere between one half and two thirds of the available bromine was displaced. The presence of residual bromine in the polymer is also manifest by the sharp ^{13}C resonance at 118.6 ppm. This residual bromine is also the likely cause of the characteristic difference in the 130 ppm region between polymers derived from 2-bromo-4,6-dichloro (2 peaks) and 2,4,6-trichlorophenoxide (3 peaks). In addition to the units 1 and 2, the 2-bromophenoxide could also contain units 3 and 4.



Intuitively one would expect the greater ease of displacement of bromine to strongly favour units 2 over 4 and that most of the residual bromine would be present in units 3. This could lead to the conclusion that the "missing" peak is due to the replacement of units 1 by units 3 in the 2-bromo, relative to the 2,4,6-trichlorophenoxide derived polymer. Unfortunately this hypothesis is not consistent with the spectrum of the 2-bromo/4-bromophenoxide copolymer which, despite the undoubted inclusion of many units 1, only has two peaks in the 130 ppm region.

One postulate which does resolve many of the inconsistencies in the present data base is that there is a strong tendency towards alternation in these polymers. Such alternation could result from a combination of an intrinsically greater reactivity of the ortho-relative to para-positions of coordinated phenoxide, coupled with an inability of radicals of the type 5 to attack coordinated phenoxide at the ortho-position due to steric blocking. (An examination of models shows this to be a plausible assumption).

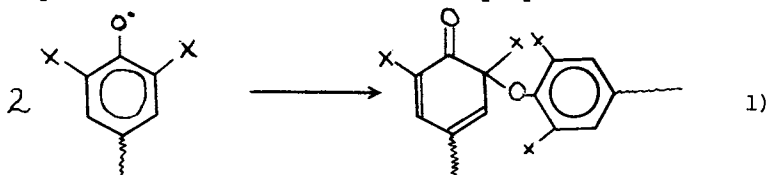


In the case of trichlorophenoxide such a model would give rise to an alternating 1,2-/1,4-sequence (which we call syndioregic by analogy with syndiotactic). In the case of 4-bromo-2,6-dichlorophenoxide the much greater reactivity of the bromine relative to chlorine overrides the factor which favoured ortho over para reactivity in the trichlorophenoxide case to such an extent that 1,4 catenation results. In the case of 2-bromo-4,6-dichlorophenoxide the 2-bromo position should be very reactive, but the radicals produced by such reaction (type 5) are not able to attack again at the ortho position of coordinated phenoxide.

The advantage of the alternation postulate is that it resolves the anomaly that the deliberate introduction of large numbers of units 1 into polymers derived from either 2,4,6-trichlorophenoxide, or from 2-bromo-4,6-dichlorophenoxide, whilst sharpening the spectra does not radically change the characteristic numbers and positions of the homopolymer resonances. A tendency to alternate would suppress both the generation of longer runs of 1,4-coupled units and the expression of the presence of such runs in the ^1H and ^{13}C spectra.

A clearer understanding of the structures of these polymers will require synthesis of model compounds for the triad and tetrad sequences. Only with such models can more confident assignments of the nmr spectra be made. Despite the tediousness of such syntheses we believe the postulate of syndioregic polymerization to be sufficiently interesting to warrant the effort and are presently working on the problem.

Branching and the polymerization mechanism. The realisation that the unusual properties of poly(dihalophenyleneoxides) are due to 1,2-coupling rather than branching removes an anomaly from the previously proposed mechanism for the polymerization reaction (13). Following suggestions of earlier workers (3) it was assumed that branching would occur by coupling of two polymer radicals according to equation (1). However, since the polymer radicals



are rapidly scavenged by attack on coordinated phenoxide until the reactant copper phenoxide has been almost completely removed, equation (1) cannot assume any importance until late in the reaction. Since displacement of coordinated phenoxide cannot lead to branching, it would be expected that the degree of branching would increase progressively as the reaction proceeded. This was contrary to earlier experimental evidence which seemed to indicate little significant change in what was assumed to be branching, from the earliest to the latest stage of reaction.

The above anomaly is removed by the recognition that branching is not a significant factor in the reaction.

Conclusions

The polymerization of halophenoxides by copper (II) mediated halide displacement is a mechanistically complicated reaction. Elucidation of the structure of the polymers is essential to an understanding of both the polymerization chemistry and the peculiar physical properties of the polymers. The physical tool which has yielded most information on the polymer structure is ^{13}C nmr. The first conclusion which derives from a study of the ^{13}C spectra of poly(dihalophenyleneoxides) is that regioselectivity in halogen displacement is more likely the source of the polymer properties than branching. A more rigorous confirmation of the polymer structures will depend on a detailed analysis of the ^{13}C spectra of model compounds for the chain segments.

Experimental

Synthesis of polymers. All of the polymers used in the present study were produced by methods previously described (4), or by modifications thereof. A more detailed description of the synthesis and characterisation of these polymers is in preparation.

^1H nmr spectra. Proton spectra were measured with a Varian T-60 spectrometer at 35° . Solutions were prepared in CS_2 . Low molecular weight polymers were typically examined as concentrated solutions (up to 20 per cent by weight) without encountering problems with solution viscosity. High molecular weight polymers, particularly those with high proportions of 1,4-coupled units had to be run at much lower concentrations due to viscosity problems. A few test runs on a Varian HA-100 spectrometer did not yield spectra of better quality than those obtained at 60 MHz.

Variable temperature measurements were carried out on a Varian A-60 spectrometer with a Varian-4060 probe heater attachment. Probe temperatures were calibrated with a standard ethylene glycol sample. Samples were prepared by dissolving up to 25 per cent by weight of polymer in tetrachloroethylene. Even with such concentrated solutions no viscosity problem was encountered above 60° .

^{13}C nmr spectra. All ^{13}C spectra were measured with a Bruker WH90 22.63 MHz pulsed FT instrument equipped with a Nicolet BNC-12

computer with 4K real data points. Probe temperature was maintained at 30°. Samples were prepared as 10 per cent solutions in deuteriochloroform, which also served as the frequency lock. Chemical shifts were computed relative to an internal standard of tetramethylsilane. Spectra were proton decoupled with 5W broad band irradiation.

Acknowledgements. The authors thank the National Research Council of Canada for financial support and for a Scholarship (P.v.G.). The assistance of Dr. Gordon Hamer in the measurement of ^{13}C spectra is gratefully acknowledged.

Literature cited.

1. Part V in a series Chemistry of Phenoxo Complexes. Part IV, Harrod J.F. and Taylor K.R. Inorg. Chem. (1975) 14, 1541.
2. Hunter W.H. and Joyce F.E., J. Am. Chem. Soc. (1917), 39, 2640
3. Blanchard H.S., Finkbeiner H. and Russell G.A., J. Polymer Sci. (1962), 58, 469.
4. Carr B.G., Harrod J.F. and van Gheluwe P., Macromolecules (1973), 6, 498.
5. Stamatoff G.S. U.S. Patent No. 3,228,910 (Jan. 11, 1966).
6. Tsuruya S. and Yonezawa T. J. Catalysis, (1975), 36, 48.
7. Hedayatullah M. and Denivelle L., Compt. Rend., (1962), 254, 2369.
8. Hay A.S., Polym. Eng. Sci. (1976), 16, 1.
9. JEOL ^{13}C FT-nmr spectra, Vol. 4.
10. Wilson N.K., J. Am. Chem. Soc. (1975), 97, 3573.
11. Dahmi K.S. and Stothers J.B., Can. J. Chem. (1966), 44, 2855.
12. Lauterbur P., J. Am. Chem. Soc. (1961), 83, 1846.
13. Carr B.G. and Harrod J.F., ibid, (1973), 95, 5707.
14. Kochi J., Science, (1967), 155, 415.

RECEIVED March 13, 1979.

Carbon-13 NMR in Organic Solids: The Potential for Polymer Characterization

A. N. GARROWAY, W. B. MONIZ, and H. A. RESING

Chemistry Division, Naval Research Laboratory, Washington, DC 20375

This Symposium bears witness to the analytical power of ^{13}C NMR as applied to organic polymers in the liquid state. But what of the solid state: can a polymer's chemistry or perhaps even engineering properties be extracted from solid state studies? A promising technique for such studies has recently evolved from the combination (1-4) of two well established NMR tools: cross-polarization (5) with dipolar decoupling (6) (proton enhanced ^{13}C NMR (7)) and magic angle spinning (8-11). We shall outline the method and illustrate with results for epoxy polymers. Emphasis will be on NMR, for not only the potential but also the limitations of this approach should be recognized. At the outset we state that while solid state techniques produce narrow spectra and can formally imitate relaxation experiments which are well-established in the liquid state, the nature of the solid specimen may dictate (i) a fundamental restriction to spectral resolution which is independent of spectrometer limitations and (ii) a competition between spin-spin and spin-lattice processes which may hopelessly obscure the connection between the observed rotating frame relaxation rate and molecular motions. We examine first the solid state techniques, then spectral resolution and finally relaxation effects.

MAGIC ANGLE SPINNING

In liquids rapid molecular motion serves to isolate nuclear spins from one another as the isotropic average of nuclear dipolar interactions vanishes. Also the chemical shift tensor collapses to its isotropic value. In a solid such fast isotropic motions are absent, although a partial averaging may occur through anisotropic motions, as in liquid crystals or rotating methyl groups. The averaging of these interactions provides the NMR definition of a liquid, a definition which may be at odds with the mechanical notion that a liquid cannot support a shear stress. In solids some other means must be found to reduce dipolar and chemical shift broadening (12-13).

In an organic solid representative broadenings are 150 ppm for aromatic carbon chemical shift anisotropy and 25 kHz (full width at half-height) for a rather strong carbon-proton dipolar interaction. At a carbon Larmor frequency of 15 MHz, the shift anisotropy corresponds to 2.25 kHz. In high magnetic fields the forms of the respective Hamiltonians are

$$H_{CS} = \hbar\omega_0s \sum_j \sigma_{zzj} S_{zj}, \quad (1)$$

This chapter not subject to U.S. copyright.
Published 1979 American Chemical Society

$$H_D = \hbar^2 \sum_{j < k} \gamma_I \gamma_S I_{zj} S_{zk} (1 - 3 \cos^2 \theta_{jk}) r_{jk}^{-3}, \quad (2)$$

where I_z , S_z are the proton and carbon spin operators along the static field direction z and r_{jk} , θ_{jk} determine the orientation of nuclear pairs with respect to z . Other terms have their usual meaning. For each nucleus the chemical shift σ_{zj} represents the projection of the chemical shift tensor along z :

$$\sigma_{zj} = \lambda_{1j}^2 \sigma_{1j} + \lambda_{2j}^2 \sigma_{2j} + \lambda_{3j}^2 \sigma_{3j}, \quad (3)$$

where $\sigma_{1j} - \sigma_{3j}$ are the principal values of the shift tensor and $\lambda_{1j} - \lambda_{3j}$ are the direction cosines orienting the tensor relative to the static field. If the rigid specimen is mechanically spun at the angle Θ to z , then the time average of Eq. (3) is (9)

$$\bar{\sigma}_{zj} = \frac{3}{2} \sin^2 \Theta \sigma_{0j} + \frac{1}{2} (3 \cos^2 \Theta - 1) \sigma_{\Theta\Theta j}. \quad (4)$$

Here σ_{0j} is the isotropic shift ($1/3 [\sigma_{1j} + \sigma_{2j} + \sigma_{3j}]$) and $\sigma_{\Theta\Theta}$ is the projection of the chemical shift tensor along the spinning axis and defined analogously to Eq. (3). In general this latter term produces a powder pattern when summed over nuclei at all orientations. However, when $\cos \Theta = 1/\sqrt{3}$, the powder pattern collapses and only the isotropic value remains, i.e., $\bar{\sigma}_{zj} = \sigma_{0j}$. If this magic angle is misset by ϵ radians, then (9)

$$\bar{\sigma}_{zj} \approx \sigma_{0j} - \sqrt{2}\epsilon [\sigma_{\Theta\Theta j} - \sigma_{0j}]. \quad (5)$$

The second term leads to a vestigial powder pattern, scaled down by $\sqrt{2}\epsilon$. The rule of thumb becomes: to reduce chemical shift anisotropy by a factor of 100, set the angle to within about $\pm 1/2$ degree.

Though spectral resolution is sensitive to orientation, it is insensitive to spinning rate. Chemical shift is an inhomogeneous broadening mechanism; as the spins dephase in a coherent manner, the phase information can be recovered by a refocussing technique. The classic example of inhomogeneous broadening is the dephasing of a liquid free induction decay in an inhomogeneous static field. Here the signal can be recovered by a Carr-Purcell pulse sequence (14) with pulses applied at a rate determined by the "real" T_2 rather than by the field inhomogeneity. A similar criterion applies for magic angle spinning; the spinning rate need only be greater than "real" or homogeneous broadening and not necessarily greater than the chemical shift anisotropy (15). When the spinning is slow, distinct sidebands appear and in the frequency domain they trace out a tent-like pattern related to the spatial anisotropy (3, 16). With increasing spinning rate, the sidebands move out and diminish in amplitude; it is convenient though not essential (16) to avoid complications of the sidebands by spinning at or above the frequency dictated by the width of the anisotropy pattern. For carbon at 15 MHz, a spinning rate of 2 kHz is adequate.

Magic angle spinning originated as a method to remove homonuclear dipolar broadening (8-11). There, however, speed is a necessity, and the technical problems of spinning above 10 kHz (with a centrifugal acceleration of $2 \times 10^6 g$'s on a 1 cm diameter sample) require great care. At 2-3 kHz, spinning is rather straight forward and conventional materials (Kel-F, Delrin, Macor and boron nitride) have been used to fabricate the rotors. Alternatively, the specimen itself can be directly machined or cast in a mold; that approach was used for the epoxies discussed here.

Two rather different spinner geometrics have evolved. The Beams (17) rotor, popularized by Andrew (9), looks like a spinning top cradled in a close fitting stator containing air jets. The Lowe and Norberg (10,11) geometry uses a right cylindrical disk supported by an axle along its center.

DIPOLAR DECOUPLING

Proton-carbon dipolar coupling can be reduced by irradiating the protons near or on resonance with a very strong rf field. Originally called spin stirring (6), this is also known as dipolar decoupling or high power decoupling to distinguish it from the scalar decoupling of liquid state ^{13}C NMR. If $\langle \omega_{\text{CH}}^2 \rangle$ is the second moment due to heteronuclear dipolar coupling, then, on irradiating protons spins $\Delta\omega_{0\text{H}}$ away from their resonance with rf field $\gamma_{\text{H}} B_{1\text{H}} \equiv \omega_{1\text{H}}$, the broadening collapses as (6)

$$\Delta \langle \omega_{\text{CH}}^2 \rangle^{1/2} = \langle \omega_{\text{CH}}^2 \rangle^{1/2} \cos \psi, \quad (6)$$

where $\tan^{-1} \psi \equiv \omega_{1\text{H}}/\Delta\omega_{0\text{H}}$ and $\psi = \frac{\pi}{2}$ is the "magic angle" for dipolar decoupling.

For a small deviation ϵ' away from $\psi = \frac{\pi}{2}$, the residual broadening is

$$\Delta \langle \omega_{\text{CH}}^2 \rangle^{1/2} \approx \epsilon' \langle \omega_{\text{CH}}^2 \rangle^{1/2}. \quad (7)$$

A single proton species can be irradiated exactly on resonance; in general the spread in Larmor frequencies is determined by the breadth of the chemical shift spectrum $|\Delta\sigma_{\text{H}}| \omega_{0\text{H}}$. Chemical shift anisotropy must be included, *even* if the sample is spun. If the protons are irradiated at the center of their spectrum, then $\epsilon' \approx \frac{1}{2} |\Delta\sigma_{\text{H}}| \omega_{1\text{H}}$. For an organic solid, take $|\Delta\sigma_{\text{H}}|$ as 10 ppm and a proton-carbon broadening as 25 kHz (full width at half height); then to achieve a residual width of 25 Hz at 15 MHz requires an rf field of 75 kHz. For the same residual broadening (in Hz) higher static fields mandate higher rf fields.

Even for on resonance irradiation, the coupling does not vanish identically. From a perturbation calculation, Haeberlen (18) finds the coupling is reduced by the factor

$$\Delta \langle \omega_{\text{CH}}^2 \rangle^{1/2} = \frac{1}{24} \frac{\langle \omega_{\text{CH}}^2 \rangle}{\omega_{1\text{H}}^2} \langle \omega_{\text{CH}}^2 \rangle^{1/2} \quad (8a)$$

$$\approx \frac{1}{134} \left[\frac{\Delta\omega}{\omega_{1\text{H}}} \right]^2 \langle \omega_{\text{CH}}^2 \rangle^{1/2} \quad (8b)$$

where Eq. (8b) is obtained by assuming the line is a Gaussian of full width $\Delta\omega$. To reduce a 25 kHz linewidth to 25 Hz requires then an rf field of 68 kHz; this correction term is independent of the size of the static field. The assumption of a Gaussian lineshape is for illustration only; the mutual proton-proton spin flips will serve to partly decouple the $I_z S_z$ interaction and make the lineshape more Lorentzian (19).

The forgoing examples indicate only two aspects of the rather complex problem of decoupling. The conclusion is that proton fields of around 75 kHz (17.5 G) are required to reduce a 25 kHz dipolar coupling by a factor of one thousand.

CROSS-POLARIZATION

Cross-polarization is based on the notion that the vast proton spin system can be tapped to provide some carbon polarization more conveniently than by thermalization with the lattice (7). Advantages are two-fold: the carbon signal (from those ^{13}C nuclei which are indeed in contact with protons) is enhanced and, more importantly, the experiment can be repeated at a rate determined by the proton longitudinal relaxation time T_{1H} , rather than by the carbon T_{1C} (7). There are many variants (7) of cross-polarization and only two common ones are described below (12,20).

In *spin lock* (SL) cross-polarization the protons are first spin locked in a resonant rf field of strength B_{1H} . Then an rf field is applied near the carbon resonance with an amplitude selected so that

$$\gamma_C B_{1C} = \gamma_H B_{1H}. \quad (9)$$

These two fields create carbon and proton reservoirs corresponding to ordering along the respective rf fields. When the above prescription is filled (the Hartmann-Hahn condition (5)), mutual carbon-proton spin flips become energy conserving and the two reservoirs are tightly coupled with a time constant comparable to the proton-carbon T_2 . When Eq. (9) is well met over the entire sample and for sufficiently large rf fields, the carbon signal is enhanced by a factor of four (γ_H/γ_C) compared to the carbon magnetization which would arise by thermalization with the lattice.

A second variation (21) initially prepares the system in a state of dipolar order; this ordering is used to cross-polarize the ^{13}C spins. We prefer to call this *dipolar* cross-relaxation although it is also known as cross-polarization under ADRF conditions, though *adiabatic demagnetization in the rotating frame* is only one way to create dipolar order. Here the cross-polarization rate is diminished from the spin lock rate and is extremely sensitive to the proton local field. This cross-polarization rate also gives a measure of spin fluctuations which are effective at relaxing the spin lock state and hence indicates how much the measured (carbon) $T_{1\rho}^c$ arises from spin-spin rather than spin-lattice processes. This observation will be discussed further in section V.

With this background we can now illustrate and refine these notions with some experimental results for cured epoxy resins.

SPECTRAL RESOLUTION, SPECTRAL FIDELITY AND MOLECULAR MOTION IN CURED EPOXIES

Figure 1 presents the three stages of resolution of ^{13}C NMR for a model organic solid, the epoxy resin diglycidyl ether of bisphenol A (DGEBA) cured with piperidine. In the top spectrum, for which only a portion is shown, neither magic angle spinning nor high power decoupling were employed; the result is representative for a conventional liquid state spectrometer. In the middle figure high power decoupling is added (proton rf field of 60 kHz). Finally in the bottom trace magic angle spinning at 2.2 kHz aids the decoupling. All the chemical shift anisotropy information, previously distributed throughout the line, has now piled up at the isotropic average; not only is interpretation easier but also the signal-to-noise ratio is increased as the lines have sharpened.

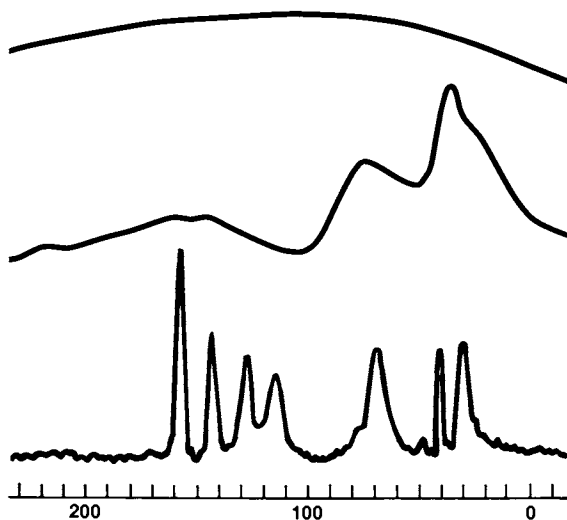


Figure 1. Three stages of resolution in a C-13 spectrum of a cured epoxy. The top spectrum is obtained under conditions appropriate to a liquid-state spectrometer: no dipolar decoupling and no magic angle spinning. Dipolar decoupling at 60 kHz is used for the middle spectrum and to that is added magic angle rotation at 2.2 kHz for the bottom figure.

Figure 2 indicates the potential for chemical identification of cured epoxy polymers; shown are the ^{13}C spectra of DGEBA based epoxies reacted with four different curing agents. Commercial DGEBA (DOW DER 332) and commercial curing agents were used. Proportions and curing cycles are in Table 1. An earlier version of these spectra appears in Ref. 4.

Table 1. Composition and Cure Cycle
for DGEBA Based Epoxies.

(a) Piperidine (PIP) 5%	120°C (16 hr)	
(b) Metaphenylene diamine (MPDA) 13.7%	20° (15 hr)	
	60° (1 hr)	
	149° (3 hr)	
(c) Hexahydrophthalic anhydride (HHPA) 31.1%	50° (16 hr)	
	90° (2 hr)	
	N, N-dimethylbenzylamine (DMBA) 0.2%	120° (2 hr)
(d) Nadic methyl anhydride (NMA) 46.2%	107° (2 hr)	
	N, N-dimethylbenzylamine	135° (2 hr)
	(DMBA) 0.8%	166° (2 hr)

Superposed in Figure 2 are liquid state spectra (with Overhauser enhancement) and peak assignments of the *unreacted* components, in a solvent. Solvent peaks are denoted with a slash (/).

In the sample of Figure 2a the curing agent piperidine was used and the epoxy approximates polymerized DGEBA. In the solid state spectrum, peaks *f* and *g* of the epoxide group are not apparent, giving a crude indicator of the degree of polymerization. A previous study (22) of partially cured piperidine-DGEBA observed lines near the methylene peak (*e*) which increased in strength and complexity as the cure progressed. These peaks were attributed to the carboxyl-methine ether carbon and to the methylene carbon near the reaction site. The expanded aliphatic region in Fig. 2a shows some reaction has occurred, even in the acetone solvent. (The insert spectrum was taken later than the full liquid state spectrum.) Hence in the solid all functional groups can be identified, except for the composite peak at *e*.

The contributions of the curing agents are easily seen in spectra 2b-d. Though there are unresolved peaks there is sufficient detail for chemical identification.

Two technical questions naturally arise: (i) are all the carbons counted and (ii) what limits resolution? In liquids all carbons are represented provided that the repetition period is substantially longer than the longest carbon T_{1C} . Resolution is generally restricted by static field inhomogeneity or by lifetime broadening. Circumstances in solids are less clearcut.

In the solid state experiments the protons are used to cross-polarize the carbon nuclei. Sufficient time must be allowed for the protons to thermalize with the lattice in between experiments but, unless the sample is inhomogeneous, all protons will share a

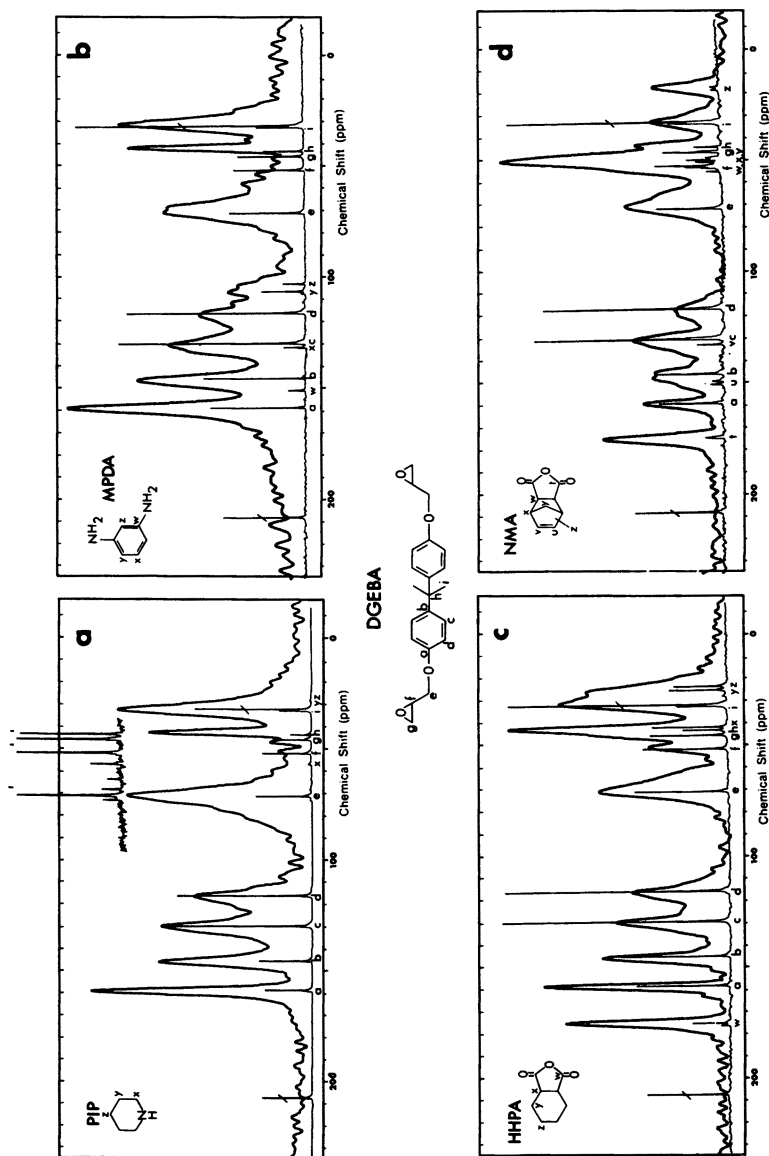


Figure 2. Solid-state spectra of four different epoxies (based on the resin diglycidyl ether of bisphenol-A) are compared with the liquid-state spectra of their respective unreacted components. The chemical compositions are in Table I. Here the epoxies are identified by their main curing agent: (a) PIP—piperidine; (b) MPDA—metaphenylenediamine; (c) HHPA—hexahydrophthalic anhydride; (d) NMA—nadic methyl anhydride.

common T_{1H} and so the carbon spectrum will be undistorted, albeit slightly attenuated for repetition periods shorter than T_{1H} .

Let us look more closely at the cross-polarization process. It is most convenient to regard the spin locked protons and carbons as reservoirs which equilibrate by a simple first order process. This simplistic view can break down. If the protons are isolated from one another, then polarization transfer is not between two monolithic reservoirs which, by assumption, quickly reach internal equilibrium, but rather between isolated carbon-proton spin pairs. The system then oscillates coherently at a frequency determined by the CH dipolar coupling constant (5,20,23). This is the circumstance just as the two reservoirs are brought into contact; it takes approximately the proton-proton T_2 for spin-pairs to respond to peer pressure and reach internal equilibrium. The oscillations are most pronounced when the Hartmann-Hahn condition is severely mismatched, by an amount greater than the proton local field; the resultant of the mismatch and the dipolar spin coupling determines the frequency. In the limit that the carbons are polarized from the dipolar state, the amplitude of this oscillation is approximately $M_o < \omega_{CH}^2 > / \omega_{1C}^2$ where M_o is the equilibrium carbon magnetization when the two reservoirs have reached thermal equilibrium, $< \omega_{CH}^2 >$ is the proton-carbon second moment, and ω_{1C} is the carbon rf field (20,21). (One difference between this and the oscillatory cross-polarization in liquids (5,24) is that the spatial dependence of the dipolar coupling renders "inequivalent" the protons in CH_2 or CH_3 groups, whereas in liquids the rotating frame cross-polarization retains the character of AX_2 , AX_3 coupling (24).)

If the protons are well coupled to one another, then after a few proton-proton T_2 's the simple thermodynamic picture is adequate. The dynamics of equilibration are particularly pure when the protons can be considered an infinite reservoir; then the ^{13}C magnetization grows as (25)

$$M(t) = M_o \lambda^{-1} \{1 - \exp(-\lambda t / T_{CH})\} \exp - t / T_{1\rho}^H \quad (10a)$$

where

$$\lambda \equiv \left[1 + \frac{T_{CH}}{T_{1\rho}^C} - \frac{T_{CH}}{T_{1\rho}^H} \right] \quad (10b)$$

and where $T_{1\rho}^C$, $T_{1\rho}^H$ are the carbon and proton rotating frame relaxation times and T_{CH} is the cross-polarization time under spin lock conditions. The equilibrium carbon magnetization M_o is determined by the proton spin temperature. For large rf fields, $M_o = \frac{\gamma_H}{\gamma_C} M_o^C$ where M_o^C is the ordinary carbon thermal magnetization appropriate to the static field. (Here $T_{1\rho}^C$ should be defined somewhat differently: as both carbon and proton systems are irradiated, this $T_{1\rho}^C$ will be sensitive to fluctuations at $\omega_{1C} + \omega_{1H} = 2\omega_{1C}$, rather than at ω_{1C} as usual.) If the rf fields are not so large, the proton spin lock ordering is fractionally reduced (26-28) by $1/4 < \omega_{HH}^2 > / \omega_{1H}^2$, where $< \omega_{HH}^2 >$ is the proton second moment. This leads to a slight decline in M_o but is not serious as it will influence all ^{13}C spins uniformly and not warp the spectrum.

Equation (10) admits a simple interpretation: the carbon magnetization rises with the rate λT_{CH}^{-1} while being depleted at $(T_{1\rho}^H)^{-1}$. As T_{CH} is, for matched Hartmann-Hahn condition, of the order of the carbon-proton T_2 , then $T_{CH} \ll T_{1\rho}^H < T_{1\rho}^C$ unless one is near a $T_{1\rho}$ minimum or unless T_{CH} has been artificially increased. This behavior

is illustrated in Figure 3 for the piperidine cured epoxy. The magnetizations of the chemically distinct carbons rise up with different rates and then diminish in concert. From the decay of the carbon magnetization, $T_{1\rho}^H \approx 2.6$ ms, a value in agreement with direct observation of the proton system.

What is the influence of magic angle spinning on the cross-polarization process? In principle magic angle spinning reduces heteronuclear dipolar coupling. In order for this averaging process to be efficient, the local proton-carbon coupling must remain static for, say, one-half revolution or $(2f_{rot})^{-1}$ where f_{rot} is the spinning frequency. Locally the coupling jumps around because of spin fluctuations in the proton spin system, fluctuations with a correlation time of the order of $3/\langle\omega_{HH}^2\rangle^{1/2}$. Only in special circumstances (29) is $2\pi f_{rot} > \langle\omega_{HH}^2\rangle^{1/2}$; for most organic solids enormous spinning rates (≥ 10 kHz) would be required to break completely the cross-polarization link. At more modest rates there is some effect; Figure 4 shows the cross-polarization times for protonated and non-protonated carbons in the piperidine cured epoxy for spinning speeds of 1-3 kHz. The relaxation times of the protonated aromatic, the methylene-methine and the methyl carbon increase marginally if at all. This follows from the observation that a reduction in heteronuclear line narrowing will show up only on the time scale of $(2f_{rot})^{-1}$, here 500 to 166 μ s. But the cross-relaxation proceeds so rapidly ($T_{CH} \approx 100$ μ s for the protonated aromatics), that the cross-polarization is virtually completed by the time the spinner has rotated very far. For the unprotonated aromatic and quaternary carbons relaxation times increase by about 50% on speeding up from 1 to 3 kHz. Even so, the longest cross-polarization time (of around 500 μ s at 3 kHz) is still substantially shorter than the proton $T_{1\rho}^H$ (2.6 ms). Further, at the rf field of 55 kHz, we find $T_{CH}/T_{1\rho}^C < 0.015$ for all carbons in the epoxy and so that correction in Eq. (10) can be ignored. The large gap between the T_{CH} 's and $T_{1\rho}^H$ insures that the ^{13}C spectrum gives a reasonable relative indication of the carbon intensities, for contact times greater than a few T_{CH} 's. For more careful work, the intensities should be corrected by Eq. (10). As T_{CH} approaches $T_{1\rho}^H$, the corrections become larger and, more importantly, the maximum available carbon signal is diminished by the rapid decay of the proton rotating frame magnetization.

So, for moderate spinning rates and for spin lock cross-polarization in rf fields well away from any $T_{1\rho}$ minimum, the cross-polarization spectrum counts all the ^{13}C nuclei in contact with the proton bath. In this homily we presume that the heteronuclear coupling is not abridged by rapid, nearly isotropic motion as in a lightly cross-linked polymer above its glass transition temperature. Some estimate of the efficiency of cross-polarization can be gleaned by recognizing (5) that the Hartmann-Hahn condition transforms the heteronuclear dipolar coupling which was static in the laboratory frame into one which is static in the doubly rotating frame. That is, if a ^{13}C resonance line (without dipolar decoupling) is not substantially broadened, then cross-polarization will not proceed. One says then that cross-polarization discriminates against liquid-like lines or indeed any carbon line not broadened by proton coupling. The discrimination of the cross-polarization process has been used to count the organic carbon rather than total carbon content of oil shale (30) and to distinguish between mobile and polymeric phases in hemoglobin (31).

Epoxyes are good candidates for solid state ^{13}C studies because of their relative chemical simplicity but even so some spectral lines overlap, as was shown in Fig. 2. We enquire into the limits of resolution to see what improvements can be expected.

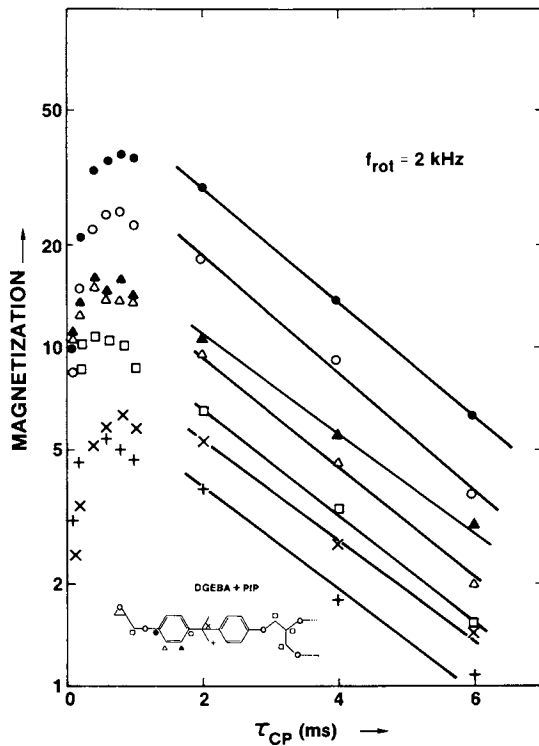


Figure 3. Cross polarization magnetization for the PIP-cured epoxy under the SL (Hartmann-Hahn) condition. The cross polarization contact time is τ_{CP} . The decay corresponds to proton $T_{1\rho}$ relaxation.

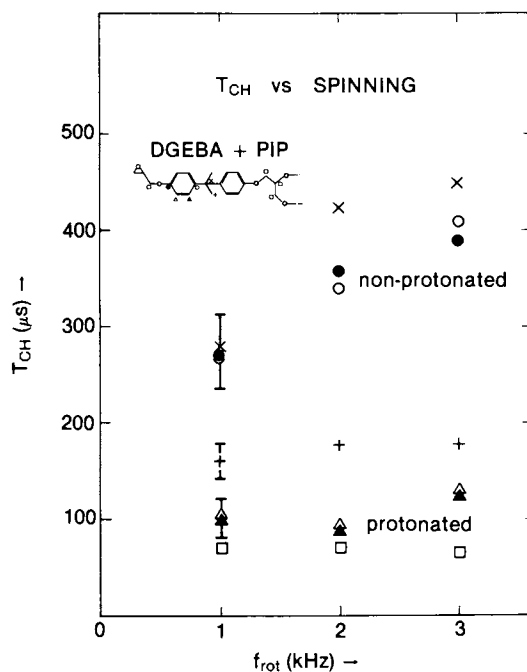


Figure 4. The influence of magic angle spinning on the SL cross polarization time constant. In the PIP-cured epoxy the time constant for the nonprotonated carbons increases by about 50% for $f_{rot} = 1-3$ kHz while those of the protonated carbons are virtually unchanged over this range.

Both inhomogeneous and homogeneous line broadening can occur; in some sense the former can be refocused by 180° rf pulses while the latter cannot. In the epoxies two prime candidates for line broadening are (i) a distribution of isotropic chemical shifts (inhomogeneous) and (ii) inadequate proton decoupling (homogeneous). Substantial fluctuation of the heteronuclear dipolar coupling, Eq. (2), from molecular motion or spin flipping at the decoupling frequency reduces decoupling efficiency.

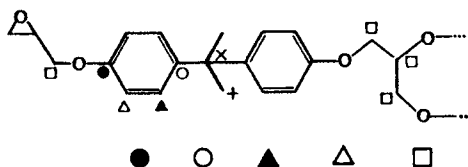
To distinguish between these two mechanisms, create a ^{13}C spin echo while simultaneously decoupling, as in Fig. 5. Naively one expects any chemical shifts to refocus at 2τ and hence Fourier transformation of the second half of the echo allows a value of T_2 (decoupled) to be determined for each line. Any diminution of the echo will then correspond to *homogeneous* broadening. (As proposed this experiment is complicated by the magic angle spinning at f_{rot} ; the chemical shift anisotropy is reintroduced (3) to the echo when $(4\tau)^{-1} \approx f_{rot}$, $2f_{rot}$.) Alternatively, the carbon $T_{1\rho}^C$ can provide similar information; the relevant pulse sequence is also shown in Fig. 5. (As we shall no longer discuss proton relaxation, we drop the superscript and let $T_{1\rho}$ refer to carbon relaxation.)

The heteronuclear dipolar coupling between spin species I and S is given in Eq. (2) and rewritten below in different form:

$$H_D(t) = \sum_{j,k} I_z(t) S_{zk}(t) f(\mathbf{r}_{jk}(t)), \quad (11)$$

where $I_z(t)$, $S_z(t)$ are spin operators which may be explicitly time dependent and $f(\mathbf{r}(t))$ is a geometrical factor with argument \mathbf{r} defining internuclear orientation which is implicitly time dependent. The relaxation rate in a carbon T_2 or $T_{1\rho}$ experiment involves the Fourier transform of the auto-correlation function of $H_D(t)$ (32). Figure 6 contrasts the two experiments. The respective rf fields impress coherent motion on the carbon spin operators S_z in the $T_{1\rho}$ experiment and on the proton spin operators I_z in the (decoupled) T_2 experiment. Molecular motions are random and so contributions to the spectral density function (the integral in the figure) will obtain for those fluctuations in $f(\mathbf{r}(t))$ at $(-\omega_{1S})$ and $(-\omega_{1I})$ respectively. When the Hartmann-Hahn condition is matched, then $T_{1\rho} = T_2$ (decoupled) in the instance that molecular motion determines the relaxation rates. (When spin-spin rather than spin-lattice effects determine the relaxation rates, then $T_{1\rho} \neq T_2$ (28).)

We then compare the measured linewidth to that implied by the "lifetime broadening" of $T_{1\rho}$ at an rf field of 66 kHz and static field of 15 MHz for the piperidine cured DGEBA epoxy at 33°C .



$T_{1\rho}$ (ms)	60	55	10.5	9.5	14	35	26
$(\pi T_{1\rho})^{-1}$ (Hz)	5.3	5.8	30	34	22	9	12
linewidth (Hz)	49	61	86	98	122	40	86

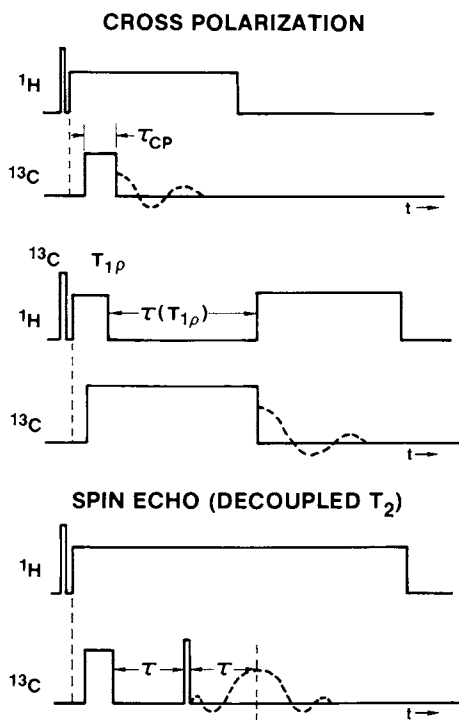


Figure 5. The rf pulse sequences for determining SL cross polarization time constant T_{CH} , C-13 $T_{1\rho}$ and C-13 T_2 under proton decoupling. Each experiment starts with a SL cross polarization.

At this rf field only a small fraction of the apparent linewidth can be attributed to broadening by $T_{1\rho}$ mechanism. At lower rf fields the contribution to the protonated aromatic carbon will become more important.

The source of this excess broadening must arise from chemical shift-like (inhomogeneous) terms. Magnet inhomogeneity is about 6 Hz and a missetting of the magic angle could contribute to lines with a large chemical shift anisotropy. There may also be a distribution of isotropic chemical shifts (2); evidence for this is seen on comparing the spectra for the piperidine cured epoxy at -36 and $+58^\circ\text{C}$ (Fig. 7). At the lower temperature there is a well-defined splitting of about 5.5 ppm of the resonance from the protonated aromatic carbon nearest the ether linkage. The splitting likely arises from the three bond removed methylene group according to the proximity of the ortho carbon and the methylene group. In polycrystalline 1,4 dimethoxybenzene a similar splitting of the ortho carbons is observed (33), imposed by the alignment of the methyl group. In the epoxy at the higher temperature large amplitude rotation of the aromatic group averages out this splitting. Indeed in the epoxy the methyl resonance narrows by a factor of two on going from -36 to $+58^\circ\text{C}$, also suggesting its chemical shift distribution is averaged by the motion of the aromatic group.

This effect — the broadening of resonance lines in highly cross-linked polymers on the order of ppm — is certainly not restricted to the epoxies. In a liquid, molecular tumbling averages out chemical shift anisotropy. But furthermore *intra* molecular motions may average *intra* molecular contributions to the isotropic chemical shift: these are not averaged to zero but rather the higher symmetry of the average chemical environment can lead to a simpler spectrum. One might call this a *conformational anisotropy*, which is unaffected by magic angle spinning. If these conformations cannot rapidly interconvert, or if they are interconverting but with a distribution of motional correlation times, then one should expect rather diffuse lines. In this event then, at a given temperature, spectral resolution will be determined primarily by the inherent chemical shift distribution and cannot be improved by spectroscopic techniques. This appears to be the case for the model epoxy at an rf field of 66 kHz.

If highest resolution is required, then the strategy is to use thermal activation to stir away the conformational anisotropy: operate as close to the glass transition temperature as possible, up to the point at which $T_{1\rho}$ lifetime broadening predominates. On the other hand, at lower temperatures the lineshape and its temperature dependence may provide useful information (34).

SPIN-LATTICE RELAXATION IN THE ROTATING FRAME

Spin-lattice relaxation of ^{13}C nuclei is, in principle, very attractive for it is determined by local fluctuations at ω_{1C} or ω_{0C} (rotating or lab frame respectively): spin diffusion among ^{13}C nuclei does not average relaxation rates among chemically distinct carbons. In solids one must append a cautionary note.

Spin-spin fluctuations can compete with spin-lattice effects: an energy $h\omega_{1C}$ can be supplied by a phonon as well as by a spin fluctuation in the dipolar field. A simple thermodynamic view is shown in Fig. 8. For convenience only two distinct carbon species are shown, protonated (primed) and unprotonated (unprimed). During the $T_{1\rho}$

$$H_D(t) = \sum I_{Zi} S_{Zk} f(r_{jk}(t))$$

$$\int_{-\infty}^{\infty} d\tau \langle H_D^*(t + \tau) H_D(t) \rangle e^{-i\omega\tau}$$



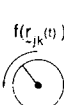



I_{Zi}	S_{Zj}	$f(r_{jk}(t))$	$J(\omega_{1S})$
			
			$J(\omega_{1H})$

Figure 6. The proton(I)–carbon(S) dipolar coupling during a C-13 $T_{1\rho}$ and decoupled T_2 experiment are compared. The relaxation rate is determined by the molecular fluctuation at the spin lock frequency ω_{1C} or decoupling frequency ω_{1H} .

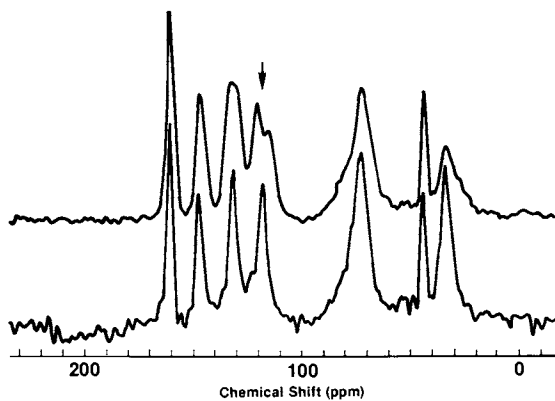


Figure 7. Spectra of the PIP-cured epoxy at -36°C (top) and 58°C (bottom). The splitting of the peaks denoted by the arrow is 5.5 ppm.

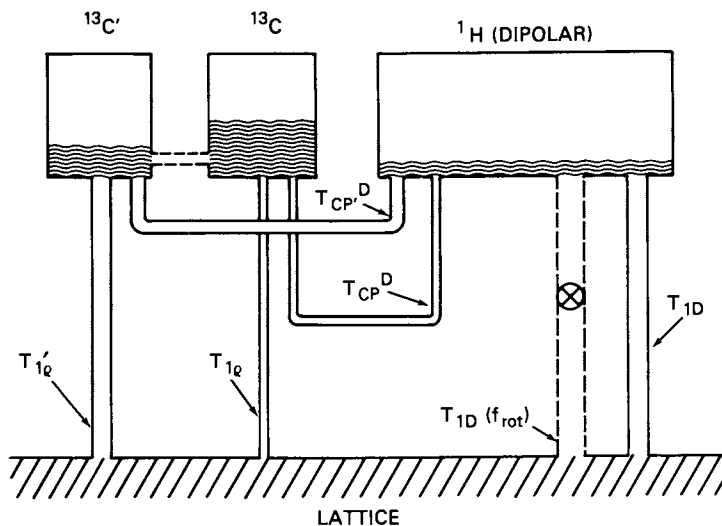


Figure 8. During the C-13 $T_{1\rho}$ experiment, the protonated (primed) and unprotonated (unprimed) carbons are in contact with not only the lattice but also the proton dipolar reservoir. Here $T_{1D} (f_{\text{rot}})$ indicates a dipolar spin-lattice process which depends on spinning speed and orientation.

experiment (ω_{1C} on, ω_{1H} off), the two carbon reservoirs are in thermal contact with not only the lattice by $T_{1\rho}$ and $T_{1\rho}$ but also with the proton dipolar reservoir with T_{CP}^D and T_{CP}^D . The later two are cross-polarization times appropriate to magnetization transfer from the (proton) dipolar state to the carbon spin lock state (or vice versa). This pathway was first mentioned in Section III. The dipolar state is thermally linked to the lattice by spin-lattice process T_{1D} and an additional process $T_{1D}(f_{rot})$ which depends on the rotational speed and orientation of the spinner (27,35,36). For an organic solid spun at around 2 kHz at the magic angle, $T_{1D}(f_{rot})$ is of the order of 100 μ s (28). The dipolar reservoir is thus strongly coupled to the lattice. For modest rf fields and for naturally abundant ^{13}C , the specific heat of the dipolar reservoir outweighs that of any ^{13}C spin lock reservoir. In this limit the effective rotating frame relaxation time $(T_{1\rho})_{eff}$ becomes

$$(T_{1\rho}^{-1})_{eff} = (T_{1\rho})^{-1} + (T_{CP}^D)^{-1}. \quad (12)$$

The cross-polarization (spin-spin) rate $(T_{CP}^D)^{-1}$ competes on an equal footing with the spin-lattice (motional) rate $(T_{1\rho})^{-1}$. Now the cross-polarization rate is strongly field dependent (12,20,21,37):

$$(T_{CP}^D)^{-1} = \frac{\pi}{2} \langle \omega_{CH}^2 \rangle \tau \exp - \omega_{1C} \tau, \quad (13)$$

where $\langle \omega_{CH}^2 \rangle$ is the proton-carbon second moment and the correlation time is determined by the proton-proton coupling (37),

$$\tau^{-2} = \frac{1}{9} \langle \omega_{HH}^2 \rangle K. \quad (14)$$

Here $\langle \omega_{HH}^2 \rangle$ is the proton-proton second moment and K is a geometrical factor varying from 0.5 to 1 in crystals of cubic symmetry (37).

Rewriting Eqs. (13), (14) in terms of the proton local field $\omega_L^2 \equiv 1/3 \langle \omega_{HH}^2 \rangle$,

$$T_{CP}^D = T_2' \exp \sqrt{\frac{3}{K}} \frac{\omega_{1C}}{\omega_L}, \quad (15a)$$

with

$$T_2' \equiv \frac{2}{\pi} \sqrt{\frac{K}{3}} \frac{\omega_L}{\langle \omega_{CH}^2 \rangle}. \quad (15b)$$

The exponential dependence holds out the prospect of finding a sufficiently large carbon rf field ω_{1C} to make T_{CP}^D much longer than $T_{1\rho}$, which would be expected to have a ω_{1C}^2 (38) or weaker dependence. Further away from the $T_{1\rho}$ minimum progressively larger rf fields are required to ensure $T_{CP}^D \gg T_{1\rho}$.

Only one complication to the determination of carbon $T_{1\rho}^C$ has been identified but it illustrates the role of the strongly interacting proton dipolar system, a role which must be examined in even more detail for the non-spinning case (39).

CONCLUSIONS

The methods of high power decoupling, cross-polarization and magic angle spinning produce useful spectra from intractable polymers. In a rather rigid material

increasing the decoupling power improves spectral resolution up to the limit imposed by the distribution of chemical shifts, which in a polymeric system below the glass transition temperature can be of the order of 5 ppm. At higher temperatures rapid interconversion of the conformations may reduce linewidths to a residual imposed by lifetime broadening due to the carbon $T_{1\rho}$.

In organic solids the determination of rotating frame relaxation is severely complicated by the presence of the strongly interacting proton spin system. Spin-spin fluctuations compete with spin-lattice fluctuations to produce an effective relaxation time; large rf field amplitudes are mandated to discriminate against the spin-spin event. The burden of proof lies with the experimenter to establish that a rotating frame relaxation rate actually reflects a motional effect seen by the carbon nuclei.

Acknowledgements

Discussions with D. L. VanderHart have helped to identify the role of dipolar order in carbon rotating frame studies. This work is sponsored in part by the Naval Air Systems Command.

Abstract

Combining magic angle spinning with proton enhanced ^{13}C NMR, one extracts from organic solids not only ^{13}C spectra, but also certain NMR relaxation rates. The spectra are useful for coarse chemical identification of intractable polymers although spectral resolution is limited to a few ppm in amorphous polymers; the origin of this restriction is discussed. Under certain conditions the ^{13}C rotating frame relaxation rate $T_{1\rho}^{-1}$ as well as the spectra can reflect the nature of the molecular motions monitored at each resolvable ^{13}C functional group. These points are illustrated in a piperidine cured epoxy based on diglycidyl ether of bisphenol A (DGEBA).

LITERATURE CITED

- (1). J. Schaefer, E. O. Stejskal, *J. Am. Chem. Soc.* **98**, 1031 (1976).
- (2). J. Schaefer, E. O. Stejskal and R. Buchdahl, *Macromolecules* **10**, 384 (1977).
- (3). E. Lippmaa, M. Alla and T. Tuherm, *Proc. XIX Congress Ampere, Heidelberg* 113 (1976).
- (4). A. N. Garroway, W. B. Moniz, and H. A. Resing, *Preprints of the Div. of Organic Coatings and Plastics Chem.* **36**, 133 (1976).
- (5). S. R. Hartmann and E. L. Hahn, *Phys. Rev.* **128**, 2042 (1962).
- (6). (a) F. Bloch, *Phys. Rev.* **111**, 841 (1958), (b) L. R. Sarles and R. M. Cotts, *Phys. Rev.* **111**, 853 (1958).
- (7). A. Pines, M. G. Gibby and J. S. Waugh, *J. Chem. Phys.* **59**, 569 (1973).

- (8). E. R. Andrew, *Arch. Sci. (Geneva)* **12**, 103 (1959).
- (9). E. R. Andrew, *Prog. in NMR Spectroscopy* **8**, 1 (1971).
- (10). I. J. Lowe, *Phys. Rev. Lett.* **2**, 285 (1959).
- (11). H. Kessemeier and R. E. Norberg, *Phys. Rev.* **155**, 321 (1967).
- (12). M. Mehring, "High Resolution NMR Spectroscopy in Solids," *NMR: Basic Principles and Progress* **11** (1976).
- (13). U. Haeberlen, "High Resolution NMR in Solids: Selective Averaging," *Adv. in Magn. Resonance supplement* **1** (1976).
- (14). H. Y. Carr and E. M. Purcell, *Phys. Rev.* **94**, 630 (1954).
- (15). E. R. Andrew, W. S. Hinshaw and R. S. Tiffen, *J. Magn. Resonance* **15**, 191 (1974).
- (16). (a) M. M. Maricq and J. S. Waugh, *Chem Phys. Lett.* **47**, 327 (1977). (b) J. S. Waugh, M. M. Maricq and R. Cantor, *J. Magn. Resonance* **29**, 183 (1978).
- (17). J. W. Beams, *Rev. Sci. Instrum.* **1**, 667 (1930).
- (18). Ref. 13, p. 82.
- (19). (a) Ref. 12, chapt. 4.4, (b) M. Mehring, G. Sinnig and A. Pines, *Z. Physik B* **24**, 73 (1976).
- (20). D. E. Demco, J. Tegenfeldt and J. S. Waugh, *Phys. Rev. B* **11**, 4133 (1975).
- (21). D. A. McArthur, E. L. Hahn and R. Walstedt, *Phys. Rev.* **188**, 609 (1969).
- (22). S. A. Sojka and W. B. Moniz, *J. Appl. Polym. Sci.* **20**, 1977 (1976).
- (23). L. Muller, A. Kumar, T. Baumann, and R. E. Ernst, *Phys. Rev. Lett.* **32**, 1402 (1974).
- (24). R. D. Bertrand, W. B. Moniz, A. N. Garroway and G. C. Chingas, *J. Am. Chem. Soc.* (in press).
- (25). Ref. 12, chapt. 4 discusses cross-polarization under a number of conditions; here we have included the effects of carbon $T_{1\rho}^C$ during cross-polarization.
- (26). P. Mansfield and D. Ware, *Phys. Rev.* **168**, 318 (1968).
- (27). A. N. Garroway, submitted to *J. Magn. Resonance*.
- (28). A. N. Garroway, W. B. Moniz and H. A. Resing, to be published in *Faraday Soc. Symposium* **13**.

- (29). E. O. Stejskal, J. Schaefer and J. S. Waugh, *J. Magn. Resonance* **28**, 105 (1977).
- (30). H. A. Resing, A. N. Garroway, and R. N. Hazlett, *Fuel*, in press (1978).
- (31). D. A. Torchia and D. L. VanderHart, *Topics in ^{13}C NMR* **3**, in press (1979).
- (32). A. Abragam, *The Principles of Nuclear Magnetism*, Clarendon Press, Oxford (1961), chapt. 8.
- (33). E. T. Lippmaa, M. A. Alla, T. J. Pehk, G. Engelhardt, *J. Am. Chem. Soc.* **100**, 1929 (1978).
- (34). A. N. Garroway, W. B. Moniz and H. A. Resing, in preparation.
- (35). J. F. J. M. Pourquie and R. A. Wind, *Phys. Lett.* **55A**, 347 (1976).
- (36). J. Jeener, VI International Symposium on Magnetic Resonance, Banff Canada (1977) unpublished.
- (37). H. T. Stokes and D. C. Ailion, *Phys. Rev. B* **15**, 1271 (1977).
- (38). N. Bloembergen, E. M. Purcell and R. V. Pound, *Phys. Rev.* **73**, 679 (1948).
- (39). D. L. VanderHart and A. N. Garroway, submitted to *J. Magn. Resonance*.

Discussion

A. A. Jones, Clark University, Mass.: Could you summarize briefly those conditions under which rotating frame relaxation provides a good probe of molecular motion in the solid state? Is this something that can be achieved with a good spectrometer and a rf large rf field?

A. N. Garroway, Naval Research Laboratory, Wash., D. C.: I would like to give a simple prescription but I can't. There are a number of tests to establish that spin-lattice effects predominate: (i) direct measurement of spin-spin cross-relaxation time T_{CP}^D (cf Fig. 8) and (ii) the sensitivity of the observed relaxation rate to rf field strength. However the spin-spin contribution must always be compared to the motional contribution and so any simple prescription for the minimum rf field required to see molecular motion necessitates *a priori* knowledge of the extent of molecular motion. As I mentioned, even with a large rf field of 80 kHz, VanderHart (Ref. 39) has not found evidence of a spin-lattice component to the carbon rotating frame relaxation in drawn polyethylene at room temperature; there is simply not very much motion in that frequency regime. Now in a system closer to the $T_{1\rho}$ minimum, the rf requirements are far less stringent.

A. A. Jones: When the specimen is spun, it is mechanically perturbed at 2-3 kHz. This is a frequency not too different from that of the motion. Does the mechanical spinning excite the system and therefore affect the rotating frame relaxation, assuming that one does everything else correctly?

A. N. Garroway: Probably not. There are two aspects. Naively what happens is that the 2 kHz spinning puts sidebands on the molecular motion. Motions which were at 60 kHz will then appear at 58 and 62 kHz. This is not of any importance provided the rf field (γB_1) and spinning frequency are well separated. In a system in which the dipolar strengths are of the order of only a few kHz, the effect of magic angle spinning can be seen on cross-polarization, which is a spin-spin process. A very interesting paper by Stejskal, Schaefer and Waugh (Ref. 29) addressed just this issue in adamantane. The second aspect is connected with centrifugal forces. Spinning at 2 kHz does not sound ominous yet we are talking about a force of 100,000 G's on the circumference of a 1 cm diameter sample. There is some evidence at that slight changes in chemical shifts can occur. However, a few tenths of a kilobar is not really the sort of pressure one expects to change the molecular orbitals substantially. The story might well be different in an elastomer.

S. Borwnstein, NRC, Ont.: Can one use the same approach to rate processes for the motion which changes the spectra in the solid state as one normally uses in solution? In Fig. 7 a doublet collapses into a singlet with temperature. This can be very easily analyzed in the liquid state to give information on the rate of a particular motion that is causing the averaging.

A. N. Garroway: Yes, however it is unlikely that a single rate is responsible.

C. J. Carmen, B. F. Goodrich, Ohio: Would you care to make any comment on the limitations of measuring carbon $T_{1\rho}$ because of the spin-spin effects? Do you feel that the practical goal understanding mechanical properties of polymer mixtures is going to be somewhat dubious as far as using carbon $T_{1\rho}$ in a real, dirty system?

A. N. Garroway: I would still hold out the hope for using these carbon relaxation rates to interpret mechanical properties, but the onus is on the experimenter to show he is actually measuring the effects of motion. Once that is done I think the idea will compete as freely as any other scientific concept.

RECEIVED March 13, 1979.

Carbon-13 NMR Studies of Model Anionic Polymerization Systems

S. BYWATER and D. J. WORSFOLD

Division of Chemistry, National Research Council of Canada,
Ottawa, Canada K1A 0R9

The marked variation in stereostructure of diene polymers caused by changes in the counter-ion and solvent when butadiene or isoprene are polymerized anionically, are as yet not fully explained. Much progress has been made on elucidating the causes of variations in the cis/trans ratio of the 1:4 structures in these systems (1, 2, 3), but the causes of the change in the ratio of 1:2 to 1:4 structures in butadiene for example has been left largely unresolved. In dioxane, for instance, the amount of 1:2 structure decreased from 87% with Li counter-ion at 15°, to 41% with Cs (4). Less variation is found in THF because a substantial part of the reaction is carried by the free ion. Changes are also observed in polyisoprene (5).

It is likely that these changes in microstructure are a result of changes in the active chain end with the various counterions. More specifically the decrease in the 1:2 to 1:4 ratio of polybutadiene when the series is spanned from Li to Cs might be caused by different types of bonding or changes in the charge distribution in the growing allylic carbanion. Information on bonding changes in this type of system may be sought from C-C coupling constants (6), and from the effect of deuterium substitution on the C-13 spectra. Changes of charge distribution are obtainable from variations in the C-13 chemical shifts which are sensitive to charge (7).

Hence measurements have been made on unsubstituted allyl alkali-metal compounds, and also on neopentylallyl (I, 5,5-dimethylhexen-2-) and neopentylmethallyl (II, 2,5,5-trimethylhexen-2-) alkali-metal compounds which are models of the polymerizing chain end in the anionic polymerization of butadiene and isoprene respectively.

Experimental

All the allylic compounds were prepared by reacting the appropriate mercury allyl compounds with either a film of the alkali metal, or finely divided lithium, in an evacuated apparatus fitted with breakseals, a glass filter, and a nmr tube. The solvent was THF except where noted.

0-8412-0505-1/79/47-103-089\$05.00/0
Published 1979 American Chemical Society

Allylmercury was prepared substituted in one end only of the allyl radical with either C-13 or D₂. The starting materials were the appropriately labelled paraformaldehydes, purchased from Merck, Sharp and Dohme. First the formaldehyde was sublimed into vinylmagnesium bromide, in THF, at -10°C to give, after work up, labelled allylalcohol. The allylalcohol was not isolated but retained as a THF solution which, on distilling from ZnCl₂ and conc. HCl, gave a THF solution of allylchloride. At this point the labelling was scrambled between the two end positions. After drying on CaH₂, the allylchloride solution was used to prepare allylmagnesium chloride which on reacting with mercuric chloride gave diallylmercury. The product was distilled at low pressure, to give an overall yield of 14%.

The mercury compounds corresponding to I and II were prepared as described before (3).

All nmr measurements were made on a Varian XL100 instrument. Shifts are reported in ppm downfield of TMS.

Results

The allyl alkali-metal compounds give 2 line C-13 nmr spectra. The terminal positions are equivalent either because of rapid equilibrium between two covalent structures, or because the structure is a delocalized symmetrical ion. The chemical shifts and C-C coupling constants are recorded in table I. Reasonable agreement with literature δ values are found (8,9).

Deuteration of one end of the allyl moiety in these compounds removes the equivalence of the two positions and in place of the single line for the terminal position, two separate absorptions should appear (10). One, in the normal decoupled spectrum, is a singlet for the hydrogen substituted carbon, and the other a weak quintet for the deuterated end which would be difficult to observe. These two signals would bracket the normal singlet. If a mixture of deuterated and undeuterated allyl compound is used, therefore, two easily observable peaks should appear, one in the normal position, the other shifted. In the spectra of allyllithium and allylsodium the line from the deuterated compound appeared 14 and 11 Hz upfield respectively, at 0°C, of the normal lines. The potassium compound only showed a somewhat broadened line. At -80°C the separation for allyl-lithium was 22 Hz.

C-13 nmr spectra were taken of all the alkali metal compounds of I and II from Li to Cs. As expected, changing the counter-ion had very little effect on the chemical shifts of the carbons in the neopentyl group in either I or II, or on the extra methyl group in II, compared with the parent hydrocarbon (3). The substantial variation in the shifts of the allyl carbons are shown in table II. In both series the β carbon is moved 10-14 ppm downfield from its position for the parent hydrocarbon. The position of the γ peak moves markedly upfield from Li to K and then remains fairly constant, while the α carbon moves downfield over the whole series.

Table I.
Allyl M ^{13}C Chemical Shifts^a and Coupling Constants^b at 0°C

M	δC_1	δC_2	$J_{\text{C-C}}$
Li	50.8	146.0	55.9
K	52.4	143.3	59.8
Cs	60.0	143.6	61.1

- a. In THF solution, the shifts recorded are as ppm downfield of TMS, using THF $\delta = 26.21$ as standard
b. in hertz

Discussion

If the allyl alkali metal compounds are ionic compounds, then if the allyl ion is delocalized the terminal carbons would be equivalent and contain most of the charge, and all three carbons would be sp^2 hybridized.

^{13}C - ^{13}C nmr coupling constants have been shown to be primarily sensitive to the s character of the bonded carbons. In simple aliphatic compounds most $J_{\text{C-C}}$ values of sp^3 - sp^2 bonded carbons lie between 40 to 60 hz, whilst those between two sp^2 hybridized carbons are in general above 65 hz. Aromatic compounds, however, such as benzene and its derivatives seem to have intermediate values of 55-60 hz for the aromatic carbon coupling constants.

Studies on benzyl alkali metal compounds, models of styrene anionic polymerization systems, showed coupling constants to the enriched α carbon in the above ranges, showing it to be sp^2 hybridized (11). Little effect of the charge could be detected in the $J_{\text{C-C}}$ values, although it should be noted that the charge is extensively delocalized into the benzene ring.

In the present study, the $J_{\text{C-C}}$ values in the allyl alkali metal compounds are near 60 hz for the K and Cs compounds, and a little lower for the Li compound, table I. Allyl mercury is not a delocalized system, and if any exchange occurs between the two ends of the allyl radical it is slow on the NMR time scale. Consequently $J_{\alpha-\beta}$ and $J_{\beta-\gamma}$ are measurable and are 42 and 69 hz respectively. Thus taking the mercury compound as a model of a static covalent structure, rapidly equilibrating allylic compounds would be expected to give an averaged coupling constant near 55 hz which is close to the Li compound's value, and not very far from those of the K and Cs compounds. This predicted value is not sufficiently different from that observed in delocalized structures such as benzene to distinguish between these two

possible types of structure although higher coupling constants are observed in other olefinic cpds (~ 70 cps). But the evidence of West(12) on non-equivalence of the hydrogens at -80° in allyl-lithium and the small probability of allyl cesium being covalent suggest that for allylic sp^2 hybridized species coupling constants lower than 70 cps are to be expected. Even lower values were observed for compound II-Li in THF (46 hz) and in benzene (36 hz) (11). The latter value is even below the reported range for systems having the sp^3 - sp^2 hybridization expected of covalent compounds. Uncertainty as to the effect of association and charge on J_{C-C} make it unwise to speculate at this time on the meaning of these results.

It has been suggested that the effect of selective deuteration on C-13 spectra of symmetrical carbenium ions can be used to distinguish between rapidly equilibrating structures (ie. involving hydride and methide shifts) and delocalized structures (13).

Similar effects could be expected for carbanions. Normally k_H/k_D ratios in the region 1.15-1.20 are found per deuterium in the formation of carbenium ions. If this ratio is translated into the separation of C-13 shifts expected by deuteration of one side of an equilibrating ion, then $\delta/\Delta \sim 0.1$ per deuterium, where δ is the difference between the observed lines and Δ is the separation expected between the shifts of the two positions in each tautomeric structure (10). If, however, there is resonance between the two structures, this ratio appears to be considerable smaller (14).

This ratio calculated from the observations on the deuterated allyl alkalimetal compounds is 0.0044 for Li, 0.0034 and perhaps half this for K. This is sufficiently smaller than the 0.1 figure quoted above to give support to the delocalized form of all these allyl alkalimetal compounds including the lithium compound.

The upfield movement of the C-13 chemical shifts (table II) of compounds I and II, models of the polymerizing systems, indicates that the negative charge residing of the γ carbon increases from Li to K, and then remains fairly steady for Rb and Cs. The movement of the chemical shifts of the α carbon is in the opposite direction, which would indicate a lessening of charge at this position. As the movements are roughly comparable in magnitude this suggests a transfer of charge from the α to the γ position as the counter ion increases in size. Li^+ is small relative to the allylic system, but Cs^+ can overlap all three positions. Thus although the Li^+ ion could well localize the charge at a particular position (α), the Cs^+ ion could allow a charge distribution more nearly approaching that of the free anion.

Although all change of the γ carbon chemical shift from that of the parent hydrocarbon in compounds I and II is caused by charge, the overall change in the α position's shift is composed

Table II.
 ^{13}C Chemical Shifts^a of Allyl Carbons in I-M^b and II-M^b

Cpd.	Solvent	δ_{α}	δ_{β}	δ_{γ}
I-H	THF	12.8	125.3	127.8
I-Li	C_6D_6	20.0	140.3	103.0
I-Li	DEE	30.7	140.0	87.6
I-Li	THF	31.0	142.5	81.9
I-Na	THF	35.7	138.8	72.3
I-K	THF	45.0	137.5	67.5
I-Rb	THF	47.4	138.2	67.5
I-Cs	THF	51.4	139.5	69.0
II-H	C_6D_6	18.0	132.4	122.3
II-Li	C_6D_6	24.2	148.4	100.4
II-Li	DEE	33.3	149.2	87.6
II-Li	THF	31.8	149.3	83.9
II-Na	THF	36.7	146.1	79.2
II-K	THF	45.8	143.2	71.9
II-Rb	THF	49.0	143.1	69.0
II-Cs	THF	53.7	143.4	70.8

- a. Shifts are recorded in ppm downfield of TMS using the following solvent peaks as standards. $\text{C}_6\text{D}_6 = 128.0$; Diethylether (DEE) = 15.55, THF = 26.10. Temperature -20° except for parent hydrocarbons and C_6D_6 solutions which were measured at $+20^\circ$.
- b. Chemicals shifts are quoted only for the *cis* isomer. In many cases the *trans* isomer is not observed.

of two contributions. There is an upfield component due to negative charge, and a downfield movement caused if there is a change in hybridization from sp^3 in the parent compound to sp^2 in the ion. The magnitude of these two effects can be obtained by using allylpotassium as a model of a delocalized ionic compound and comparing its chemical shifts with those of propene. The upfield movement of the γ shift is 63 ppm, whilst the α shift is 34 ppm downfield. Hence in this symmetrical ion this 34 ppm must be composed of the 63 ppm upfield movement (as in the γ position) caused by charge, and an extra 97 ppm downfield movement caused by the hybridization change. The figure 97 ppm is a reasonable figure for such a $sp^3 \rightarrow sp^2$ change (δC_1 , in propane and propene is 15.6 and 115.0 respectively). As the β carbon has a change of chemical shift of 11 ppm downfield, the net upfield movement of chemical shifts caused by charge is 115 ppm. If this is equated to $1e$, then the upfield shift per electron of 115 ppm is substantially smaller than that found for aromatic systems. A similar calculation on allyllithium gives a figure of 114 ppm per electron.

Using these figures, it is then possible to calculate the charges on the alkalimetal compounds I and II, and these are shown in Table III. It is seen that the total charge approaches $1e$ as expected, at least for the higher alkalimetal compounds. The charge distribution does change markedly. There is a localization of the charge at the α position in the Li compounds, but for Cs there is almost equal distribution of charge between the two positions. Despite the evidence of low coupling constants, the Li compounds appear to be in reasonable agreement with this scheme even in benzene solution.

Nmr measurements in THF are characteristic of the ion-pairs (presumably contact). Microstructure measurements in this solvent are affected by the very small amount of very reactive free anions present in dilute solutions. These produce with isoprene (15) (and probably butadiene) very high vinyl contents in the polymer. The vinyl content in THF (16) does not drop for this reason as rapidly with increasing counter-ion size as in diethylether and dioxane (Table IV). Nevertheless it can be qualitatively assumed (as did Essel (5)) that each ion-pair produces a characteristic microstructure largely independent of solvent. Such a generalization would be expected to be least valid for lithium where changes in external solvation could produce some differences. It appears generally therefore that in ether solvents there is an inverse relationship between vinyl content and the charge on the γ -position of the active centre (the reaction site for this structure). The γ -position is however much more reactive as indicated by the free anion product. The charge distribution in this case must surely be very similar to that observed with Cs^+ as counter-ion, yet the polymers produced are quite dissimilar. Steric accessibility of the γ -position must for the larger counter-ions be a major directing

Table III.
Calculated Charges^a on Allylic Positions

M	Solvent	COMPOUND I-M				COMPOUND II-M			
		α	β	γ	Σ Total	α	β	γ	Σ Total
Li	C ₆ H ₆	.79	-.13	.22	.88	.80	-.14	.19	.85
	DEE	.69	-.13	.35	.91	.72	-.15	.30	.87
	THF	.69	-.15	.40	.94	.73	-.15	.34	.92
	DME					.72	-.14	.35	.93
Na	THF	.65	-.12	.49	1.02	.69	-.12	.38	.95
K	THF	.59	-.11	.53	1.01	.61	-.09	.44	.96
Rb	THF	.55	-.11	.53	.97	.58	-.09	.47	.96
Cs	THF	.51	-.12	.52	.91	.54	-.10	.45	.89

^aCharges tabulated as fractions of 1 electron at that position.

Table IV.
Effect of Counter Ion on the % 1,2 Structure in Polybutadiene
Solvent

Counter ion	Solvent		
	DEE	Dioxan (4)	THF (15)
Li	73	87	96
Na		85	91
K	58	55	83
Rb			75
Cs	44	41	74

influence in the reaction. The ability of the lithium ion to reside close to the α -position must leave the γ more open to reaction. The large Cs⁺ can overlap both positions which apparently causes increased attack at the α position, perhaps by facilitating an end approach.

Literature Cited

1. Carton, A. and Bywater, S., *Macromolecules*, 8, 697 (1975).
2. Gebert, W., Hinz, J. and Sinn, H., *Makromol. Chem.*, 144, 97 (1971).
3. Bywater, S. and Worsfold, D.J., *Macromolecules*, in press.
4. Salle, R. and Pham, Q-T., *J. Polym. Sci., Chem. Ed.*, 15, 1799 (1977).
5. Essel, A. and Pham, Q-T., *J. Polym. Sci., A-1*, 10, 2793 (1972).
6. Frei, K. and Bernstein, H.J., *J. Chem. Phys.*, 38, 1216 (1963).
7. Spiesscke, H. and Schneider, W.G., *Tetrahedron Lett.*, 468 (1961).
8. van Dongen, J.P.C.M., van Dijkman, H.W.D. and Bie, M.J.A., *Recl. Trav. Chim. Pays Bas*, 29, 93 (1974).
9. O'Brien, D.H., Russell, C.R. and Hart, A.J., *Tetrahedron Lett.*, 37 (1976).
10. Saunders, M., Telkowski, L. and Kates, M.R., *J. Am. Chem. Soc.*, 99, 8070 (1977).
11. Bywater, S., Patmore, D.J. and Worsfold, D.J., *J. Organometal. Chem.*, 135, 145 (1977).
12. West, P., Purmort, J.I. and McKinley, S.V., *J. Am. Chem. Soc.*, 90, 797 (1968).
13. Saunders, M., Kates, M.R., Wiberg, K.B. and Pratt, W., *J. Am. Chem. Soc.*, 99, 8072 (1977).
14. Saunders, M. and Kates, M.R., *J. Am. Chem. Soc.*, 99, 8071 (1977).
15. Bywater, S. and Worsfold, D.J., *Can. J. Chem.*, 45, 1821 (1967).
16. Rembaum, A., Ells, F.R., Morrow, R.C. and Tobolsky, A.V., *J. Pol. Sci.*, 61, 166 (1962).

Discussion

J. Prud'homme, U. of Montreal, Quebec: With THF as solvent we see that the 1:2 structure is about constant, it does not change much. What would you expect for dioxane solutions?

D.J. Worsfold: In dioxane solution the polymer produced is from reaction of the ion pair almost entirely. As the NMR spectra are taken in concentrated solution, even in THF the structure observed is the ion pair and should correlate with the polymer structure formed in dioxane. In polymerizations in THF solution the polymer is quite largely formed from the reaction of the free ion chain end, and hence is less dependent on the ion pair structure. The NMR spectra were measured at -20°C because these compounds are not very stable, but as dioxane freezes at $+10^{\circ}\text{C}$ it was not possible to use it for the NMR solvent.

RECEIVED March 13, 1979.

Carbon-13 NMR High-Resolution Characterization of Elastomer Systems

CHARLES J. CARMAN

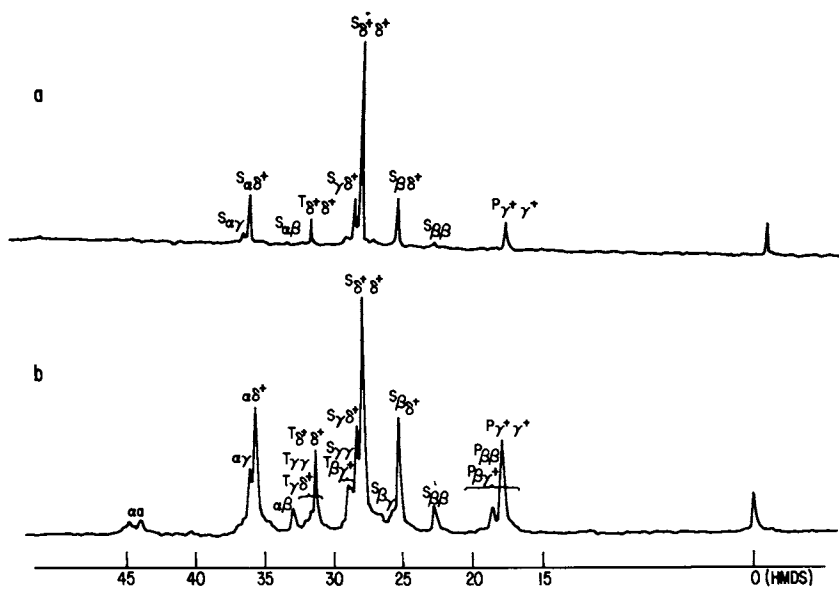
The B. F. Goodrich Research & Development Center, Brecksville, OH 44141

Identification and characterization of elastomers are important analytical information for those involved in rubber technology. It has been shown (1) that Fourier transform carbon-13 nuclear magnetic resonance (^{13}C nmr) spectroscopy is a very powerful method for determining subtle molecular structure features in elastomers. Details from typical information-rich spectra can be used to describe monomer composition and sequence distribution, chain configuration or simply materials identification of elastomer mixtures. These types of analyses are usually made from high resolution solution spectra. That is, a rubber is dissolved in a solvent with noninterfering resonances and the spectrum obtained at high temperature. This produces a spectrum typically with linewidths three to five hertz wide, and consequently has maximum chemical shift information. The advantage of obtaining ^{13}C nmr spectra on solid elastomers was suggested by the early report (2) of a high quality spectrum of solid natural rubber gum, and of the high resolution spectrum obtained when the ^{13}C nmr analysis was combined with magic angle spinning (3) on a cured carbon black filled natural rubber sample. This paper will compare some of the structural information that one can obtain from a solution spectrum with the information available from spectra obtained directly from solid elastomers. Spectra obtained from normal Bloch decays using the conventional proton decoupling typical for organic compounds will be compared to spectra obtained with magic angle spinning and high power proton decoupling. It will be shown that a combination of solution spectra of raw elastomers with solid spectra of cured products usually can be used for materials identification.

Results and Discussion

High Resolution Spectra of Solutions. An example of high resolution solution spectra of an elastomer system which illustrates the sensitivity of ^{13}C nmr to molecular structure is shown in Figure 1. Shown are spectra of ethylene propylene rubbers

0-8412-0505-1/79/47-103-097\$06.25/0
© 1979 American Chemical Society



Macromolecules

Figure 1. Pulsed FT C-13 NMR spectrum of (a) a 71 wt. % C₂H₄ and (b) a 56 wt % C₂H₄ EPDM rubbers obtained at 393 K from a trichlorobenzene solution. (Figure 1b reproduced from Ref. 11.)

TABLE I.

 ^{13}C Chemical Shifts for Ethylene Propylene Rubbers

Species	^{13}C NMR Shift ^a	Occurrence
S $\alpha\alpha$	44.6-43.7	methylene sequence length 1
S $\alpha\beta$	32.9	two in each sequence length 2
S $\alpha\gamma$	b, 35.9, 36.4	two in each sequence length 3
S $\alpha\delta^+$	35.5	two in each sequence length M>3
S $\beta\beta$	22.7	methylene sequence length 3
S $\beta\gamma$	25.8	two in each sequence length 4
S $\beta\delta^+$	25.4	two in each sequence length M>4
S $\gamma\gamma$	28.8	methylene sequence length 5
S $\gamma\delta^+$	28.4	two in each sequence length M>5
S $\delta^+\delta^+$	28.0	M-6 in each sequence length M>6
T $\alpha\beta^+$	c (38.9)	two for each $_ _ _$
T $\beta\beta$	27.0	$_ _ _$ and $_ _ _$
T $\beta\gamma^+$	b, 29.0, 28.8	$_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$
T $\gamma\gamma$	31.6	$_ _ _$ and $_ _ _$
T $\gamma\delta^+$	31.6	$_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$
T $\delta^+\delta^+$	31.3	$_ _ _$, $_ _ _$, $_ _ _$, $_ _ _$
P $\alpha\beta^+$	c (22.9)	attached to T $\alpha\beta^+$ ($_ _ _$)
P $\beta\beta$	b, 19.8, 19.6, 18.9, 18.7	attached to T $\beta\beta$ ($_ _ _$ and $_ _ _$)
P $\beta\gamma^+$	18.7	attached to T $\beta\gamma^+$ ($_ _ _$, $_ _ _$ etc.)
P $\gamma^+\gamma^+$	18.2	attached to T $\gamma\gamma$, T δ^+ , and T $\delta^+\delta^+$

^a downfield from internal HMDS (hexamethyldisiloxane) at 120°C in OCDB or TCB

^b stereostructure produces non-equivalent chemical shifts

^c not detected; predicted (10) chemical shift is shown in parentheses

(reproduced from C. J. Carman, R. A. Harrington, C. E. Wilkes, Macromolecules, (1977), 10, 536.)

TABLE II.
Numbers of Methylene Sequences of Different Lengths

<u>Length</u>	<u>Occurrence</u>	<u>Number in "representative sample"</u>
1	┌┌ and └└	$S_1 = N_2 p_{22} + N_3 p_{33}$
2	┌└ and └┌┌	$S_2 = N_2 p_{23} + N_3 p_{31} p_{12}$
3	┌┌┌ and └└└	$S_3 = N_2 p_{21} p_{12} + N_3 p_{31} p_{13}$
4	┌┌└ and └└┌┌	$S_4 = N_2 p_{21} p_{13} + N_3 p_{31} p_{11} p_{12}$
5	┌┌┌┌, └└└└	$S_5 = p_{11} S_3$
6	┌┌┌└, └└└┌┌	$S_6 = p_{11} S_4$
3+2n	┌┌ and └└	$S_3 + 2n = p_{11}^n S_3$
4+2n	┌└ and └┌	$S_4 + 2n = p_{11}^n S_4$
all		$\Sigma S = N_2 + N_3 - N_3 p_{32}$

(reproduced from C. J. Carman, R. A. Harrington, C. E. Wilkes, Macromolecules, (1977), 10, 536.)

method of steepest descent. Table IV shows how well the peak areas of a ^{13}C nmr spectrum of ethylene propylene rubber (EPDM A) can be fitted using the conditional probability model. The utility of the method is that rubbers of different composition can be analyzed and compared in terms of their composition as well as in terms of relative monomer sequence distribution.

Table V gives a comparison of the two rubbers whose spectra were shown in Figure 1. The compositions are a result of the analysis and are comparable with those obtained from proton nmr or infrared (1). The information on sequence structure, however, is unique to the ^{13}C nmr analysis and is testimony to its power to obtain molecular structure information. The value for " $r_1 r_2$ " shows that these rubbers are not perfectly random but tend toward alternation. The amount of propylene inversion could not be quantitatively measured before the advent of ^{13}C nmr even though infrared could be used to estimate its presence (12). One can see that the amount of propylene inversion is not constant and ^{13}C nmr analysis (11,13) shows this can run quite high.

Table VI yields the methylene sequence distribution of these two EPDM rubbers. A measure of the fraction of long methylene runs is important to the understanding of physical properties of these and similar materials. Polyethylene blends with EPDM rubbers, having appropriate long runs of ethylene, have been shown (14,15) to have unusually high tensile strengths.

TABLE III.

Number of Each Carbon Species as a Function of Sequence

Code	Species	Number
A	S $\alpha\alpha$	S ₁
B	S $\alpha\beta$	2S ₂
D	S $\alpha\gamma$	2S ₃
E	S $\alpha\delta^+$	2 ($\Sigma S-S_1-S_2-S_3$)
C	S $\beta\beta$	S ₃
F	S $\beta\gamma$	2S ₄
G	S $\beta\delta^+$	2 ($\Sigma S-S_1-S_2-S_3-S_4$)
H	S $\gamma\gamma$	S ₅
I	S $\gamma\delta^+$	2 ($\Sigma S-S_1-S_2-S_3-S_4-S_5$)
J	S $\delta^+\delta^+$	2N ₁ + N ₂ + N ₃ - (sum of above numbers)
-	T $\alpha\beta^+$	2N ₃ p ₃₂
K	T $\beta\beta$	N ₂ p ₂₂ ² + N ₃ p ₃₃ ²
L	T $\beta\gamma^+$	N ₁ p ₁₂ p ₂₂ + N ₁ p ₁₃ p ₃₃ + N ₂ p ₂₂ (p ₂₁ + p ₂₃) + N ₂ p ₂₃ p ₃₃ + N ₃ p ₃₃ p ₃₁
M	T $\gamma\gamma$	N ₂ p ₂₃ p ₃₁ p ₁₂ + N ₃ p ₃₁ p ₁₂ p ₂₃
N	T $\gamma\delta^+$	N ₁ p ₁₁ p ₁₂ p ₂₃ + N ₁ p ₁₃ p ₃₁ p ₁₂ + N ₂ p ₂₁ p ₁₂ p ₂₃ + N ₂ p ₂₃ p ₃₁ (p ₁₁ + p ₁₃) + N ₃ p ₃₁ p ₁₂ p ₂₁
O	T $\delta^+\delta^+$	N ₁ p ₁₁ p ₁₂ p ₂₁ + N ₁ p ₁₃ p ₃₁ (p ₁₁ + p ₁₃) + N ₂ p ₂₁ p ₁₂ p ₂₁
-	P $\alpha\beta^+$	same as T $\alpha\beta^+$
P	P $\beta\beta$	same as T $\beta\beta$
Q	P $\beta\gamma^+$	same as T $\beta\gamma^+$
R	P $\gamma^+\gamma^+$	sum of numbers for T $\gamma\gamma$, T $\gamma\delta^+$, and T $\delta^+\delta^+$
	all S	2N ₁ + N ₂ + N ₃
	all T	N ₂ + N ₃
	all P	N ₂ + N ₃
	all species	2N ₁ + 3N ₂ + 3N ₃

(reproduced from C. J. Carman, R. A. Harrington, C. E. Wilkes, Macromolecules, (1977), 10, 536.)

TABLE IV.
 Computer Fit of a ^{13}C nmr Spectrum
 Using Method of Conditional Probabilities

<u>Type</u>	<u>Calculated</u>	<u>Observed</u>	<u>Difference</u>	
S $\alpha\alpha$	5.98	6.00	0.02	
S $\alpha\alpha$	10.92			
S $\alpha\delta^+$	19.99			
	<u>30.91</u>	30.00	-0.91	
S $\alpha\beta$	4.41	4.50	0.09	
T $\gamma\gamma$	0.56			
T $\gamma\delta^+$	2.38			
T $\delta^+\delta^+$	10.27			
	<u>13.31</u>	14.00	0.79	
S $\gamma\gamma$	3.40	}		
T $\beta\beta$	1.53			
T $\beta\gamma^+$	8.89			
S $\gamma\delta^+$	12.45		12.70	0.25
S $\delta^+\delta^+$	26.12		27.50	1.38
	<u>52.39</u>	<u>55.10</u>	<u>2.71</u>	
S $\beta\gamma$	0.75			
S $\gamma\delta^+$	19.24			
	<u>19.99</u>	19.00	-0.99	
S $\beta\beta$	5.46	6.00	0.54	
P $\beta\beta$	1.53			
P $\beta\gamma^+$	8.89			
P $\delta^+\delta^+$	13.21			
	<u>23.64</u>	21.50	-2.14	

TABLE V.
 ^{13}C NMR Analysis of Two EPDM Rubbers

	EPDM A	EPDM B
mol % C_3H_6	35.7	20.1
" r_1r_2 "	0.78	0.60
% inversion	8	48

TABLE VI,

Methylene Sequence Distribution of Two EPDM Rubbers

		Percent Methylene in Sequence Lengths 1-12+											
		1	2	3	4	5	6	7	8	9	10	11	12+
EPDM A		5.50	4.05	15.06	1.38	15.63	1.29	13.62	1.07	10.90	0.83	8.29	22.36
EPDM B		1.55	1.06	3.23	3.50	4.23	4.12	4.64	4.31	4.68	4.23	4.29	59.96

The measurement of methylene sequence length in ethylene propylene rubber has been performed by Randall (13) using a less complicated scheme than the conditional probability method. Ray and coworkers (16) also suggested an analysis for those ethylene propylene rubbers which have no inverted propylene present. The advantage of the statistical model analysis is to test different polymerization models and mechanisms (11) that cannot be tested with a method limited (13) to sequence distributions. Zambelli and coworkers (17,18,19,20,21) have suggested a reasonable model for vanadium-based Ziegler catalyzed ethylene propylene rubbers. Their model has a predominance of secondary insertion for propylene adding to propylene and of primary insertion for propylene adding to ethylene. We expressed the mechanisms of their model in terms of conditional probabilities, which resulted in a model using three parameters instead of five. A comparison of the two models showed little differences in monomer composition; only slight differences in " r_1r_2 " and inversion, but significant differences in a portion of the spectral fit.

Figure 2 shows the profile of the 27-29 ppm spectral region of three polymers which served as models (11) for ethylene propylene rubber. The better agreement between the observed spectrum and the five-parameter model strongly suggests the three-parameter model is less realistic as an explanation for the polymerization mechanism. Table VII compares the observed profiles of EPDM rubbers made with a Ziegler catalyst system. The ratio of $S_{\gamma\gamma}/S_{\beta\gamma}^+$ (i.e., H/L) agrees better with the five parameter model

TABLE VII.

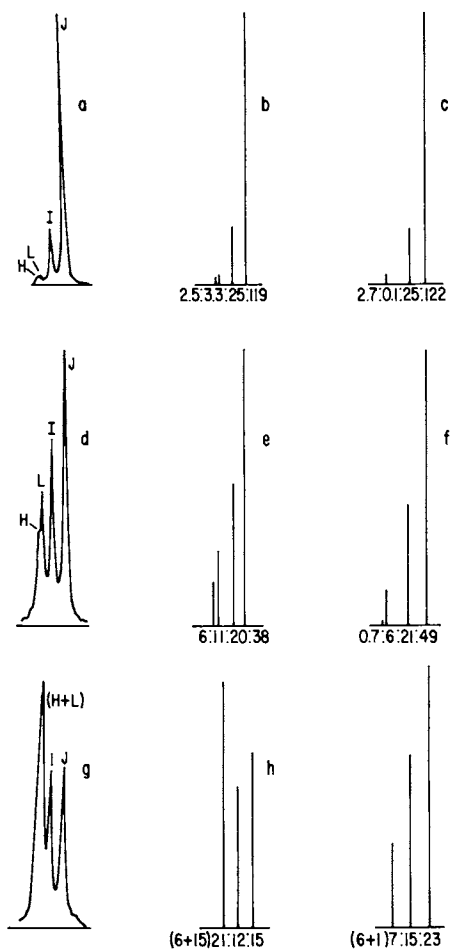
Profiles of 27-29 ppm Spectral Region for Polymerization Models (11)

	<u>mol % C₃H₆</u>	<u>CHW-5</u>	<u>Z-3</u>	<u>Observed</u>
J/I	25.3	2.9	3.0	3.0
	35.5	1.9	2.0	1.9
	51.0	1.3	1.5	-
H/L	25.3	0.98	2.60	0.8
	35.5	0.51	0.81	0.5
	51.0	0.26	0.90	0.4
L/J	25.3	0.07	.02	.07
	35.5	0.22	0.14	.27
	51.0	1.40	0.29	1.3

$$J/I = S\delta^+\delta^+/S\gamma\delta^+$$

$$H/L = S_{\gamma\gamma}/T\beta\gamma^+$$

$$L/J = T\beta\gamma^+/S\delta^+\delta^+$$



Macromolecules

Figure 2. Analysis of 27–29 ppm ^{13}C nmr spectral region (peaks H I J L) for EPDM samples containing 24, 43 and 53 wt % C_3H_6 . (a) experimental; (b) calculated using five-parameter model; (c) calculated for parameters simulating Zam-belli model (11).

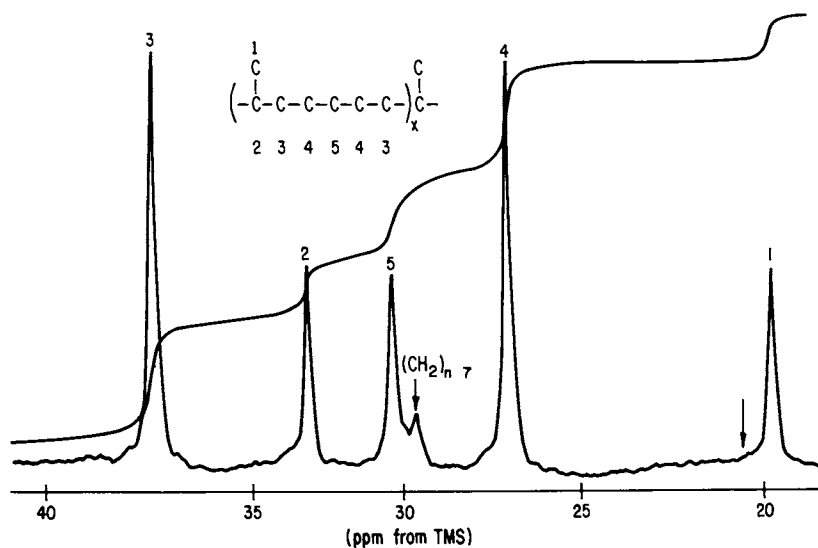
than the three parameter model over the propylene composition range cited.

Just as high resolution, solution spectra have yielded much molecular structure information on ethylene propylene rubbers, the quality of similar data on other elastomer systems has been fruitful. Figure 3 shows the ^{13}C nmr spectra of copolymers of propylene and butadiene made with both a vanadium and a titanium catalyst. The two copolymers were hydrogenated and the ^{13}C nmr spectra of the resulting polyalkanes showed they had regular repeating sequences of five methylene carbons bounded by tertiary carbons bearing methyl groups (22). Based on comparing the chemical shifts of the polyalkanes shown in Figure 4 to those seen in ethylene propylene rubbers and to empirical predictions, the ^{13}C nmr method was unambiguous in assigning a perfectly alternating sequence distribution. The extra resonances seen in Figure 3b for the titanium-made polymer are the result of cis configuration substituent effect and also the presence of cis-1,4-polybutadiene homopolymer. The spectra of the original polymers, shown in Figure 3, show that the copolymer made with a vanadium catalyst is alternating with essentially all of the butadiene in a trans configuration. There appears to be less than 2% polybutadiene homopolymer present as an impurity. On the other hand, the copolymer made with titanium is an alternating copolymer with about 89% of the butadiene in a trans configuration and 11% in a cis configuration. This copolymer also had 7-8% cis-1,4-polybutadiene present as an impurity.

The quantitative use of high resolution solution spectra for the determination of configuration has also been ably demonstrated for other elastomer systems such as polybutadiene (23,24,25), polyisoprene (1), and polyisoprene (1). One would like to explore the possibilities of analyzing solid elastomer systems in terms of high resolution spectral discrimination.

High Resolution Spectra of Solids. Figure 5 compares the spectra of various cis-1,4-polyisoprene samples obtained from the Fourier transform of Bloch decays using typical proton decoupling powers typical for small organic molecules. One can see that the line widths of the cured, carbon black filled rubber are greater than those of the solution spectra. However, the latter spectrum is still narrow enough to provide chemical shifts and allow for material identification. The sample used to obtain the spectrum in Figure 5c was run using magic angle spinning. The results are shown in Figure 6. The lines are narrower than that obtained from the high temperature spectrum shown in 5c. The narrow resonances obtained with the ambient temperature, magic angle spinning have the inherent possibility of assigning resonances of minor structures resulting from cross-linking structures or additives.

Caution should be used in assuming an inherent advantage exists in obtaining a magic angle spectrum of a rubber gum stock. We found the line widths obtained on uncured polyisoprene with



Macromolecules

Figure 4. Pulsed FT C-13 NMR spectrum of the polyalkane obtained by hydrogenating an alternating propylene butadiene copolymer made with a titanium catalyst, Copolymer B. The spectrum was obtained at ambient temperature from a 20% (w/V) solution in 1:1 CCl₄:CDCl₃ (32).

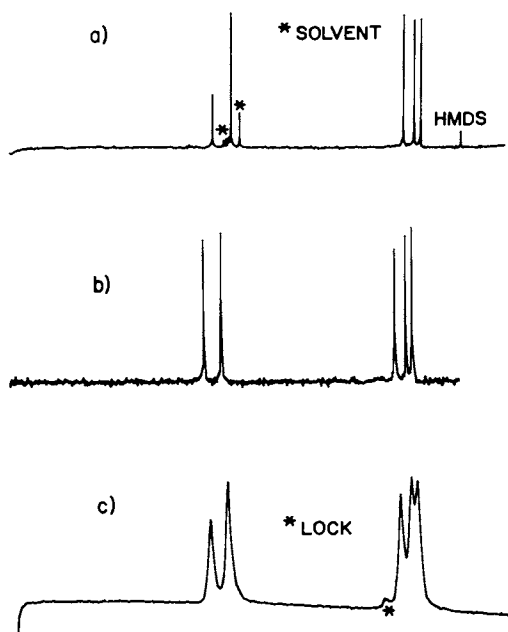


Figure 5. $C-13\{^1H\}$ NMR spectra of *cis*-1,4-polyisoprene: (a) in perchloroethylene at $90^\circ C$; (b) as a solid gum stock at $90^\circ C$; (c) as a cured solid with 50 phr carbon black at $100^\circ C$.

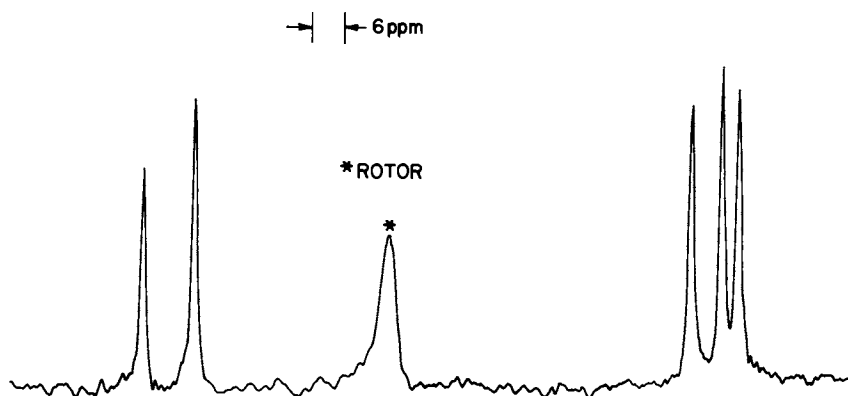


Figure 6. Magic angle spinning, high-power proton decoupling, FT $C-13$ NMR spectrum of cured, carbon-black-loaded polyisoprene at ambient temperature, FT of normal FID without proton enhancement.

magic angle spinning at room temperature were very narrow. But they were no narrower than Figure 5b, a high temperature, normal pulsed spectrum.

A cross polarization experiment (29) without magic angle spinning was attempted and the resulting spectrum is shown in Figure 7. This spectrum is consistent with an interpretation (30) of a superposition of an anisotropic carbon black spectrum and a spectrum of a natural rubber phase of lower mobility on the filler surface. Our attempts to combine the merits of magic angle spinning and cross polarization for this elastomer is a manner similar to that done with glassy polymers (31) have not been successful. Difficulties arise because a rotor must be filled with the soft rubber, rather than fashion the rotor from the polymer (31). Also, the high mobility present in elastomers creates a weak dipolar coupling so that the cross polarization is inefficient and results in weak enhancement compared to standard free induction decay spectra. As far as material identification is concerned, the spectrum resulting from acquiring a standard pulsed free induction decay at an elevated temperature is adequate. Further research will probably show the narrow lines from the magic angle spectra of natural rubber may allow assignments to lesser components.

For comparison to polyisoprene, some spectra of other solid elastomers will be shown to further demonstrate the quality of solid elastomer spectra.

We have previously demonstrated (1) that high resolution spectra of styrene butadiene rubbers (SBR) could be used to distinguish the difference between a solution polymerized SBR and a blend of SBR with high cis polybutadiene rubber (BR) having the same overall styrene content. Figure 8 compares the solution and solid spectra of a 60/40 blend of SBR and BR. The identification of the solid as a blend is evident. Figure 9a shows that the resonances are broadened when the elastomer system is cured and carbon black is present. Figure 9b shows the same sample run with magic angle spinning in a Kel F rotor. Fine structure is beginning to be seen in the aliphatic and aromatic regions and the smaller olefinic carbons are becoming evident. As with the natural rubber system, we have not been able to achieve enhanced sensitivity using cross polarization with these samples.

Figure 10 shows a spectrum of butyl rubber gum stock obtained on the solid at 80°C using normal pulsed FT techniques. Clearly it could be identified as a component in fabricated materials by direct ^{13}C nmr spectral analysis. Figure 11 shows spectra obtained from various portions of typical rubber products. These samples were cut from the rubber product, placed in an nmr tube without solvent, and spectra obtained at an elevated temperature. The data show how polyisoprene, a polyisoprene/polybutadiene blend and a polyisobutylene/polyisoprene/polybutadiene rubber blend are quickly identified in the materials. Figure 11a shows processing oil was present, and which was confirmed by solvent extraction.

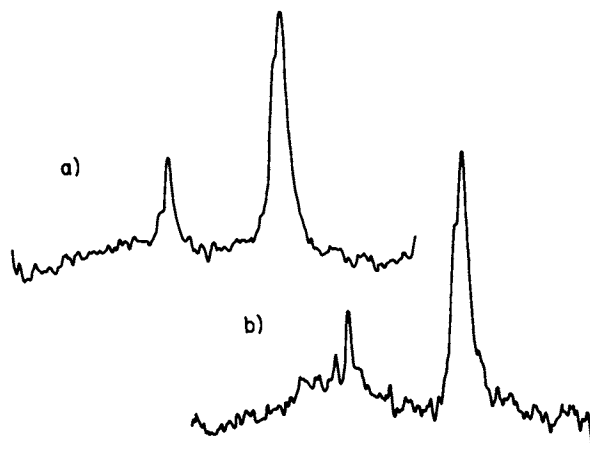


Figure 7. Cross polarization spectra of cured, carbon-black-filled polyisoprene using (a) spin lock of 20 msec, contact time 10 msec and (b) spin lock of 20 msec, contact time 5 msec.

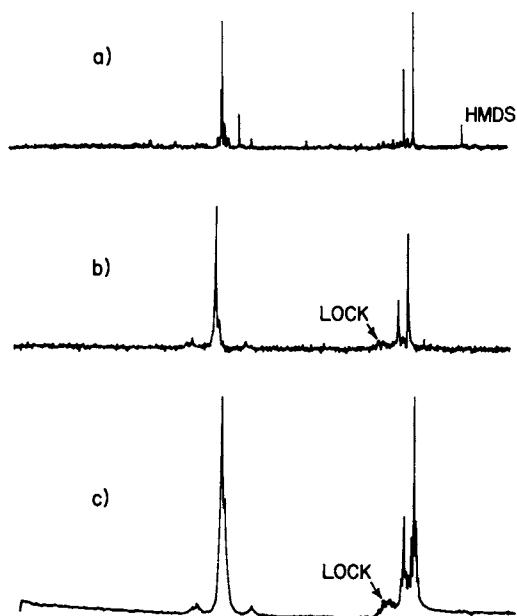


Figure 8. *C-13 NMR spectra of a 60/40 emulsion SBR/polybutadiene rubber blend: (a) in perchloroethylene at 100°C; (b) solid, uncured unfilled rubber at 90°C; (c) solid, cured unfilled rubber at 100°C.*

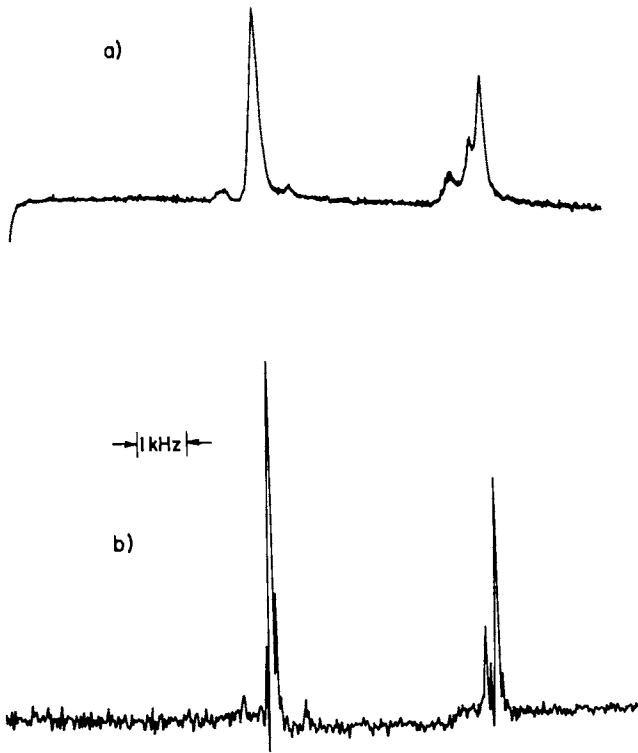


Figure 9. $C-13$ NMR spectra of a 60/40 emulsion SBR/polybutadiene rubber blend: (a) solid, cured, carbon black filled at $100^{\circ}C$; (b) same as Sample a but using magic angle spinning at ambient temperature.

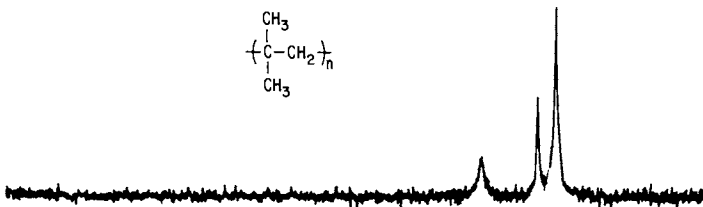


Figure 10. $C-13$ NMR spectrum of butyl rubber as a solid at $70^{\circ}C$

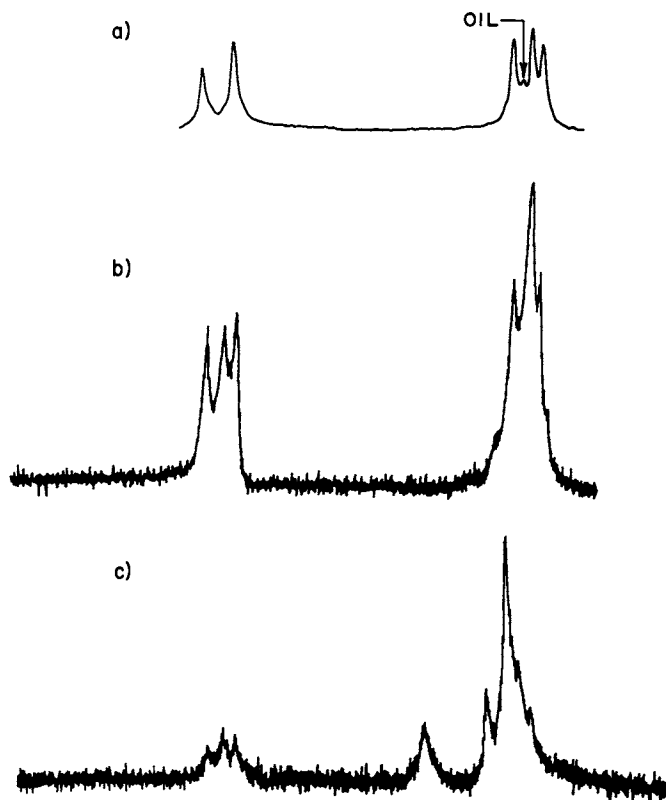


Figure 11. ^{13}C NMR spectra rubber fabricated materials run on solids at 90°C using normal pulsed FT technique: (a) tire tread; (b) tire compound; (c) liner material.

Summary

By examining sections cut from various areas of finished fabricated rubber products, one can easily identify the elastomer component by using standard Fourier transform pulsed nmr techniques combined with high temperature analysis. Such an analytical approach is useful for comparing product uniformity or for the comparison of competitive products. For detailed understanding of subtle molecular defects and associated properties or for studying rubber at filler interfaces, the further development of magic angle spinning and cross polarization techniques for elastomers is anticipated as necessary. Until these combined techniques are used more extensively with practical elastomer systems, one can only estimate their usefulness. The spectral resolution obtained directly on solid elastomers is not as great as the corresponding solution spectra. However, a combination of both types of data provides the macromolecular scientist with a new dimension in understanding both the structure and properties of polymers.

Experimental

The experimental details for the solution spectra have been previously given (1).

The normal Fourier transformed proton decoupled spectra of the solid elastomers were obtained on a Bruker HX-90-E/SXP spectrometer with a 15-inch magnet. The solid samples were cut into small pieces and placed into the inner portion of a 7 mm ID Wilmad coaxial tube. The upper portion of the inner coaxial tube was 9 mm ID and fits precisely inside a standard 10 mm Wilmad tube which contained the high temperature lock compound. Depending on the desired temperature, either D₂O, DMSO-d₆ or 1,4-dibromotetra-deuterobenzene was used as the lock. Ninety degree pulse widths of less than 15 μ s were used with a five second repetition time, 16 K data points and 6 KHz sweep width. The spectra combined with magic angle spinning were obtained courtesy of Bruker Instruments using a Bruker CXP pulsed spectrometer at 45.3 MHz, 1.5 KHz spinning rates, 6 gauss H₁ for proton and a ¹³C $\pi/2$ pulse of about 8.5 μ s. The cross polarization spectrum was obtained courtesy of Dr. J. Schaefer, Monsanto Research, using a spin lock of 20 ms, a single contact of 5 ms and 10 ms and a H₁ for proton of about eight gauss.

The polymer samples were standard commercial rubbers.

Abstract

High resolution ¹³C nmr spectroscopy of elastomers is a powerful tool for identification and characterization of elastomer systems. The ¹³C nmr spectra of elastomers in solution are rich in molecular structure details. The information provided

can be in the form of monomer sequence distribution, chain configuration, steric purity or identification of polymer mixtures. Comparison of ^{13}C spectra of solid elastomers with those from solutions shows that structural information can also be obtained on the solids. Spectra of elastomer solids under conditions of normal free induction decays, cross polarization and magic angle spinning will be described. A few applications to some typical elastomer composites will be discussed.

Literature Cited

1. Carman, C. J. and Baranwal, K. C., Rubber Chem. Technol., (1975), 48, 705.
2. Duch, M. W. and Grant, D. M. Macromolecules, (1970), 3, 165.
3. Schaefer, J., Chin, S. H., and Weisman, S. I., Macromolecules, (1972), 5, 798.
4. Carman, C. J., and Wilkes, C. E., Rubber Chem. Technol., (1971), 44, 781.
5. Wilkes, C. E., Carman, C. J., and Harrington, R. A., J. Polym. Sci., (1973), 43, 237.
6. Crain, W. O., Jr., Zambelli, A., and Roberts, J. D., Macromolecules, (1971), 4, 330.
7. Zambelli, A., Gatti, G., Sacchi, C., Crain, W. O., Jr., and Roberts, J. D., Macromolecules, (1971), 4, 475.
8. Tanaka, Y., and Hatada, K., J. Polym. Sci., Polym. Chem. Ed., (1973), 2057.
9. Grant, D. M., and Paul, E. G., J. Amer. Chem. Soc., (1964), 86, 2984.
10. Carman, C. J., Tarpley, A. R., Jr., and Goldstein, J. H., Macromolecules, (1973), 6, 719.
11. Carman, C. J., Harrington, R. A., and Wilkes, C. E., Macromolecules, (1977), 10, 536.
12. Van Schooten, J., and Mostert, S., Polymer, (1963), 4, 135.
13. Randall, J. C., Macromolecules, (1978), 11, 33.
14. Lindsay, G. A., Singleton, C. J., Carman, C. J., and Smith, R. W., Polymer Preprints, (1978), 19(1), 206.
15. Carman, C. J., Batiuk, M., and Herman, R. M., U. S. Patent 4,046,840 (1977), assigned to BFGoodrich.
16. Ray, G. J., Johnson, P. E., and Knox, J. R., Macromolecules, (1977), 10, 773.
17. Bovey, F. A., Sacchi, M. C., and Zambelli, A., Macromolecules, (1974), 7, 752.
18. Zambelli, A., Wolfsgruber, C., Zannoni, G., and Bovey, F. A., Macromolecules, (1974), 7, 750.
19. Locatelli, P., Provasoli, A., and Zambelli, A., Makromol. Chem., (1975), 176, 2711.
20. Wolfsgruber, C., Zannoni, G., Rigamonti, E., and Zambelli, A., Makromol. Chem., (1975), 176, 2765.
21. Zambelli, A., Tosi, C., and Sacchi, C., Macromolecules, (1972), 5, 649.

22. Carman, C. J., Macromolecules, (1974), 7, 793.
23. Carman, C. J., and Wilkes, C. E., Macromolecules, (1974), 7, 40.
24. Chen, H. Y., J. Polym. Sci., Lett. Ed., (1974), 12, 85.
25. Ivin, K. J., Lavery, D. T. and Rooney, J. J., Makromol. Chem., (1978), 179, 253-258.
26. Clague, A. D. H., van Broekhoven, J. A. M., and Blaauw, L. P., Macromolecules, (1974), 7, 348.
27. Elgert, K. F., Quack, G., and Stutzel, B., Makromol. Chem., (1975), 176, 759.
28. Schaefer, J., Stejskal, E. O., and Buchdahl, R., Macromolecules, (1977), 10, 384.
29. Pines, A., Gibby, M., and Waugh, J. S., J. Chem. Phys., (1973), 59, 569.
30. Schaefer, J., High Resolution ^{13}C nmr Studies of Solid Polymers, "Molecular Basis of Transition & Relaxation", Gordon & Breach Publishers, U.K., 1978, p. 103.
31. Schaefer, J., Stejskal, E. O., and Buchdahl, R., J. Macromol. Soc. Phys., (1977), B13(4), 665.
32. Carman, C. J., Macromolecules, (1974), 7, 789.

Discussion

J. Prud'homme, U. of Montreal, Que.: Did you consider the possibility of studying swollen rubbers (vulcanized materials) to obtain better resolution?

C. J. Carman: There is no advantage. We did not obtain lines any narrower than when the samples were run at higher temperatures. Solvent was excluded because we were also trying to determine effects of relaxation times. The effects are hard to interpret. Solvent present would cause even greater difficulties in the interpretation of the relaxation data. As far as the high resolution quality of the spectra, we see no advantage in having solvent present in these elastomer systems. There is an advantage in swelling a plastic system because mobility of the chains will be affected and higher resolution achieved.

J. Guillet, U. of Toronto, Ont.: It seems to me that when one talks about solids one needs to define what is meant by a solid. In this NMR measurement one is looking at something similar, in terms of its molecular properties, to the kinds of motions necessary for the diffusion of small molecules through polymers. The conventional measurement of polymer viscosity looks at the probability of motion of the center of mass of a very large molecule, which nevertheless is still very small indeed. This leads to very large viscosities for the system. A small molecule moving through a polymer will be assisted by the small scale motions (such as, rotations or vibrations of chains). These latter are the sorts of mobility examined by NMR. Probably the fact that you are dealing with a rubber may simply be that some of the restrictions have been eliminated that would be present if you had a semi-crystalline polymer. This brings me to my

question. Polyethylene for example or an ethylene-propylene rubber will have a glass transition of about -40°C . Are the same results obtained when a solid sample of polyethylene is subjected to these high temperatures as when a strictly amorphous material such as polyisoprene or some of the other rubbers you talked about are run at high temperatures?

C. J. Carman: I agree whole-heartedly with the first part of your statement. We have to define what we mean by a solid and that rubbers or elastomers on an NMR time scale really have a lot of mobility and should not be thought of as solids. This is one reason I presented these data: to stimulate discussion. There is a school of thought that all polymers should be classified as solids and therefore solids techniques should be applicable. Because differences do exist between "solids" I agree with the first part of your statement. With regard to your second point, differences are evident. Spectra of solid EPDM exhibit broader peaks, as one would expect, than do polymers such as polyisoprene. I am not willing to say a whole lot about the advantage of the cross polarization and the magic angle spinning because I found the peaks do not narrow as you just predicted they should and I would predict they would. I think there is still some work to be done. Even though there is some crystallinity present, I am not certain that we can define the crystallinity as measured by x-ray diffraction in terms of the motions and the constraints that bear on the cross polarization experiment. I can't define it. We have not seen good cross polarization effects on these samples and the magic angle spinning didn't narrow the peaks that much more than does a higher temperature. High temperatures do not give as narrow a peak as found in solution spectra of, for example, polyisoprene.

J. Guillet: Can you relate the broadness to the crystallinity or are you at a temperature well above the melting point?

C. J. Carman: We are at a temperature well above the melt.

J. Guillet: The thing that is not realized is that diffusion of benzene through rubber is about the same as benzene through benzene. The kinds of motion found in a liquid are in fact very similar to those found in a rubber. It's just that the translational motion of the large molecules is restricted. It is not too surprising that rubbers would show up in some cases as liquids.

C. J. Carman: Yes, I agree.

J. Guillet: The swelling doesn't help you very much because it hasn't affected the mobility of the molecules. It has helped the macroscopic appearance of viscosity but not the microscopic.

C. J. Carman: Agreed. But as far as those ethylene-propylene polymers which have crystallinity and a T_g of around minus forty or fifty degrees, I expect they might be in a different class. Here there might be some advantage to swelling.

A. Garroway, Naval Research Lab, Washington, DC: A few quick comments. First of all, it's worth reminding ourselves that

the NMR definition of a liquid looks at a specific dipolar or NMR type interaction and asks whether it is averaged out. Depending on what interactions are involved, what molecules are being examined, what experiment is being carried out and what nuclear species is being measured, one may have simultaneously a liquid and a solid preventing characterization of materials on a rheological basis. One can have a partial system. A methyl rotation would be a classic example where one has very fast anisotropic motion. It is necessary therefore to be very careful about applying entropomorphic notions of what a liquid or solid is. Secondly, a comment about cross polarization. It is in fact possible to do cross polarization on pure liquid type systems and by that I mean something in which all dipolar interactions are averaged out. There one uses the J interaction to couple and the advantage that one obtains is that there is some enhancement of signal to noise. There is also a speed-up in time. The experiments formally look quite different because no equilibration of temperature exists but rather an oscillation which goes on continuously. So I agree with everything said but ask that cross polarization not be indited just yet.

C. J. Carman: Actually, I didn't intend to. I'm glad you made the comment you did.

W. G. Miller, U. of Minnesota, MN: My question has reference to your cured systems which were filled with carbon black. I would have thought that a reasonable fraction of the polymer was very much immobilized with the carbon black. If so why do you not see it when cross polarization experiments with spinning are carried out?

C. J. Carman: We did try to look at the bound rubber. It turns out that there still is too much motion in such rubber to get cross polarization. You can see it by bound rubber measurements. On the time scale of this experiment it does not contribute to the spectrum.

W. G. Miller: How do you know you are not seeing bound rubber?

C. J. Carman: Let me answer this way. We know there is bound rubber present using standard techniques. I have yet to see the bound rubber with the ^{13}C NMR technique, is what I am saying. Area measurements of high resolution spectra do not give numbers for bound rubber that coincide with standard measurements. For this reason I am not convinced I have ever seen it with NMR.

J. McAndless, Defense Research, Ottawa: The C-13 NMR technique seems quite good for identifying the major component in the rubber formulations, namely the rubber itself. Is the technique sufficiently sensitive to pick out the antioxidants and the processing oils without having to go through the normal separation techniques?

C. J. Carman: In some cases, yes.

J. McAndless: Does it require use of a differential technique computer based program to subtract out the C-13 NMR rubber

signals?

C. J. Carman: I would like to ask the audience if anybody has been successful in doing NMR subtraction spectroscopy? We haven't been that successful at these very low levels because of artifacts. The appropriate signals can be seen directly without resorting to subtraction.

D. Axelson, Florida State Univ. FL: In the light of what I am going to talk about tomorrow I would like to ask whether you have any numbers for the bulk polyethylene or the bulk ethylene copolymer line widths?

C. J. Carman: I have them but not with me.

D. Alexson: Basically are we talking about 50 Hz, 100 Hz, 300 Hz...?

C. J. Carman: They are on the order of 25-50 Hz. I have to qualify that because most of our measurements are at high temperatures. So when you talk about the bulk, one should specify at what temperature, at what frequency?

D. Axelson: These line widths are for 100°C at 22 MHz?

C. J. Carman: The line widths for EPDM are on the order of 25-60 Hz at 100°C at 22 MHz. They will vary a little.

D. Axelson: Are these high molecular weight polymers?

C. J. Carman: Yes.

J. Guillet: It intrigues me that the EPDM exhibit broadening. I'm wondering whether it is crystalline order that is being observed above the melting point. Is there truly random polymer above the melting point? Would this be a possible explanation for the broadening?

C. J. Carman: I guess it could be, but again I'm not certain because there will be a distribution of chemical shifts which perhaps contributes to the line widths as well. In addition to the latter, other factors can contribute to line broadening. As was indicated by other speakers before me, conformational effects do play a role in polymer spectra, I'm convinced of it. I'm not certain of how to extract it but I'm convinced it is there. In the center of the EPDM spectrum is the bulk of the structure which arises from the long runs of methylene carbons. There are several peaks which are very temperature dependent and very solution dependent. I am not sure how much of the line width is really due to molecular motion as opposed to distribution of chemical shifts. The shifts may differ from those found for the polymer when it is in solution. This is why I am a little hesitant to assign the broadening to crystallinity effects.

RECEIVED March 13, 1979.

Characterization of Carbohydrate Polymers by Carbon-13 NMR Spectroscopy

ARTHUR S. PERLIN and GORDON K. HAMER

Department of Chemistry, McGill University, Montreal, Quebec, H3A 2K6, Canada

Many of the advantages offered by ^{13}C -NMR spectroscopy for studies on macromolecules in general, are represented in applications to carbohydrate polymers. Although for a number of years ^1H -NMR studies have furnished valuable information in the field -- early applications to dextrans (1) and glycosaminoglycans (2) may be cited as examples -- and continue to do so, a serious limitation is imposed by the fact that ^1H -signals of the polymers are often excessively broad, even in spectra recorded at high field. Usually, this problem is less serious with ^{13}C -NMR, and the wide range of chemical shifts (> 200 p.p.m.) for ^{13}C nuclei is another characteristic that favors the resolution of most of the individual signals in a spectrum.

A comparison of spectra of hyaluronic acid (3,4) furnishes an illustration of the advantage frequently offered by ^{13}C -over ^1H -NMR spectroscopy. As seen in Fig. 1, the proton spectrum at 220 MHz conveys little information about the fact that the polymer is composed of a disaccharide repeating sequence (1). By contrast, fourteen signals are evident in the ^{13}C spectrum, thus accounting for each nucleus of sequence 1. It is worth noting that both spectra in Fig. 1 were recorded at an elevated temperature so as to reduce line broadening, although the improvement in resolution obtained was greater for the ^{13}C spectrum. Because of this feature, as is well known (5,6), ^{13}C -NMR spectroscopy can be advantageously employed in studying phase transitions of polymers such as hyaluronic acid (4), and other polysaccharides (7,8).

Characteristic Chemical Shifts.

Fig. 1 also may be used to note some general characteristics of ^{13}C spectra of carbohydrate polymers (9-11). Chemical shifts of anomeric carbons (C-1), in the region of 100-110 p.p.m., are typically well separated from other signals. As compared with C-1 of the related monosaccharides (12-15), the anomeric carbon is strongly deshielded (commonly by 7-10 p.p.m.) through glycoside formation (9), i.e., by the change from O-H to an O-C bond.

0-8412-0505-1/79/47-103-123\$05.00/0
© 1979 American Chemical Society

Similarly, the C-1 resonance of an axial anomer is shielded relative to that of its equatorial isomer. Also very distinctive are signals due to the carbon of a primary alcohol group (C-6, in the region of 60-65 p.p.m.) and to the carboxyl group of an uronic acid moiety. Typically, as seen in Fig. 1, the carboxyl C=O resonance is in the region of 175 p.p.m. although, as noted below (see Fig. 3), it is strongly pH dependent.

Usually, the carbon involved in glycosidic bonding to the anomeric position of the adjacent residue is sufficiently deshielded by this bond as to produce a signal well separated from those of the other classes of carbons. Hence, in Fig. 1, the two signals near 80 p.p.m. are assigned to C-4 of the two residues in 1. The remaining, secondary, carbons have chemical shifts commonly centred around 75 p.p.m. Their signals constitute the most difficult ones to assign, although in certain applications this need not be a serious deficiency. A generally useful approach (9-11, 16,17) to an analysis of these signals involves correlations with spectral data for model compounds -- monosaccharides and oligosaccharides, and derivatives, as well as other polysaccharides. Some examples of chemical shift correlations of this type are given later. Other techniques include selective ¹H-decoupling, and chemical or enzymatic modification. It is fair to say, however, that the discovery of new approaches to this problem could very materially facilitate fuller analyses of the spectra.

The particular array of chemical shifts found for the ¹³C nuclei of a given polymer depends, of course, on such factors as bond orientation, substituent effects, the nature of nearby functional groups, solvation influences, etc. As a specific example, derivatives of the carbohydrate hydroxyl moieties may give rise to chemical shifts widely different from those of the unmodified compound, a fact that has been utilized, e.g., in studies (18) on commercially-important ethers of cellulose. Hence, as illustrated in Fig. 2, the introduction of an O-methyl function causes (14,15) a large downfield displacement for the substituted carbon. This change allows for a convenient, direct, analysis of the distribution of ether groups in the polymer. Analogously, carboxymethyl, hydroxyethyl and other derivatives may be characterized as well (18).

Differentiation of Polymers. Mucopolysaccharides

A program in our laboratory on the chemistry of mucopolysaccharides (glycosaminoglycans) has been greatly enhanced by the availability of ¹³C-NMR. For instance, its application (10,19, 20) to studies on the blood anticoagulant, heparin, has served to re-inforce evidence from ¹H spectra (2) showing that there are two main classes of heparin. One type is exemplified by material extracted from beef lung (B type), which may be depicted almost wholly by a repeat biose structure (2). A second type (A type), from hog mucosa, is shown to contain 15-30% of constituents other

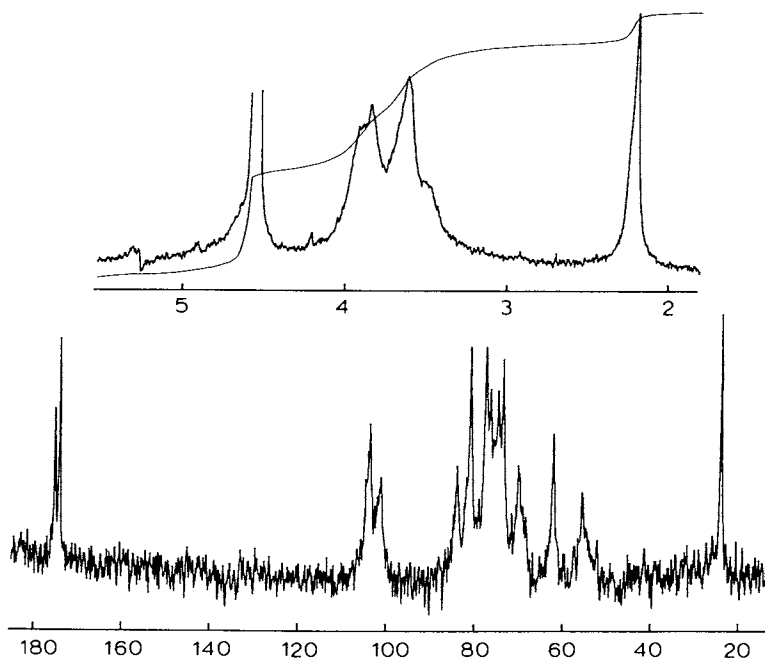


Figure 1. ^1H -NMR spectrum at 220 MHz (upper) and C-13 NMR spectrum at 22.63 MHz (lower) of hyaluronic acid (sodium salt) in D_2O solution. Descriptions of analogous spectra are given in Ref. 3 (^1H) and Ref. 4 (C-13).

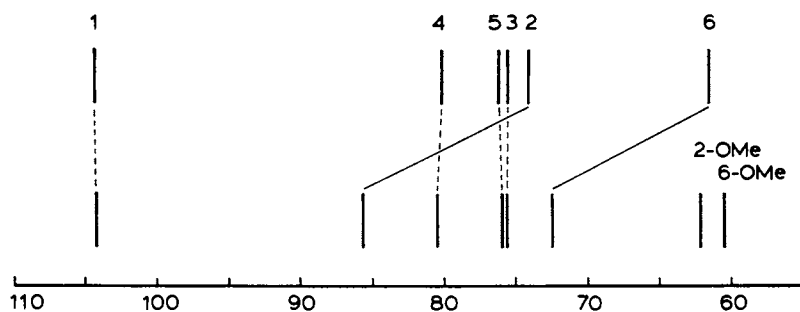


Figure 2. Stick diagram representing the C-13 spectra (18) of methyl β -cellobioside, as a model for cellulose (upper), and a partially substituted O-methylcellulose (2- and 6-O-methyl) (lower). The light lines emphasize the changes in chemical shift associated with the introduction of ether substituents.

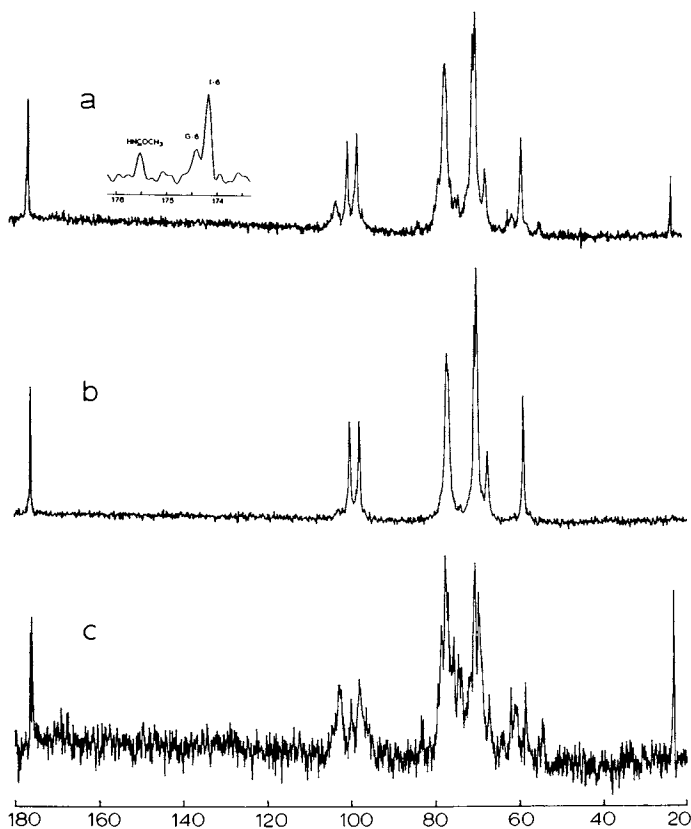


Figure 3. C -13-NMR spectra at 22.63 MHz of heparin A (a) and heparin B (b) as sodium salts in D_2O solution. The inset of spectrum (a) shows the carbonyl region of the heparin A spectrum at pH 2. The difference spectrum (c) represents a subtraction of spectrum b from spectrum a: I = iduronic, G = glucuronic.

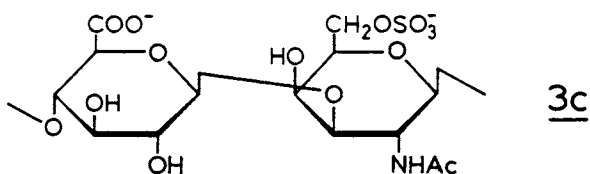
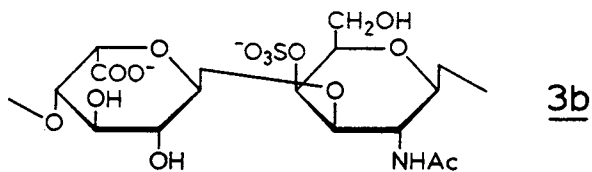
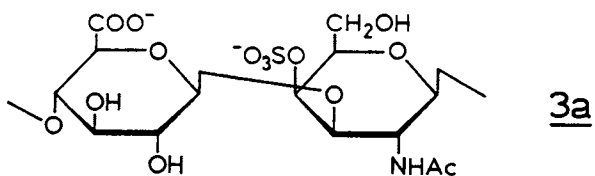
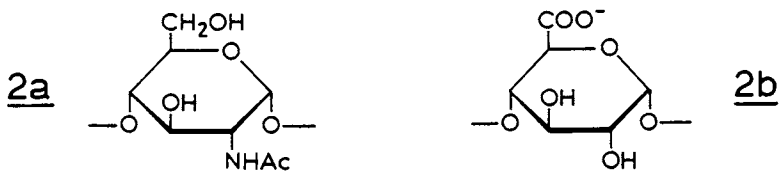
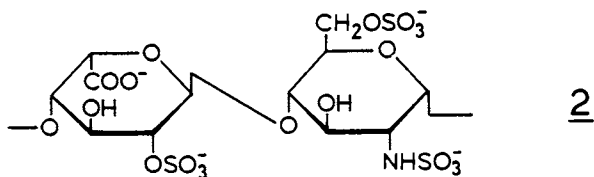
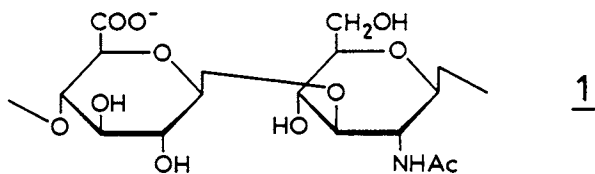
than those in 2. The most prominent evidence for this is the occurrence of an upfield signal (Fig. 3) attributable to an acetamido methyl carbon, a C-1 signal slightly to low field of those of the two main constituents, and at least two relatively weak C=O signals overlapping the main carboxyl peak. However, most of the signals due to these minor constituents are obscured by the prominent ones of the main structural components (2). To extract more information, a difference spectrum has been obtained by subtracting spectrum 3b from 3a, thus eliminating signals of sequence 2 common to both spectra: this gives the pattern shown in Fig. 3c. The latter bears a striking resemblance to the spectrum (21) of a major fraction of a related polymer, heparan sulfate, which consists mainly of acetamido derivative 2a, and β -D-glucuronic acid (2b). Hence, the difference spectrum strongly indicates that these same sugar moieties are the minor constituents of heparin A, and are bonded to adjacent residues in the polymer in an analogous manner as in heparan sulfate. These findings accord well with evidence from several other sources.

Chemical analyses have shown that the compositions of mucosal heparins vary widely. From results such as those of Fig. 3, it is clear that ^{13}C -NMR spectroscopy offers an attractive, rapid means for the characterization of heparins from different sources, a long-standing need in the pharmaceutical field.

An illustration of the power of ^{13}C -NMR for distinguishing between closely-related carbohydrate polymers, as well as for detecting polydispersity, is provided (22) by studies on chondroitins A, B and C. Although ^1H -NMR finds useful application in this area, the ^{13}C spectrum of each of these polymers is more easily recognizable, and more informative. Very striking is the downfield position of the C-6 signal of the hexosamine residue in chondroitin C (3c), due to the presence of the 6-sulfate group (Fig. 4c). In the spectrum of the 4-sulfate (chondroitin A) (3a), by contrast, the major C-6 signal is found in the region typical of an unsubstituted primary alcohol group (Fig. 4a). There is clear evidence from the spectra, moreover, that each polymer contains structural elements of the other, to the extent of 20-30% (22).

Chemical Shift Correlations.

Both chondroitins A and C have residues of 4-linked β -D-glucuronic acid in common (3a and 3c), and there is a close correspondence (22) between the chemical shifts for these residues in the two polymers (Fig. 4a and 4c). Hence structural differences in the hexosamine residues of 3a and 3c, give rise to only minor shielding changes in these acid moieties. A similar analogy is to be found on comparing the hexosamine 4-sulfate residue common to chondroitins A and B (3a and 3b). Here again, the corresponding carbons in the two polymers are characterized by essentially the same chemical shifts (Fig. 4a and 4b).



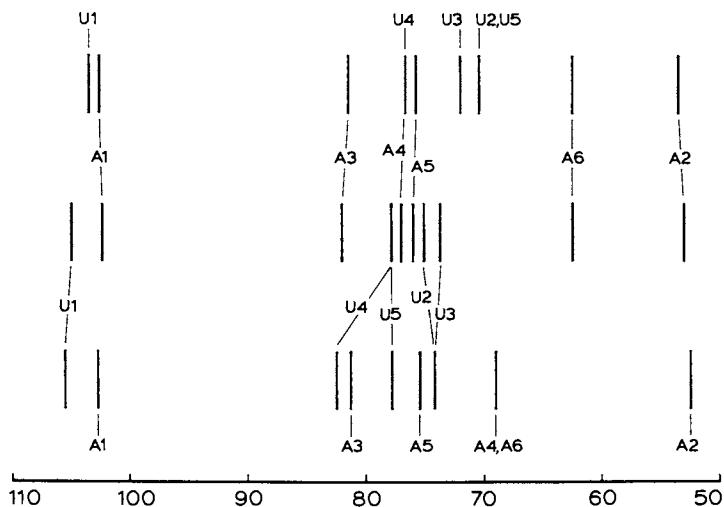


Figure 4. Stick diagrams representing C-13 spectra at 22.63 MHz (22) of chondroitins A, B, and C (4a, 4b and 4c, respectively). The vertical, light lines relate resonances for analogous C-13 nuclei in the three different polymers. Not included are signals caused by the acetamido CH₃ and C=O carbons, and the carboxyl carbons (U-6). Minor signals that demonstrate the presence of chondroitin A in C, and of C in A, are found in Figure 1 of Ref. 22. A = acetamidoxyhexose, U = uronic acid.

It is noteworthy that most of the chemical shift values for all three polymers may be closely approximated (22) by calculations based on data for monomeric reference compounds. These findings illustrate, therefore, the general validity of studies on low molecular weight model compounds for analysis of ^{13}C spectra of carbohydrate polymers. Many examples of equally satisfactory comparisons of this kind are to be found in studies on other polysaccharides (11,23). These polymers include glucans (16), mannans (24,25), limit dextrins (26), lichenin (27), agarose (28) and various polysaccharides of fungal and microbial origins (e.g., 7,8,29-31). Observed departures from expectation have been attributed to specific conformational influences (8).

The studies just cited, and others, serve to illustrate various aspects of the applications of ^{13}C -NMR described above for determination of structure, analysis of constituents, detection of heterogeneity, etc. In this limited review of the subject, unfortunately, it is feasible to deal with only one or two of those many contributions in order to highlight a particular usage.

Enzymic Applications. Agarose.

Studies on agarose and related seaweed polysaccharides, provide an example of how ^{13}C spectroscopic measurements may be combined with the use of selective polymer-degrading enzymes for structural elucidation. In turn, the spectral measurement can become a highly effective means for monitoring characteristics of the enzymic reactions. The degradation of agarose (4a) may occur either by cleavage of the β -(1 \rightarrow 4)-linkage, giving a series of neo-agaro-oligosaccharides (4b) having $\underline{\text{D}}$ -galactose at the reducing end, or by cleavage of the α -(1 \rightarrow 3)-linkage, giving agaro-oligosaccharides (4c) with 3,6-anhydro- $\underline{\text{I}}$ -galactose (in the open-chain, aldehydo form) at the reducing end. The two types of hydrolysis are clearly differentiated by the ^{13}C -NMR spectra of crude enzyme digests (Fig. 5) (28). Cleavage of the β -(1 \rightarrow 4)-linkage is indicated by the appearance of new anomeric signals at 93.8 and 97.8 p.p.m. ($\text{G}'\text{-1a}$, $\text{G}'\text{-1b}$) with intensities in the ratio $\sim 1:2$ (Fig. 5b). In contrast α -(1 \rightarrow 3)-cleavage is indicated by the appearance of a signal at 91.4 p.p.m. ($\text{A}'\text{-1}$), a chemical shift characteristic of hydrated aldehydes (Fig. 5c). The spectra show that the mode of action of each enzyme is specific; there is no evidence of α -cleavage by β -agarase, or of β -cleavage by α -agarase. Integration of the anomeric carbon peak areas provides an estimate of the average chain length of the oligomer mixture. For purified oligosaccharides this may be expressed as n , the number of biose units per oligosaccharide molecule. By examining the systematic variations in the ^{13}C spectra of homologous series of oligosaccharides (4b, 4c; $n = 0,1,2$) together with the results of model compound studies, the peak assignments shown in Fig. 5a can be made. These assignments provide a starting point for determining

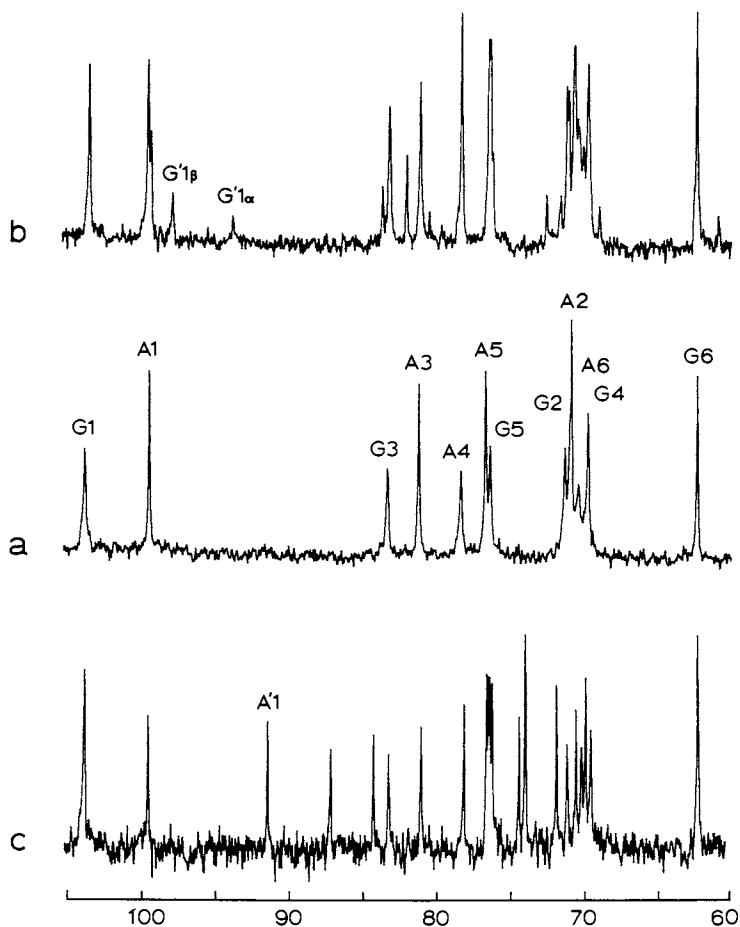


Figure 5. 22.63-MHz C-13-NMR spectra of: (a) agarose (4a); (b) neoagar-oligosaccharides (4b, $n = 3.9$) produced by enzymic hydrolysis of 4a with β -agarase; and (c) agar-oligosaccharides (4c, $n = 2.5$) produced by enzymic hydrolysis of 4a with α -agarase. Spectra recorded in D_2O solution at 35°C (5b,5c) or 95°C (5a). G = D-galactose, A = 3,6-anhydro-L-galactose.

the presence and location of substituents (e.g., $-\text{OCH}_3$, $-\text{OSO}_3^-$) in agars isolated from different seaweed species (32,33).

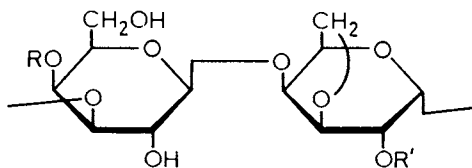
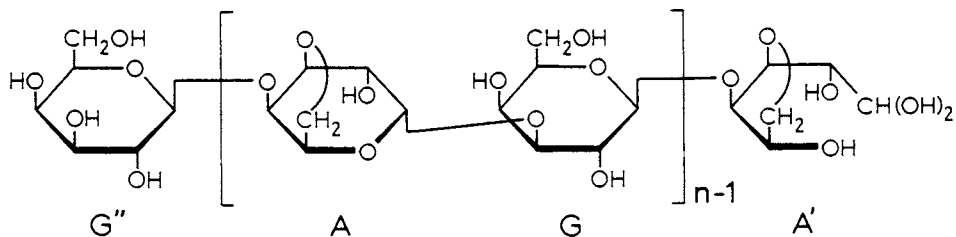
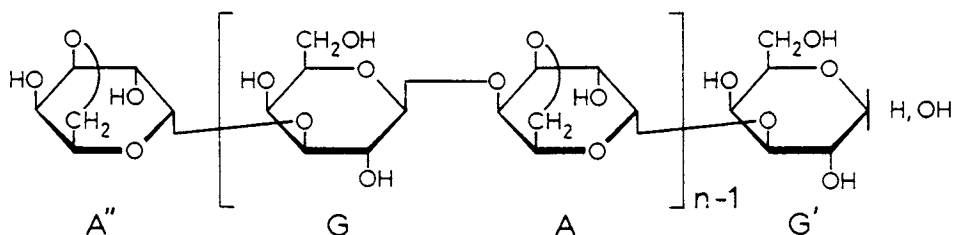
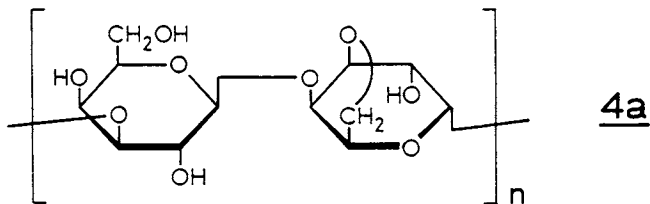
It has also been found that ^{13}C -NMR spectroscopy provides a facile means for discriminating between agars and carrageenans, a closely related family of polysaccharides obtained from red seaweeds (32-35). Both contain 3-O-linked $\underline{\text{D}}$ -galactose and 4-O-linked 3,6-anhydrogalactose residues; in carrageenans however, the 3,6-anhydrogalactose has the $\underline{\text{D}}$ configuration (κ - and ι -carrageenan, 5a and 5b respectively, are representative examples). In each polysaccharide the G-1 signal is found at ~ 103 p.p.m. The A-1 signal, however, occurs at 99.2 p.p.m. (agarose), 96.2 p.p.m. (κ -carrageenan) and 93.1 p.p.m. (ι -carrageenan). Studies of partially desulfated carrageenans (36) using the spectrum subtraction technique described above (see Fig. 2), indicate that the 3 p.p.m. shift difference between κ -carrageenan and agarose is primarily due to the change in 3,6-anhydrogalactose configuration, from $\underline{\text{L}}$ (agarose) to $\underline{\text{D}}$ (κ -carrageenan) whereas the additional 3 p.p.m. shift between ι - and κ -carrageenan is the result of sulfation at A-2.

Monitoring Chemical Change.

Another type of application in which ^{13}C -NMR spectroscopy may be employed to advantage, is in carrying out various chemical modifications of carbohydrate polymers. It is highly effective for monitoring the reduction of uronide residues (20), deaminative degradation of aminosugar-containing polymers (10), acid hydrolysis (18,37), derivatization (18,23), de-esterification; also complex formation (38), interactions with ionic species (20,39,40), etc. In a number of such instances, the spectroscopic measurement may be regarded as the analytical method of choice.

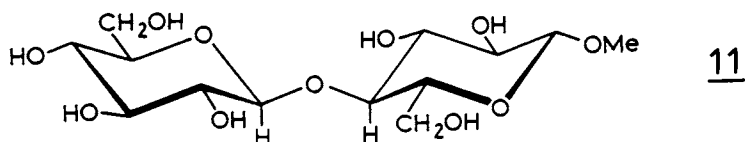
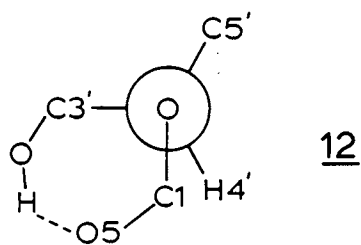
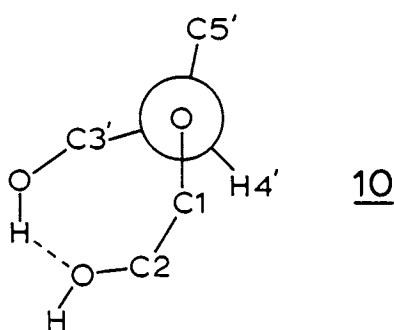
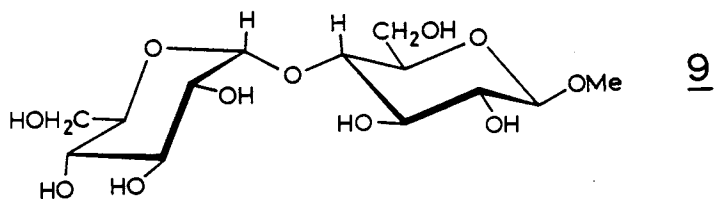
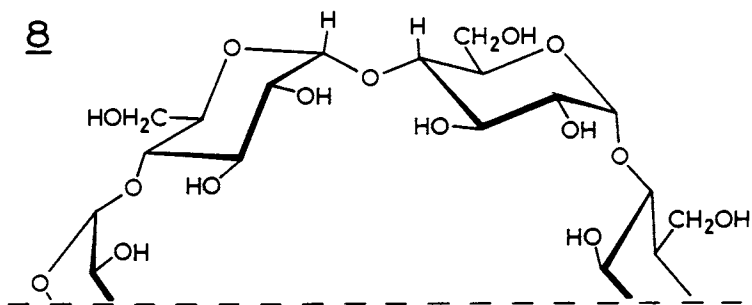
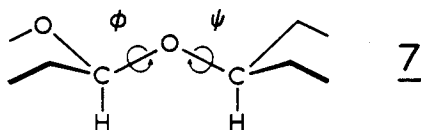
Uses of ^1H -coupled Spectra.

Experimentally, most of what has been described above involves the detection of individual ^{13}C resonance signals, and measurements of their chemical shifts and intensities. For these purposes, the ^1H -decoupled spectrum is ideal because of its relative simplicity and the fact that it gives an optimal signal response. Nevertheless, a coupled spectrum can provide valuable information as well (10). Its simplest use lies in the characterization of classes of carbon -- methyl, methylene, etc. -- from the multiplicity of the observed signal, in which case off-resonance decoupling is often employed. In addition, however, since the magnitude of ^{13}C - ^1H coupling across one, two or three bonds is geometry dependent (12,41) stereochemical information may be obtained. Given the quality of ^1H -coupled spectra now generally available, only measurements of $^1\text{J}_{\text{C-H}}$ are of practical value with polymers of moderate-to-high molecular weight. Signals of anomeric carbons are of particular interest because the magnitude of



5a $R = \text{SO}_3^-, R' = \text{H}$

5b $R = R' = \text{SO}_3^-$



$^1J_{C-H}$ is related in a consistent fashion to the orientation of the C-1,H-1 bond: for an equatorial bond (as in 6a), the coupling is about 10 Hz larger than when, in the anomer (6b), the C-1,H-1 bond is axial (e.g., 170 Hz vs 160 Hz) (12,22,41-44). Also important is the fact that the signal of an anomeric carbon is usually well separated from other signals in the spectrum, as already noted. Hence, problems of configuration and conformation may be examined in a facile manner by ^{13}C -NMR.

Stereochemistry of ^{13}C - 1H coupling in oligosaccharide models.

With monosaccharides, 1H -coupled spectra show splitting of up to ~ 8 Hz due to two- and three-bond coupling (2J and 3J), both of which are stereochemically dependent (12,41,45-47). These fine splittings are not generally resolved in polysaccharide spectra. If, however, one considers an oligosaccharide as a model for polysaccharide conformation, the observed ^{13}C - 1H coupling across the glycosidic bond ($^3J_{COCH}$) in the smaller molecule (e.g. 7) can provide information about the magnitude of inter-residue torsional angles (ϕ and ψ) characteristic of that particular type of linkage (48). This requires a knowledge of the angular dependence of 3J . Although data are available from carbohydrate model compounds for a variety of ^{13}C -O-C- 1H arrays of nuclei, it is difficult to find molecules that can furnish appropriate 3J values for the 0° - 60° region of the Karplus curve relating 3J to the dihedral angle. One suitable molecule is cyclohexaamylose (formula 8 depicts three of its six glucose residues). According to crystallographic and theoretical studies the glycosidic bonds are so constrained by its cyclic structure that ϕ (the angle relating H-1 with C-4 in 8) and ψ (that relating C-1 with H-4) are about $\pm 10^\circ$.

Although C-1 (or C-4) in 8 may couple with several protons other than H-4 (or H-1), and hence give rise to a complex signal (49), the introduction of deuterium at positions 2,3 and 6 through facile catalytic H-D exchange (50) improves things materially. Analysis of the C-1 signal for deuterated 8, with the aid of computer simulation has given a value of 4.8 Hz due to the inter-residue coupling between C-1 and H-4, and of 5.2 Hz due to that between C-4 and H-1. With these data, the Karplus curve has been extended to cover the overall range, as shown in Fig. 6.

This curve could then be used in examining the conformations of disaccharide models of interest. Methyl β -maltoside (9), for which inter-residue couplings of 2.5-3.0 Hz have been estimated (49), may be depicted by orientations of the linkage region in which the time averaged torsional angles are of the order of 45 - 50° (10) (the alternative of $\sim 140^\circ$ is regarded as improbable on the basis of other data for 9 from experimental and theoretical sources). Hence torsional angles ϕ and ψ of the β -maltose moiety in aqueous solution are much larger than those -- 0° - 15° -- that characterize the molecule in the solid state.

Measurements of inter-residue coupling in methyl

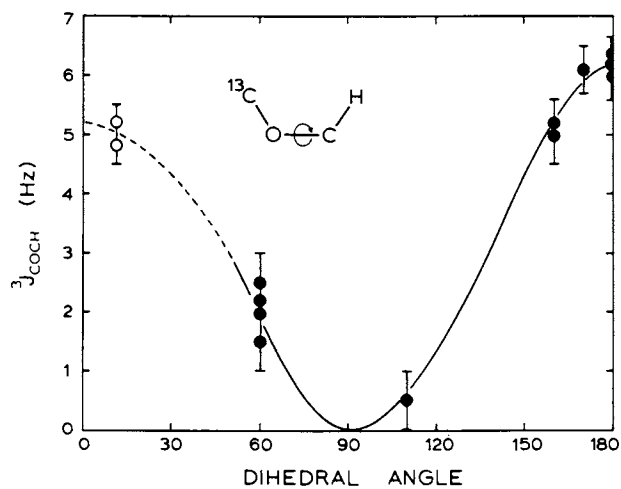


Figure 6. A plot of $^3J_{C-H}$ vs. torsional (dihedral) angle for $^{13}C-O-C-^1H$ arrays of nuclei: (○), values obtained from the spectrum of cyclohexaamylose- d_4 .

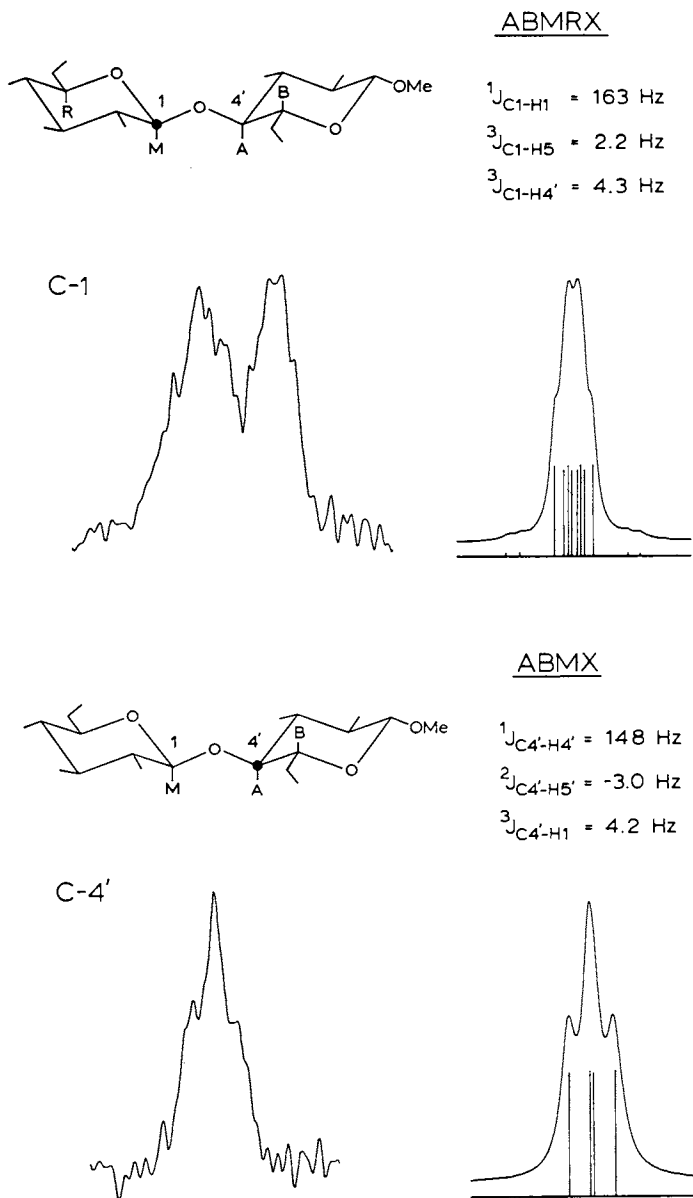


Figure 7. (Upper) C-1 signal of methyl β -cellobioside- d_8 (in D_2O solution). The multiplet to the left that overlaps it partially is the C-1' signal. Computer-simulated C-1 signal is shown on the right. (Lower left), C-4' signal of methyl β -cellobioside- d_8 ; and (lower right), computer-simulated C-4' signal.

β -cellobioside (11) (51) proved to be highly complex but, as with 8, extensive deuteration helped very materially in simplifying the ^{13}C spectra. The C-1 and C-4' signals for methyl β -cellobioside- d_8 , shown in Fig. 7, are much more amenable to analysis than those of the non-deuterated molecule. Nevertheless, the ^3J coupling constants have been extracted (51) by computer simulation, because the spectra of Fig. 7 are not first order. For example, the coupling of C-1 with H-4' is affected by strong coupling between the latter and H-5', and the overall treatment required is that of an ABMRX case. With the aid of simulation, the value of inter-residue coupling between C-1 and H-4' was found to be 4.3 Hz, and between C-4' and H-1, 4.2 Hz. Hence, torsional angles ϕ and ψ in methyl β -cellobioside in D_2O solution are $\sim 30^\circ$ (12), values that correspond rather closely to those in the conformation favored in the crystalline state of the molecule.

Summary.

In recent years, ^{13}C -NMR spectroscopy has found extensive use in studies on carbohydrate polymers, in some series overshadowing the importance of ^1H -NMR spectroscopy. Applications range through determination of primary structure, analysis of mixtures, monitoring of chemical and enzymic transformations or of chelation reactions, and studies on conformational change. Measurements of ^{13}C - ^1H coupling are utilized in determining the configuration and conformation of glycosidic bonds and, in addition, difference methods and simulation experiments are employed as an aid in the analysis of complex spectra.

Acknowledgments.

The authors express their gratitude to the National Research Council of Canada and the Faculty of Graduate Studies, McGill University, for generous support.

Literature Cited.

1. W.M. Pasika and L.H. Cragg, *Can. J. Chem.*, 41, 293,777(1963).
2. A.S. Perlin, M. Mazurek, L.B. Jaques and L.W. Kavanagh, *Carbohydr. Res.*, 7, 369(1968); L.B. Jaques, L.W. Kavanagh, M. Mazurek and A.S. Perlin, *Biochem. Biophys. Res. Commun.*, 24, 447(1966).
3. A.S. Perlin, B. Casu, G.R. Sanderson and L.F. Johnson, *Can. J. Chem.*, 48, 2260(1970).
4. A. Darke, E.G. Finer, R. Moorhouse and D.A. Rees, *J. Mol. Biol.*, 92, 477(1975).
5. M.W. Duch and D.M. Grant, *Macromolecules*, 3, 165(1970).
6. J. Schaeffer, *Macromolecules*, 4, 110(1971).
7. H. Saito, T. Ohki and T. Sasaki, *Biochemistry*, 16, 908 (1977).

8. H. Saito, T. Ohki, N. Takasuka and T. Sasaki, *Carbohydr. Res.*, 58, 293(1977).
9. D.E. Dorman and J.D. Roberts, *J. Am. Chem. Soc.*, 93, 4463 (1971).
10. A.S. Perlin, N.M.K. Ng Ying Kin, S.S. Bhattacharjee and L.F. Johnson, *Can. J. Chem.*, 59, 2437(1972).
11. A.S. Perlin, in G.O. Aspinall (Ed.), *M.T.P. Rev. Sci., Org. Chem., Series 2, Carbohydrates*, 7, 21(1976).
12. A.S. Perlin and B. Casu, *Tetrahedron Lett.*, 2921(1969).
13. L.D. Hall and L.F. Johnson, *Chem. Commun.*, 509(1969).
14. D.E. Dorman and J.D. Roberts, *J. Am. Chem. Soc.*, 92, 1355 (1970).
15. A.S. Perlin, B. Casu and H.J. Koch, *Can. J. Chem.*, 48, 2596 (1970).
16. H.J. Jennings and I.C.P. Smith, *J. Am. Chem. Soc.*, 95, 606 (1973).
17. P.A.J. Gorin, *Carbohydr. Res.*, 39, 3(1975).
18. A. Parfondry and A.S. Perlin, *Carbohydr. Res.*, 57, 39(1977).
19. A.S. Perlin, *International Symposium on Macromolecules (IUPAC)*, Rio de Janeiro, Brazil, July 26, 1974.
20. A.S. Perlin, *Federation Proceedings*, 36, 106(1977).
21. E. Mushayakarara, M.Sc. thesis, McGill University, Montreal, Quebec, August 1977.
22. G.K. Hamer and A.S. Perlin, *Carbohydr. Res.*, 49, 37(1976).
23. M. Vincendon, *Bull. Soc. Chim. Fr.*, 3501(1973).
24. P.A.J. Gorin and J.F.T. Spencer, *Can. J. Microbiol.*, 18, 1709(1972).
25. P.A.J. Gorin, *Can. J. Chem.*, 51, 2105(1973).
26. T. Usui, N. Yamaoka, K. Matsuda, K. Tuzimura, H. Sugiyama, S. Seto, *J. Chem. Soc. Perkin Trans. I*, 2425(1973).
27. D. Gagnier, R. Marchessault, M. Vincendon, *Tet. Lett.* 53(1975)
28. G.K. Hamer, S.S. Bhattacharjee and W. Yaphe, *Carbohydr. Res.*, 54, C7(1977).
29. A.K. Bhattacharjee, H.J. Jennings, C.P. Kenny, A. Martin and I.C.P. Smith, *J. Biol. Chem.*, 250, 1926(1975).
30. D.R. Bundle, I.C.P. Smith and H.J. Jennings, *J. Biol. Chem.*, 249, 2275(1974).
31. G.G.S. Dutton, K.L. Mackie, A. V. Savage and M.D. Stephenson, *Carbohydr. Res.*, 66, 125(1978).
32. S.S. Bhattacharjee, W. Yaphe and G.K. Hamer, IX th International Seaweed Symposium, Santa Barbara, California (1977).
33. A.S. Shashkov, A.I. Usov and S.V. Yarotsky, *Bioorgan. Khimiya*, 4, 74(1978).
34. S.V. Yarotsky, A.S. Shashkov and A.I. Usov, *Bioorgan. Khimiya*, 3, 1135(1977).
35. S.S. Bhattacharjee, W. Yaphe and G.K. Hamer, *Carbohydr. Res.*, 60, C1(1978).
36. G.K. Hamer, unpublished results.
37. J.W. Blunt and M.H.G. Munro, *Aust. J. Chem.*, 29, 975(1976).
38. P.A.J. Gorin and M. Mazurek, *Can. J. Chem.*, 51, 3277(1973).

39. G. Gatti, B. Casu, N. Cyr and A.S. Perlin, Carbohyd. Res., 41, C6(1975).
40. A.S. Perlin, G.K. Hamer, B. Casu and G. Gatti, Polymer Preprints, 19, No. 2, 10(1978).
41. J.A. Schwarcz and A.S. Perlin, Can. J. Chem., 50, 3667(1972).
42. K. Bock, J. Lundt and C. Pederson, Tetrahedron Lett., 1037 (1933).
43. K. Bock and C. Pederson, J. Chem. Soc. Perkin Trans. II, 293(1974).
44. F.R. Taravel and P.J.A. Vottero, Tetrahedron Lett. 2341 (1975).
45. J.A. Schwarcz, N. Cyr and A.S. Perlin, Can. J. Chem., 53, 1872(1975).
46. N. Cyr, G.K. Hamer and A.S. Perlin, Can. J. Chem., 56, 297 (1978).
47. R.U. Lemieux, T.L. Nagabhushan and B. Paul, Can. J. Chem., 50, 773(1972).
48. A.S. Perlin, N. Cyr, R.G.S. Ritchie and A. Parfondry, Carbohyd. Res., 37, C1(1974).
49. A. Parfondry, N. Cyr and A.S. Perlin, Carbohyd. Res., 59, 299(1977).
50. F. Balza, N. Cyr, G.K. Hamer, A.S. Perlin, H.J. Koch and R.S. Stuart, Carbohyd. Res., 59, C7(1977); H.J. Koch and R.S. Stuart, Carbohyd. Res., 59, C4(1977).
51. G.K. Hamer, F. Balza, N. Cyr and A.S. Perlin, Can. J. Chem., in press.

Discussion.

R.H. Marchessault (Xerox, Mississauga): You are to be congratulated on carrying out the analysis of the dihedral angles, because I know this is a difficult thing to do and that you have been trying to do it for some time in Dr. Perlin's lab. The precision you got though, I would guess, is in the neighborhood of ± 20 degrees, indicating the reliability of the measurement. Unequivocally one can say you are on one side of the curve or the other. For the cellobiose case the results seem to fall in with what we expect for β -glycosides. Can the reliability of the measurement be increased? Obviously, the crystallographer has an ideal case. He is dealing with a single crystal where everything is stable and there is only one angle involved. There must be a certain amount of averaging in your case.

G.K. Hamer: That is certainly true. I think the problem is not that we can't measure the coupling constants. I think the accuracy with which we can measure the coupling constants is pretty good because it turns out that the simulations are extremely sensitive to small changes in chemical shifts, and the changes in chemical shifts that are important are in the proton spectrum. They are minute changes. The real problem is in the Karplus equation and in parameterising the curve. I think one

can't push it a great deal further. Another point involves the coupling pathways. One is H-1 through to C-4', the other is H-4' through to C-1. They are not strictly identical because in the one case C-1 is attached to two oxygens whereas C-4' is attached to only one oxygen. The question is, can one use the same curve to derive angles for both interglycosidic bonds?

R.H. Marchessault: I would like to encourage you to keep going because its the first direct measure of the ϕ - ψ angles in solution that we have seen.

F. Seymour (Baylor Medical Center, Texas): In the heparin spectrum for the hog mucosa there was a rather prominent peak around 25 p.p.m. Do you know its origin? Can you comment on that peak, and its relation to the spectrum of normal bovine lung?

G.K. Hamer: Yes, that peak comes from the N-acetyl function which is substituted for N-sulfate on the glucosamine.

F. Seymour: Is it present in the spectrum of the bovine lung?

G.K. Hamer: No, it is not. Using the educated eye (once you know its there in the A type) if you look at the B type very closely you see a tiny peak in the same position. If you look at the 220 MHz proton spectrum it is much more obvious. There is a very small amount in the bovine. The main differences between the two sources are a) N-acetyl substitution for N-sulfate and b) the change in uronic acid.

RECEIVED March 13, 1979.

Relaxation Studies in the System Poly(ethyl methacrylate)-Chloroform by Carbon-13 and Proton NMR

LENAS J. HEDLUND, ROBERT M. RIDDLE, and WILMER G. MILLER

Department of Chemistry, University of Minnesota, Minneapolis, MN 55455

Nuclear magnetic resonance spectroscopy of dilute polymer solutions is utilized routinely for analysis of tacticity, of copolymer sequence distribution, and of polymerization mechanisms. The dynamics of polymer motion in dilute solution has been investigated also by proton¹⁻⁷ and by carbon-13⁵⁻¹⁵ NMR spectroscopy. To a lesser extent the solvent dynamics in the presence of polymer has been studied.¹⁶⁻¹⁸ Little systematic work has been carried out on the dynamics of both solvent and polymer in the same system.

The concentration dependence of polymer or solvent motion has been studied only rarely over a wide range in concentration.¹⁹⁻²² Typically, polymer carbon-13 relaxation is not concentration dependent up to 20-30 percent polymer. Little is known concerning the concentration dependence of the solvent motion.

Carbon-13 relaxation depends predominantly on intramolecular contributions, whereas proton relaxation is sensitive to intermolecular as well as intramolecular interactions. However, by use of isotope dilution²³ the two types of interactions may be separated. Studies utilizing both nuclei can thus yield complementary information.

The experimental design was to study both the carbon-13 and proton relaxation as a function of temperature for both polymer and solvent, and to extend these to as high a polymer concentration as the available equipment permitted. Inasmuch as the mechanical properties of polymers can be affected considerably by small amounts of diluents, we would ultimately like to approach the bulk polymer state, where use of strong dipolar decoupling and magic angle spinning are necessary.²⁴

We chose the system poly(ethyl methacrylate)-chloroform (PEMA-CHCl₃) for several reasons. Karim and Bonner,²⁵ using a PEMA packed gas chromatography column, have shown that CHCl₃ has the strongest interaction with bulk PEMA out of the thirty solvents investigated. Secondly, although PEMA has not been so thoroughly studied as poly(methyl methacrylate), PMMA, its chain dynamics in solution should resemble PMMA, which in bulk has also been studied.²⁶ Finally, the dynamics of neat chloroform has been studied by NMR²⁷⁻³² and by dielectric relaxation.^{33,34}

0-8412-0505-1/79/47-103-143\$05.00/0

© 1979 American Chemical Society

Experimental

Poly(ethyl methacrylate) (Cellomer Associates) was vacuum dried at 50°C. The molecular weight (M_w) was determined to be 3.3×10^5 from its intrinsic viscosity in ethyl acetate.³⁵ Chloroform (spectral grade) and deuteriochloroform (MSD Isotopes) were used as received. Prior to sample preparation the solvent was degassed using five freeze-thaw cycles. The solvent was vacuum distilled onto the polymer in a 12 nm NMR tube, and sealed. ^1H and ^{13}C spin-lattice relaxation times were made on a Varian Associates XL-100 - 15/VFT-100 Spectrometer operating at 100.1 and 25.2 MHz, respectively, using differential mode³⁶ inversion recovery or saturation recovery³⁷ using homospoil. ^1H and ^{13}C T_1 measurements were made using either an external ^{19}F or an internal ^2H field frequency lock. Either integrated peak areas or peak intensities were used to determine T_1 from a linear plot of intensity versus delay depending on the linewidth of the resonance. For sharp resonances no difference was found in T_1 's determined from integrated areas or intensities.

Results

The ^{13}C spectrum of 6, 20 and 40 wt. % PEMA solutions at 34°C are shown in Figure 1. All of the resonances are easily discernible except for the backbone methylene at 40%. At low concentration the polymer $\alpha\text{-CH}_3$, quaternary carbon, and backbone methylene carbon exhibit resolved or partially resolved chemical shifts due to the various stereochemical sequences since the polymer was not stereoregular. A rough estimate indicates the polymer is essentially atactic.

The ^{13}C relaxation behavior of chloroform as a function of temperature and polymer concentration is shown in Figure 2, and the corresponding nuclear Overhauser enhancement factors in Figure 3. The values for the neat solvent are in rather good agreement with literature values.²⁸ Addition of as little as three percent polymer is seen to have a measurable affect on the solvent relaxation. As the polymer concentration is increased further there is a systematic lowering of T_1 . At 30% and higher, a T_1 minimum is observed.

In Figures 4-6 the temperature and concentration dependence of the quaternary carbon, α and ester methyls, and ester methylene ^{13}C relaxations are shown. Data on the backbone methylene have not yet been accumulated, nor have the nuclear Overhauser enhancements on the polymer carbons. At low polymer concentration the difference in T_1 among the stereotriads for a given ^{13}C was less than experimental error. At higher concentration only an average could be determined. In general the concentration dependence of T_1 is considerably less than that observed with the solvent. A T_1 minimum is found for the ester methylene, and the quaternary carbon relaxation. The minimum moves to higher temperature with increasing polymer concentration, analogous to the

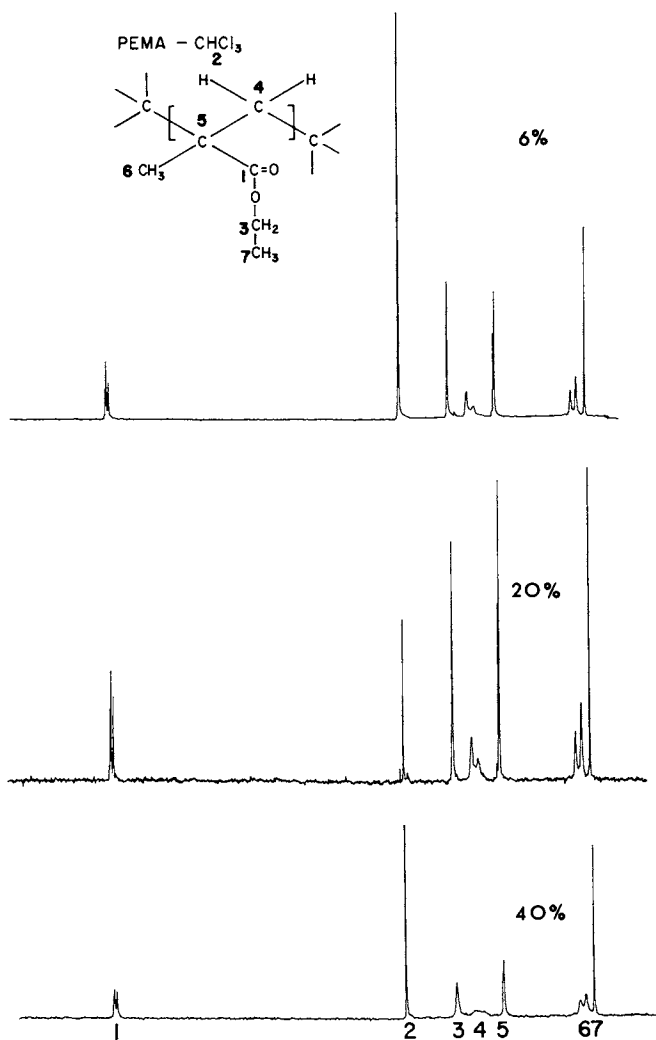


Figure 1. C-13 spectrum of 6, 20, and 40 wt % PEMA solutions at 34°C

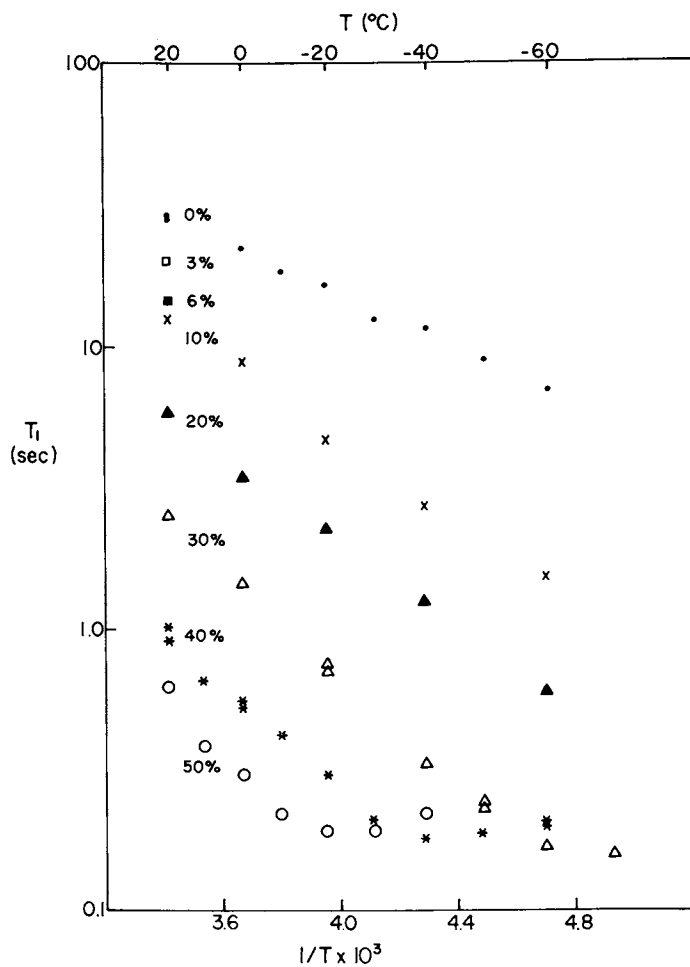


Figure 2. Chloroform C-13 relaxation as a function of temperature. The polymer concentration (weight percent) is as indicated.

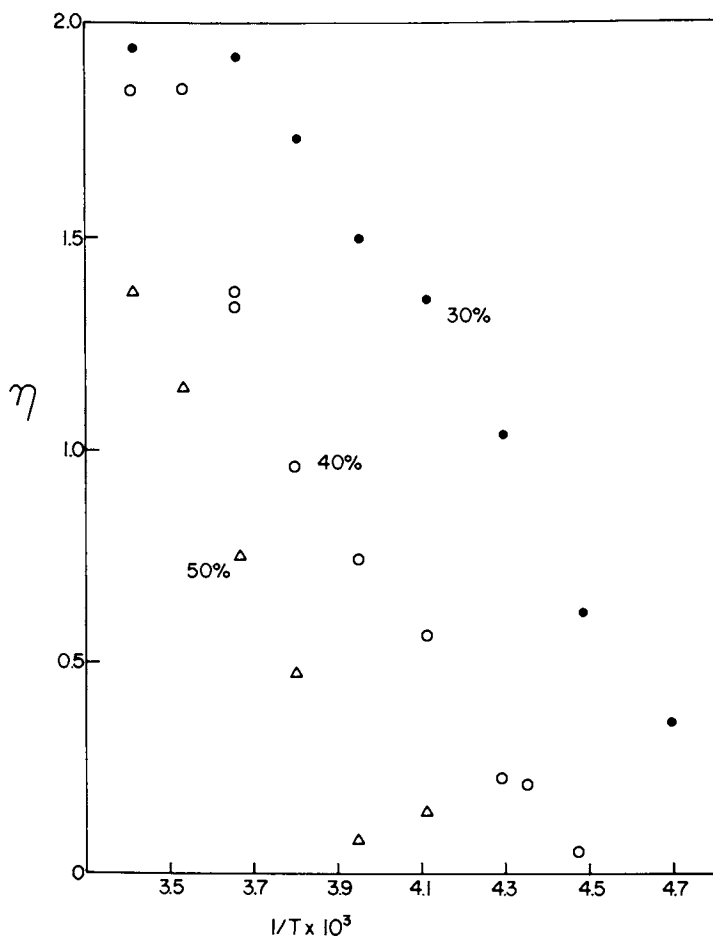


Figure 3. Chloroform nuclear Overhauser enhancement factor as a function of temperature at the indicated polymer concentrations.

**American Chemical
Society Library**

1155 16th St. N. W.

In Carbon-13 NMR in Polymer Science; Pasika, W.;
ACS Symposium Series; American Chemical Society: Washington, DC, 1979.

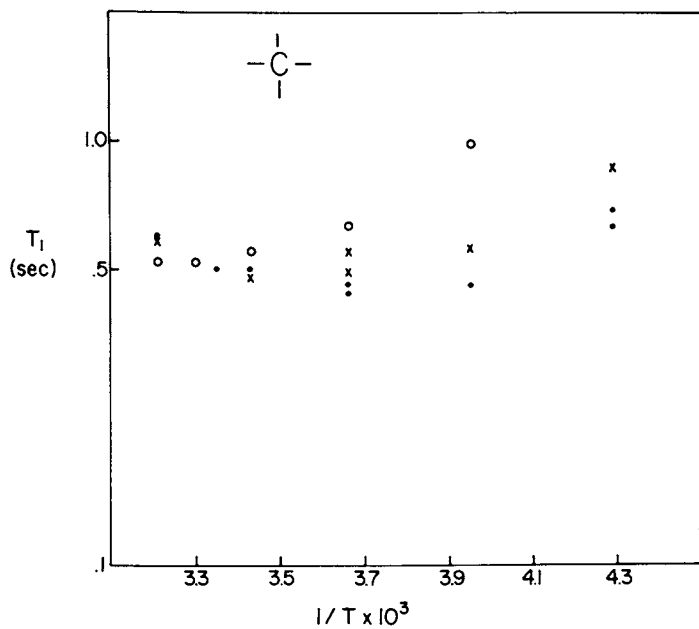


Figure 4. Temperature and concentration dependence of the quaternary C-13 relaxation: (●), 10%; (×), 20%; (○), 30%.

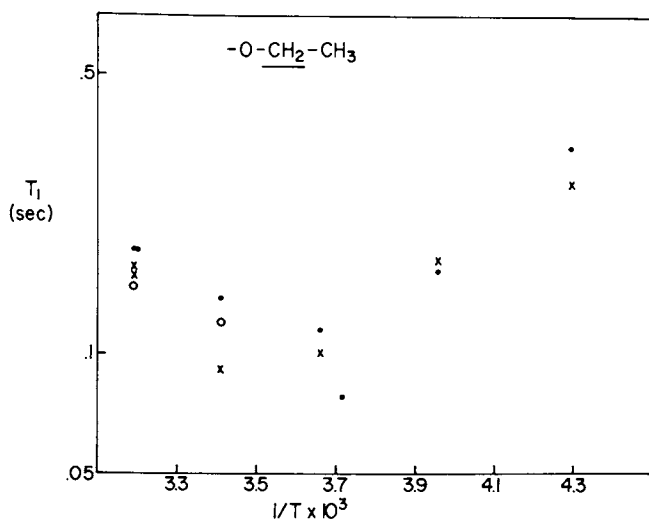


Figure 5. Temperature and concentration dependence of the ester methylene C-13 relaxation: (●), 10%; (×), 20%; (○), 30%.

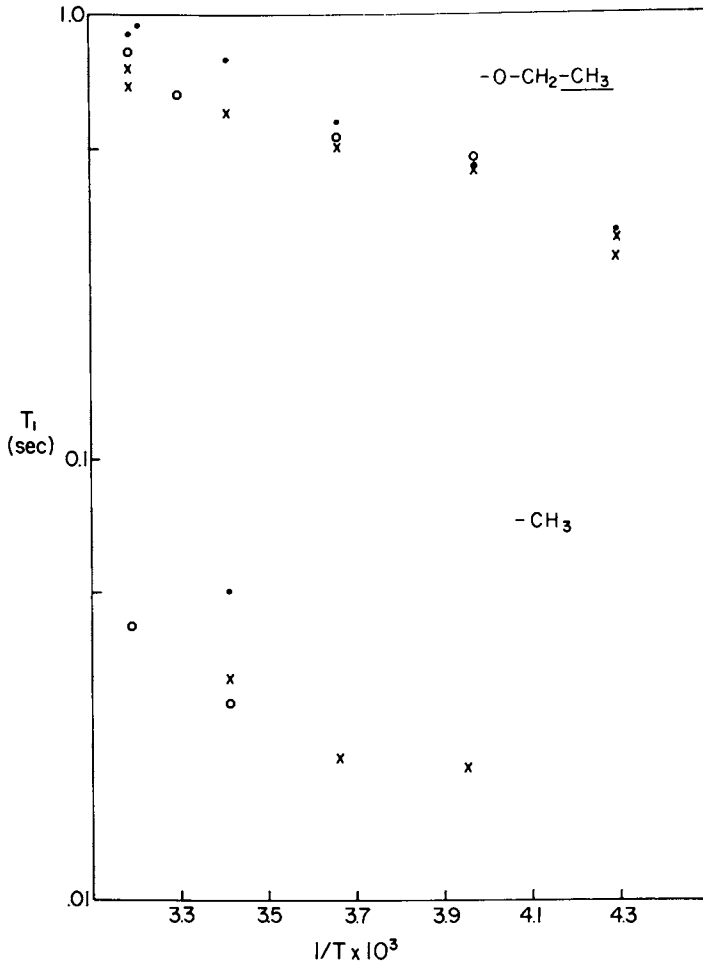


Figure 6. Temperature and concentration dependence of the methyl C-13 relaxation: (●), 10%; (×), 20%; (○), 30%.

results for the solvent. The ester methyl has a considerably larger T_1 than the ester methylene due to the internal rotation of the methyl. The α -CH₃ T_1 is more than an order of magnitude smaller than that of the ester methyl, as it reflects more closely the backbone motion and probably a higher barrier for internal rotation as well.³⁸

In Figure 7 is shown the solvent ¹H relaxation time as a function of temperature and polymer concentration. Analogous to the ¹³C results, the proton relaxation is affected by addition of only a small amount of polymer. Further addition of polymer decreases T_1 systematically.

The neat chloroform proton T_1 data are in substantial agreement with those of Bender and Zeidler.²⁷ Unlike chloroform ¹³C relaxation, which is almost exclusively by intramolecular dipole-dipole relaxation, ¹H relaxation has intermolecular as well as intramolecular contributions. By means of dilution with CDCl₃ one can obtain separately the intramolecular and the various intermolecular contributions.²⁷ Additional intermolecular terms must be included in the presence of polymer. Expressed in terms of relaxation rates,

$$(1/T_1) = (1/T_1)_{\text{intra}} + (1/T_1)_{\text{inter}} \quad (1)$$

where

$$(1/T_1)_{\text{intra}} = (1/T_1)_{\text{intra}}^{\text{spin rotation}} + (1/T_1)_{\text{intra}}^{\text{H-Cl}} \quad (2)$$

and

$$(1/T_1)_{\text{inter}} = (1/T_1)_{\text{inter}}^{\text{H-Cl}} + (1/T_1)_{\text{inter}}^{\text{H-HS}} + (1/T_1)_{\text{inter}}^{\text{H-P}} \quad (3)$$

The superscripts H-Cl refers to solvent proton-chlorine interaction, H-HS to solvent proton-proton interaction, and H-P to interaction of the solvent proton with the polymer. The latter should be dominated by solvent proton-polymer proton interaction. Upon addition of CDCl₃, followed by extrapolation to pure CDCl₃, the extrapolated rate $[(1/T_1)_0]$ is given by

$$(1/T_1)_0 = (1/T_1)_{\text{intra}} + (1/T_1)_{\text{inter}}^{\text{H-Cl}} + (1/T_1)_{\text{inter}}^{\text{H-D}} + (1/T_1)_{\text{inter}}^{\text{H-P}} \quad (4)$$

Equations 1-4, plus a knowledge³⁹ of the theoretical ratios

$$(1/T_1)_{\text{inter}}^{\text{H-D}} / (1/T_1)_{\text{inter}}^{\text{H-H}} \text{ and } (1/T_1)_{\text{inter}}^{\text{H-Cl}} / (1/T_1)_{\text{inter}}^{\text{H-H}}, \text{ allows the}$$

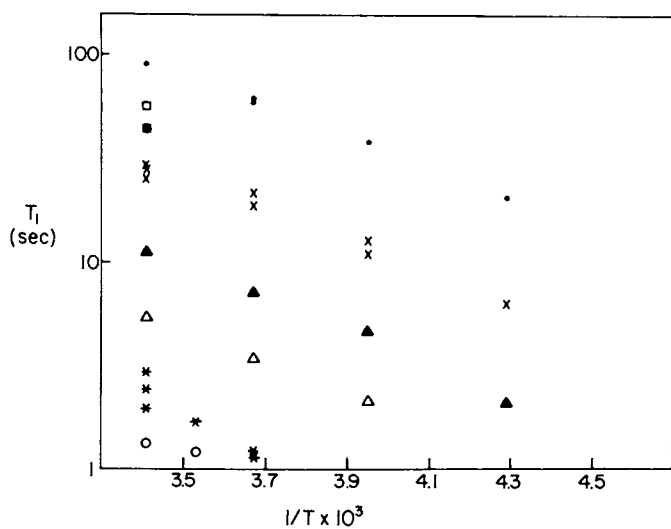


Figure 7. Temperature and concentration dependence of the chloroform ^1H relaxation: (●), 0%; (□), 3%; (■), 6%; (×), 10%; (▲), 20%; (△), 30%; (*), 40%; (○), 50%.

intramolecular and intermolecular contributions to T_1 to be assessed. In pure chloroform the intramolecular relaxation is found to be dominated by spin rotation.²⁷ The intermolecular contributions dominate the intramolecular at all temperatures, with the proton-proton interaction being the major contributor to the intermolecular relaxation mechanisms. In the presence of polymer the quantity $(1/T_1)_{\text{inter}}^{\text{H-P}} + (1/T_1)_{\text{intra}}$ is more easily determined from the experimental data than the proton-polymer interaction alone. Shown in Figure 8 is the solvent proton-solvent proton contribution as a function of polymer concentration. Shown in Figure 9 is the solvent proton-polymer contribution, where we have assumed the intramolecular contribution is small, and also scaled the results to reflect the number of polymer protons present.

Discussion

In a polymer solution the translational diffusion of solvent is well known to be related to the internal motion of the polymer, and its concentration dependence.⁴⁰⁻⁴² It would not be surprising to find a relationship between solvent rotational motion and polymer segmental motion, particularly at high polymer concentration. There is little systematic literature bearing on this point. Rothschild,⁴³ by analyzing the band shape of a far infrared band of CH_2Cl_2 , found only a small change in solvent rotational correlation time upon addition of up to 60% polystyrene. Anderson and Liu,¹⁶ studying the proton relaxation of benzene in the presence of PMMA, found a systematic decrease in T_1 with increasing polymer concentration. Addition of 35% PMMA resulted in a factor of three reduction in T_1 . Through the use of isotope dilution they were able to separate the relaxation into intramolecular and intermolecular contributions. Inasmuch as their analysis indicated that the intramolecular relaxation was independent of polymer concentration, they concluded that up to 35% PMMA had no effect on the rotational motion of benzene. Finally we turn to the work of Heatley and Scrivens,¹⁸ who measured the ^{13}C and ^1H relaxation in acetone at 0, 5, 10 and 20% PMMA. At high temperatures spin rotation dominated the relaxation and little information was obtainable concerning solvent motion. At low temperatures, where dipolar relaxation dominated, no effect of polymer on acetone rotation was evident up to 10% PMMA, but quite perceptible at 20% PMMA.

Returning to our data, it is especially interesting that we find a systematic reduction in solvent ^{13}C relaxation upon increase in polymer concentration, even at low polymer concentration, inasmuch as the relaxation should be almost exclusively by intramolecular dipolar relaxation.²⁸ The existence of a T_1 minimum at higher polymer concentration permits us to make a meaningful comparison with various models. If the chloroform ^{13}C relaxation

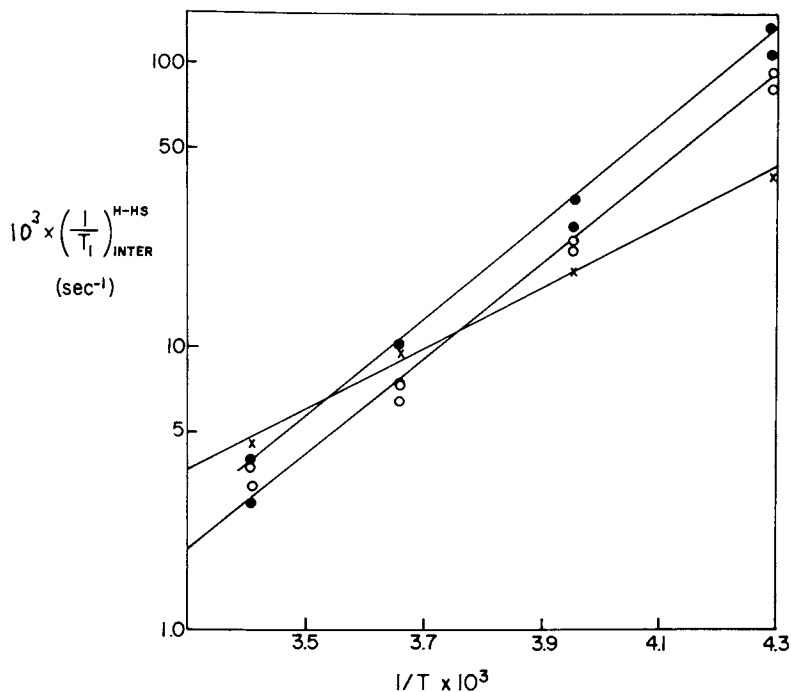


Figure 8. Temperature and concentration dependence of the solvent proton-solvent proton relaxation rate: (●), 20%; (○), 10%; (×), 0%.

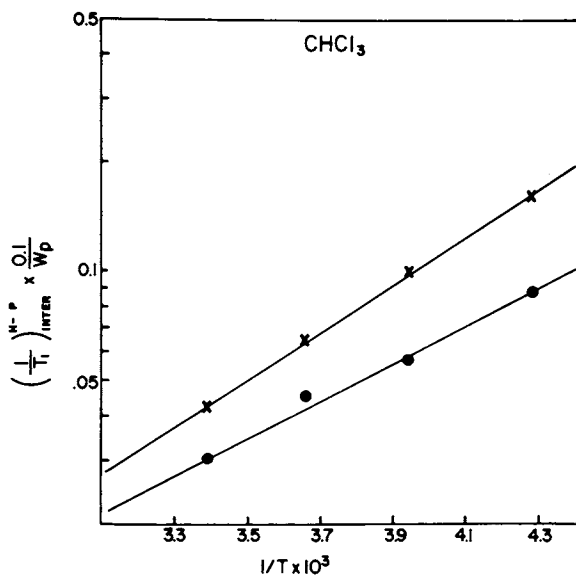


Figure 9. Temperature and concentration dependence of the solvent proton-polymer proton relaxation rate: (●), 10% polymer; (×), 20% polymer.

was describable by a single isotropic rotational correlation time, τ_c , the spectral densities $J_n(\omega)$ would be given by

$$J_n(\omega) = \tau_c / (1 + \omega^2 \tau_c^2), \quad (5)$$

leading to a T_1 minimum of 0.040 sec. The fact that the T_1 minimum is more than 4 times larger in itself rules out a single rotational correlation time. The next logical step is to assume a distribution of correlation times, as it will raise the T_1 minimum. The appropriate spectral densities for both symmetric and asymmetric distributions of isotropic rotations have been given^{10,44}, wherein the relaxations are characterized by a mean correlation time and a distribution width parameter. Keeping the width parameter and type of distribution function fixed, T_1 as a function of mean correlation time may be calculated. Assuming, for instance, the symmetric Cole-Cole distribution, a width parameter of 0.25 will give a T_1 minimum in agreement with the 30% PEMA data. However, upon keeping the width parameter constant, a mean correlation time of less than 10^{-15} sec. would be required to yield a calculated T_1 in agreement with the 20°C data. This is clearly a meaningless value as solvent rotational correlation times are never that small. As an example, for pure deuteriochloroform at 20°C, Huntress²⁹ calculates τ_{\perp} to be 1.8×10^{-12} sec. and τ_{\parallel} to be 0.92×10^{-12} sec. It is highly unlikely that the rotational diffusion of the solvent in a polymer solution could possibly exceed that of the pure solvent. Additionally the calculated η corresponding to this mean τ and distribution width is much lower than the experimental value at 20°C of nearly 2. Unsymmetric distributions give unsatisfactory results also. The assumption of isotropic motion is not a significant source of error, as chloroform rotation diffusion coefficients differ by less than a factor of two.²⁹ Relaxation by scalar coupling and spin rotation is small in pure chloroform²⁸, and we can see little reason why it should be more significant in the presence of polymer. Hence another mechanism must be devised to explain the experimental results.

A possible step in this direction can be made through use of earlier relaxation studies on other systems. Hunt and Powles,⁴⁵ when studying the proton relaxation in liquids and glasses, found the relaxation best described by a "defect-diffusion" model, in which a non-exponential correlation function corresponding to diffusion is included together with the usual exponential function corresponding to rotational motion. The correlation function is taken as the product of the two independent reorientation processes. This type of model has more recently been applied to ^{13}C relaxation in polymers,¹⁰ where the nonexponential part corresponds to conformational jumps in a Monnerie type⁴⁶ local mode

mechanism plus an exponential contribution corresponding to over-all tumbling. A continuum version of this has recently been given.⁴⁷ This can at least qualitatively explain the experimental data in that the T_1 minimum can be significantly increased over a single exponential model while keeping the nuclear Overhauser enhancement factor much closer to the experimental values, while at the same time not leading to physically absurd values for the exponential reorientation. One can give a physical picture to this scheme which makes some intuitive sense. In the presence of polymer the solvent is relaxed by two mechanisms. One is the rotation of the solvent, an exponential reorientation contribution. The nonexponential or diffusive contribution could result from the reorientation when the solvent drops into the hole resulting from the conformational jump of the polymer. This would be compatible with the mechanisms used to explain the relationship between solvent translational diffusion and polymer segment motion, as mentioned earlier. The solvent proton results can also be analyzed in terms of this mechanism, as well as the polymer backbone motion. We are currently trying to quantitatively analyze the data from this viewpoint.

References

1. K. J. Liu and R. Ullman, J. Chem. Phys., **48**, 1158 (1968).
2. K. J. Liu and W. Burlant, J. Polym. Sci., Part A-1, **5**, 1407 (1967).
3. K. Hatada, Y. Okamoto, K. Ohta, and H. Yuki, J. Polym. Sci., Polym. Letters Ed., **14**, 51 (1976).
4. F. Heatley and M. K. Cox, Polymer, **18**, 225 (1977).
5. D. Ghesquiere, B. Ban, and C. Chachaty, Macromolecules, **10**, 743 (1977).
6. J. Spevacek and B. Schneider, Polymer, **19**, 63 (1978).
7. D. Daskocilova, B. Schneider, and J. Jakes, J. Mag. Res., **29**, 79 (1978).
8. J. Schaefer, Macromolecules, **6**, 882 (1973).
9. J. R. Lyerla and T. T. Horikawa, J. Polym. Sci., Polym. Letters Ed., **14**, 641 (1976).
10. F. Heatley and A. Begum, Polymer, **17**, 399 (1976).
11. F. A. Bovey, F. C. Schilling, T. K. Kwei and H. L. Frisch, Macromolecules, **10**, 559 (1977).
12. K. Hatada, T. Kitayama, Y. Okamoto, K. Ohta, Y. Umemura and H. Yuki, Makromol. Chem., **178**, 617 (1977).
13. R. E. Cais and F. A. Bovey, Macromolecules, **10**, 752 (1977).
14. R. E. Cais and F. A. Bovey, Macromolecules, **10**, 757 (1977).

15. W. H. Stockmayer, A. A. Jones and T. L. Treadwell, Macromolecules, 10, 762 (1977).
16. J. E. Anderson and K. J. Liu, J. Chem. Phys., 49, 2850 (1968).
17. K. Sato and A. Nishioka, J. Polym. Sci. Part A-2, 10, 489 (1972).
18. F. Heatley and J. H. Scrivens, Polymer, 16, 489 (1975).
19. W. P. Slichter and D. D. Davis, Macromolecules, 1, 47 (1968).
20. F. Heatley, Polymer, 16, 493 (1975).
21. T. Cosgrove and R. F. Warren, Polymer, 18, 255 (1977).
22. R. Kimmich and Kh. Schmauder, Polymer, 18, 239 (1977).
23. G. Bowers and A. Rigamonti, J. Chem. Phys., 42, 171 (1965).
24. J. Schaefer, in "Structural Studies of Macromolecules by Spectroscopic Methods", K. J. Irvin, Ed., John Wiley & Sons, 1976, pp 201-226.
25. A. K. Karim and D. C. Bonner, Soc. Plast. Eng., Tech. Pap., 22, 3 (1976).
26. D. W. McCall, Natl. Bur. Stand. (U.S.) Spec. Publ. No. 301, 475 (1969).
27. H. J. Bender and M. D. Zeidler, Ber. Bunsenges Physik. Chem., 75, 236 (1971).
28. R. Shoup and T. Farrar, J. Mag. Res., 7, 48 (1972).
29. W. T. Huntress, J. Phys. Chem., 73, 103 (1969).
30. D. E. O'Reilly and G. E. Schacher, J. Chem. Phys., 39, 1768 (1963).
31. G. Bowers and A. Rigamonti, J. Chem. Phys., 42, 175 (1965).
32. K. Gillen, M. Schwartz and J. Noggle, Mol. Phys., 20, 899 (1971).
33. S. Mallikarjun and N. E. Hill, Trans. Faraday Soc., 61, 1389 (1965).
34. S. Mallikarjun and N. E. Hill, Rheologica Acta, 5, 232 (1966).
35. J. Brandrup and H. Immergut, Polymer Handbook, 2nd ed., Interscience, New York, 1975.
36. R. Freeman and H. D. W. Hill, J. Chem. Phys., 54, 3367 (1971).
37. J. L. Markley, W. J. Horsley, and M. P. Klein, J. Chem. Phys., 55, 3604 (1971).
38. J. R. Lyerla, T. T. Horikawa and D. E. Johnson, J. Am. Chem. Soc., 99, 2466 (1977).

39. A. Abragam, *The Principles of Nuclear Magnetism*, Oxford (1961).
40. R. S. Moore and J. D. Ferry, *J. Phys. Chem.*, 66, 2699 (1962).
41. C. P. Wong, J. L. Schrag, and J. D. Ferry, *J. Polym. Sci. Part A-2*, 8, 991 (1970).
42. J. D. Ferry, "Viscoelastic Properties of Polymers," Wiley, New York (1970).
43. W. G. Rothschild, *Macromolecules*, 1, 43 (1968).
44. J. Schaefer, *Macromolecules*, 6, 882 (1973).
45. B. I. Hunt and J. G. Powles, *Proc. Phys. Soc.*, 88, 513 (1966).
46. B. Valur, J. P. Jarry, F. Geny and L. Monnerie, *J. Polym. Sci., Polym. Phys. Ed.* 13, 667, 675 (1975).
47. J. T. Bendler and R. Yaris, *Macromolecules*, 11, 650 (1978).

RECEIVED March 13, 1979.

The Use of Carbon-13 NMR to Study Binding of Hormones to Model Receptor Membranes: The Opioid Peptide Enkephalin

HAROLD C. JARRELL, P. TANCRÉDE, ROXANNE DESLAURIERS,
and IAN C. P. SMITH

Division of Biological Sciences, National Research Council of Canada,
Ottawa, Ontario, Canada K1A 0R6

W. HERBERT MCGREGOR

Wyeth Laboratories, P. O. Box 8299, Philadelphia, PA 19101

The recent discovery of a class of peptides, the enkephalins, which act as opiate agonists has led to a number of physical chemical studies aimed at understanding the structure-activity relationships between the enkephalins and the opiates (1-9). The ^1H spin-spin coupling constants [2-4] of the enkephalins in solution can be interpreted in terms of folded conformations resembling that of morphine in the placement of the residues which appear important for biological activity. X-ray crystallography and theoretical calculations (4-9) have also shown that methionine and leucine enkephalin adopt conformations similar to those concluded from ^1H NMR studies. Hence it would appear that opioid peptides can topographically resemble the opiates by assuming preferred, folded, conformations. However, earlier studies from this laboratory (10) have shown that ^{13}C NMR data can be interpreted in terms of a conformationally flexible structure for methionine enkephalin.

Aboud and coworkers (11-13) have reported that acidic lipids, in particular phosphatidyl serine (PS), enhanced binding of opiates to membranous components of brain. Furthermore, PS has been shown to bind opiates stereospecifically whereas other acidic lipids bind to a lesser extent and neutral lipids bind very little (11). Loh and coworkers (14-16) have reported that cerebroside

*Issued as N.R.C.C. Publication No. 17068.

0-8412-0505-1/79/47-103-159\$05.50/0
© 1979 American Chemical Society

sulfate exhibits strong binding of opiates and that the cerebroside sulfate-opiate complex resembles an opiate complex which had been isolated from brain components. Lipids, therefore, have been implicated in the interaction of opiates with their receptors.

It is known (17) that opiates and opioid peptides may exhibit specific receptor binding (high affinity) and nonspecific (low affinity) binding. Phosphatidyl serine and other lipids exhibit weak binding of opiates and therefore serve as models for non-specific binding. Preliminary to a study of high affinity binding we chose to study the interaction of methionine enkephalin to various lipids, in particular PS, as a model for binding. In these studies we have followed a number of ^{13}C NMR parameters for the glycine-2 and glycine-3 residues of methionine enkephalin. The glycyI residues were 90% enriched in ^{13}C at the α -carbons. This allowed us to measure chemical shifts (δ), line widths ($\Delta\nu$), and spin-lattice relaxation times (T_1) for the α -carbons of the glycine residues as a function of various parameters. It is generally believed that the binding is electrostatic in nature (11,15); we therefore sought a pH dependence of the binding of enkephalin to the lipids. We further sought competition between morphine and enkephalin for binding to PS.

Materials and Methods

[2-[2- ^{13}C]glycine] and [3-[2- ^{13}C]glycine] methionine enkephalin (TyrGlyGlyPheMet), enriched to 90% in ^{13}C , were prepared as described previously (10). Phosphatidyl serine (PS) was obtained from Serdary Research Laboratories, London, Canada or Lipid Products, South Nutfield, England. Egg lecithin was obtained from Lipid Products, South Nutfield, England. Cerebroside sulfate (sulfatides) was obtained from Analabs, Inc. Morphine sulfate pentahydrate was purchased from Ingram and Bell Limited, Don Mills (Toronto), Canada. Samples for binding studies were prepared as described previously (10).

NMR measurements were performed on samples prepared in

deuterium oxide at 30°C. pH values are meter readings that have not been corrected for the deuterium isotope effect. The pH of samples was varied using hydrochloric acid and sodium hydroxide diluted in deuterium oxide.

^{13}C NMR measurements were performed on Varian CFT-20 and XL-100 spectrometers under conditions which have been described previously (10). ^{13}C chemical shifts are reported with respect to external tetramethylsilane (TMS). Dioxane was added to the samples as an internal standard for linewidth and chemical shift measurements.

Results and Discussion

Three parameters are readily obtainable from ^{13}C NMR spectra which may be useful in studying binding interactions: the chemical shift (δ), the linewidth ($\Delta\nu$) or the apparent or effective spin-spin relaxation time (T_2^*), and the spin-lattice relaxation time (T_1). ^{13}C chemical shifts can reflect steric strain and change in the electronic environment within a molecule when it binds to another species. Spin-lattice and spin-spin relaxation times can yield information on the lifetimes, sizes and conformations of molecular complexes.

Studies on morphine binding to PS have shown the interaction to have a maximum at pH 8-9 (11). In order to interpret the pH-dependence of binding it is necessary to determine the dissociation constants (pKa) of the titratable groups in each of the interacting species. For PS pKa values of 3.7, 4.0 and 7.5 have been determined for the phosphate, carboxyl, and amino groups, respectively (18). Figure 1 shows the ^{13}C chemical shift of the tyrosyl β -carbon as a function of the pH-meter reading of a solution of methionine enkephalin in deuterium oxide (D_2O). The pKa (measured in D_2O) was determined to be 7.5 for the amino group of the tyrosine residue while the methionine carboxyl group had a pK of 3.9.

The pH-dependence of the chemical shifts measured for the

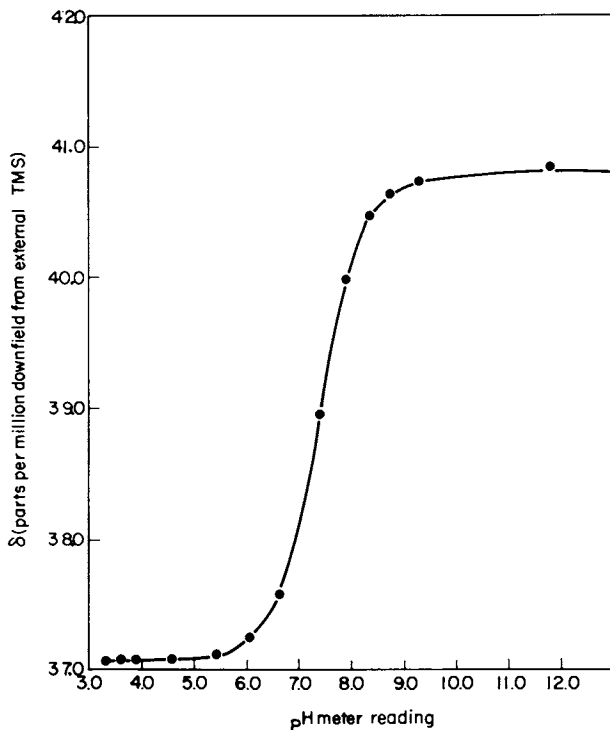


Figure 1. Plot of C-13 chemical shift of C- β of tyrosine in enkephalin as a function of pH meter reading in D₂O, 30°C

enriched gly-2 and gly-3 residues of methionine enkephalin in the absence and presence of PS are shown in Figures 2 and 3, respectively. It is evident that the ^{13}C chemical shifts of the glycyI residues were not greatly affected by the presence of lipid. The observed linewidths (corrected for field inhomogeneity) for the gly-2 and gly-3 residues of methionine enkephalin in the presence of PS were also measured as a function of pH; the results are given in Table I. The observed linewidths vary only slightly over the reported pH range. The above results indicate that neither chemical shifts nor linewidths (or T_2^*) would be useful for monitoring interaction of enkephalin with PS - at least via the glycyI residues.

Figures 4 and 5 show a pH-dependence of the T_1 values of the glycyI residues in [2-[2- ^{13}C]glycine] and [3-[2- ^{13}C]glycine] methionine enkephalin (1.1 mg) respectively, in the presence of PS (75 mg). An earlier study (10) showed that the T_1 values of the glycyI units of enkephalin free in solution did not change over the pH range 1.5-10.0. In the presence of PS, we observe a dramatic decrease in T_1 from that of free enkephalin at "pH" 10, to values close to 60 msec around "pH" 4, for the gly-3 residue. This decrease in T_1 values as a function of pH is reversed at "pH" 1.0, where the T_1 values again reflect those of free enkephalin. At "pH" values less than 4.0 the intensity of the glycyI resonances was greatly reduced from that of the glycyI resonances of free enkephalin and hence a mapping of the pH-dependence of T_1 between "pH" 4.0 and 1.0 was not possible. The above pH-dependence would seem to indicate an ionic-type of interaction between enkephalin and PS. The observed pH-dependence of binding is consistent with interaction between the NH_3^+ group of tyrosine in enkephalin and the CO_2^- group of serine in PS, analogous to the earlier model for morphine binding (11). A maximum interaction would be expected when the amino group of tyrosine is fully protonated and the carboxyl group of serine is unprotonated. Thus at pH 10.0 or 1.0 little effect on the T_1 values would be expected. It should

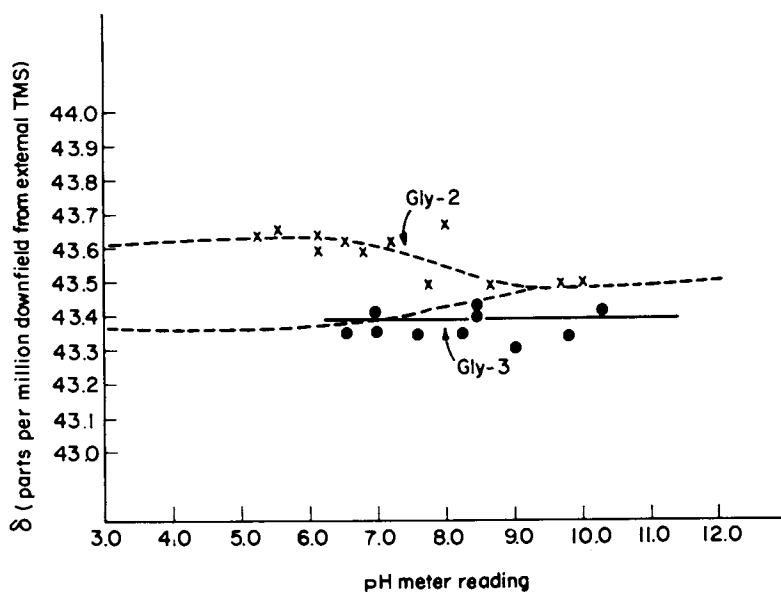


Figure 2. C-13 chemical shifts of the glycylic residues in [2-[2-C-13]glycine]methionine enkephalin and [3-[2-C-13]glycine]methionine enkephalin in the presence of 75.0 and 78.5 mg of PS, respectively, as a function of pH, 30°C. Shifts observed for enkephalin in the absence of PS (see Figure 3), (---).

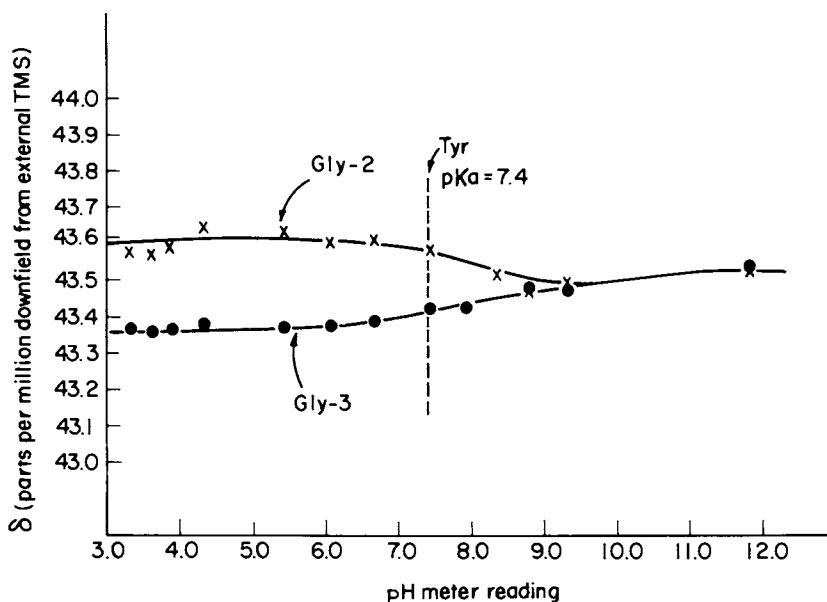


Figure 3. ^{13}C chemical shifts of the glycyl-2 and glycyl-3 residues of methionine enkephalin as a function of pH, 30°C

Table I. Linewidths of [2-[2-¹³C]glycine] and [3-[2-¹³C]glycine]Methionine Enkephalin in the Presence of PS at Various "pH"'s

"pH"	$\Delta\nu^a$ (Hz)	"pH"	$\Delta\nu^b$ (Hz)
5.0	5.4	6.5	5.0
5.5	5.4	6.9	2.5
6.1	3.2	7.6	2.7
6.5	3.1	8.0	3.3
7.5	3.0	8.6	1.2
8.1	3.0	9.8	2.0

^aLinewidth for [2-[2-¹³C]glycine] methionine enkephalin (1.1 mg) in the presence of PS (75 mg) in D₂O (1 ml) and corrected for field inhomogeneity.

^bLinewidth for [3-[2-¹³C]glycine] methionine enkephalin under the same conditions as in (a).

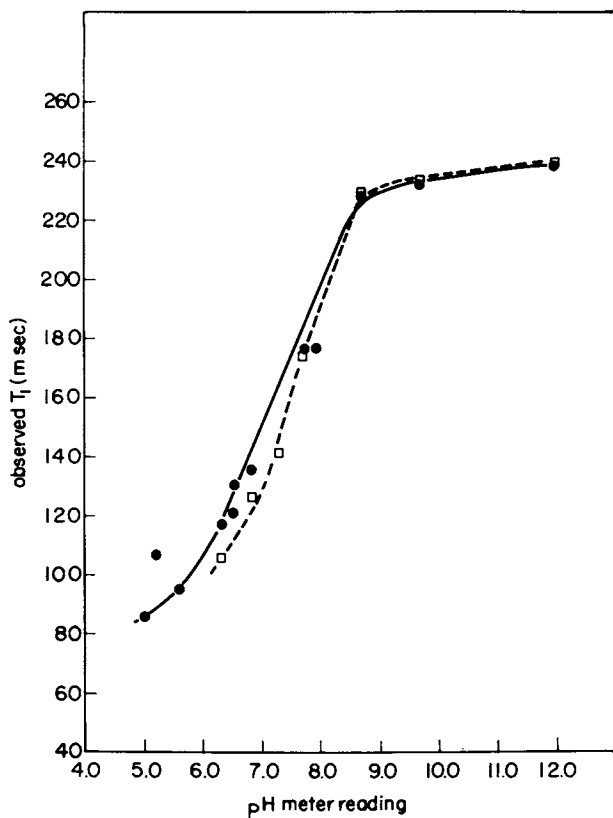


Figure 4. The pH dependence of T_1 of [2-[2-C-13]glycine]methionine enkephalin (1.1 mg) in presence of phosphatidyl serine [8.5 mg (□) and 75 mg (●)]. Spectra were run at 20 MHz, 30°C.

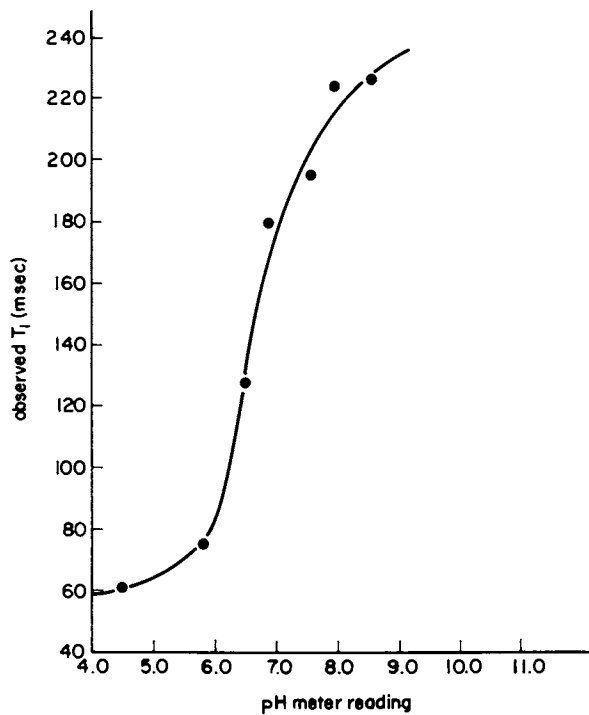


Figure 5. The pH dependence of T_1 of [3-[2-C-13]glycine]methionine enkephalin (1.1 mg) in presence of phosphatidyl serine (78.5 mg). Spectra were run at 20 MHz.

be noted that due to the very similar pKa's of the amino groups of PS and enkephalin, the phosphate and carboxyl group of PS, and the carboxyl group of enkephalin, we cannot determine which groups are specifically involved in binding using the present method.

Rotational correlation times obtained from the observed ^{13}C T_1 values of the glycyI residues in methionine enkephalin as a function of "pH" are given in Figure 6. The 3-glycyI residue shows a longer correlation time than does the 2-glycyI moiety at low pH. Although binding is thought to occur via the NH_3^+ group of the tyrosyl residue, the 2-glycyI unit is known (10) to undergo a greater degree of segmental motion in free solution and this could contribute to the shorter observed correlation time at low pH.

Dissociation Constants. Having established that T_1 values could be used to monitor the enkephalin-PS interaction, we sought to obtain a quantitative estimate of the strength of this interaction. If we assume that we are dealing with a 1:1 peptide-lipid complex, the interaction may be represented as in equation 1,



where P and L are methionine enkephalin and lipid, respectively, and PL is the peptide-lipid complex. The dissociation constant K_d , which reflects the strength of the interaction, is therefore given by equation 2

$$K_d = \frac{[\text{P}_F][\text{L}_F]}{[\text{PL}]} \quad (2)$$

where $[\text{P}_F]$ and $[\text{L}_F]$ are the concentrations of free enkephalin and free lipid, respectively, and $[\text{PL}]$ is the concentration of the peptide-lipid complex. In the case of fast exchange it can be shown (19) that the observed relaxation time $T_{1,\text{obs}}$ can be related to the mole fraction of the bound species by equation 3

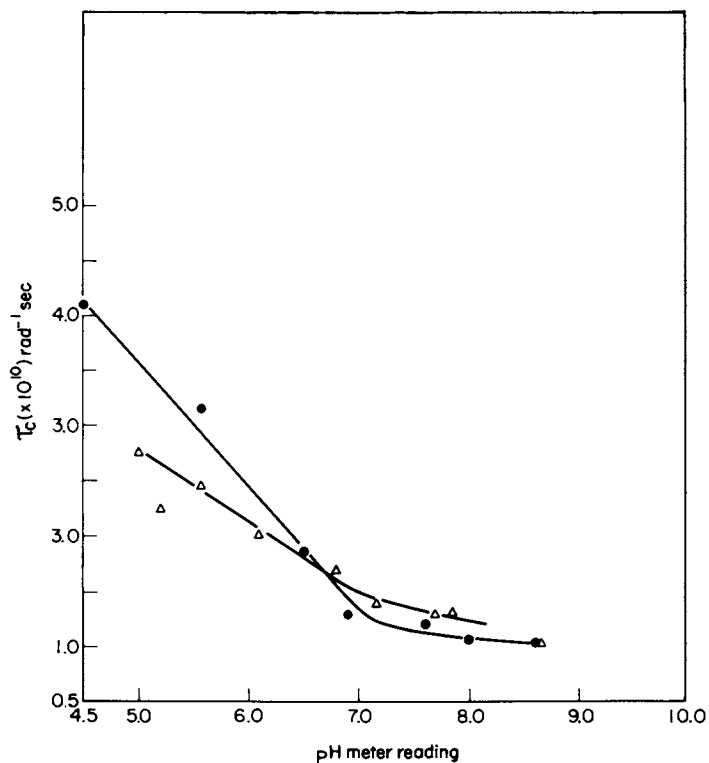


Figure 6. Plot of correlation times corresponding to observed T_1 values of [2-[2-C-13]glycine]methionine enkephalin (\times) and of [3-[2-C-13]glycine]methionine enkephalin (\circ) in presence of PS as a function of pH (1.1 mg of peptide in the presence of ca. 75 mg PS), 30°C

$$\frac{1}{T_{1,obs}} = \frac{1}{T_{1,F}} + \frac{f_B}{T_{1,B}} \quad (3)$$

where $T_{1,F}$ and $T_{1,B}$ are the spin-lattice relaxation times of the free and bound species and f_B is the mole fraction of the bound species. For an interaction represented by equation 1 it can be shown that the mole fraction bound can be represented by equation 4

$$f_B = \frac{[L_T]}{K_d + [L_T] + [P_T]} \quad (4)$$

where $[L_T]$ and $[P_T]$ are the total concentrations of lipid and enkephalin, respectively. Substituting equation 4 into equation 3 gives:

$$\frac{1}{T_{1,obs}} = \frac{1}{T_{1,F}} + \frac{1}{T_{1,B}} \frac{[L_T]}{K_d + [L_T] + [P_T]} \quad (5)$$

Equation 5 can be rearranged into a more useful form,

$$\frac{T_{1,obs} T_{1,F}}{T_{1,F} - T_{1,obs}} = \frac{T_{1,B} [K_d + [P_T]]}{[L_T]} + T_{1,B} \quad (6)$$

Thus, a plot of $[T_{1,obs} T_{1,F} / (T_{1,F} - T_{1,obs})]$ vs. $1/[L_T]$ should give $T_{1,B}$ as the intercept and $T_{1,B}(K_d + [P_T])$ as the slope.

We have determined K_d at "pH" 6.3 for $[2\text{-}[2\text{-}^{13}\text{C}]\text{glycine}]$ and $[3\text{-}[2\text{-}^{13}\text{C}]\text{glycine}]$ methionine enkephalin by varying the lipid concentration in the presence of a 1.2 mg/ml sample of enkephalin; Figures 7 and 8 represent plots of $(T_{1,obs} T_{1,F} / (T_{1,F} - T_{1,obs}))$ vs. $1/[L_T]$. These plots yield K_d values of 5.4×10^{-1} M and 3.3×10^{-1} M, respectively, and a $T_{1,B}$ value of 35 msec in both cases. An estimate of the sensitivity of the value determined for $T_{1,B}$ and K_d was obtained by varying $T_{1,F}$ by ± 10 msec (within the expected experimental error (10)). This resulted in a change of ± 20 msec for $T_{1,B}$ and a value of K_d which differed by a factor of three.

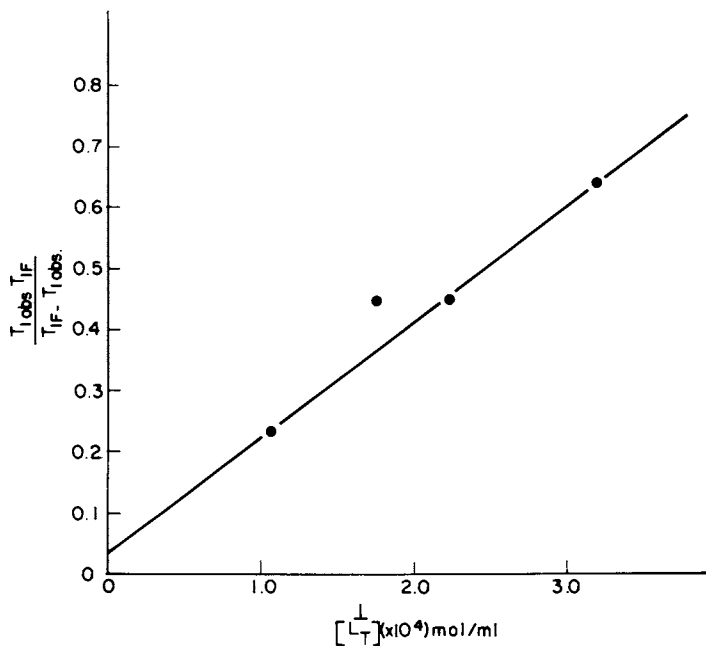


Figure 7. Plot of $(T_{1\text{obs}} T_{1F}) / (T_{1\text{obs}} - T_{1F})$ vs. $1/[L_T]$ for [2-[2-C-13]glycine]-methionine enkephalin. Spectra were obtained using 1.1 mg of enkephalin at 30°C on a CFT-20 spectrometer operating at 20 MHz.

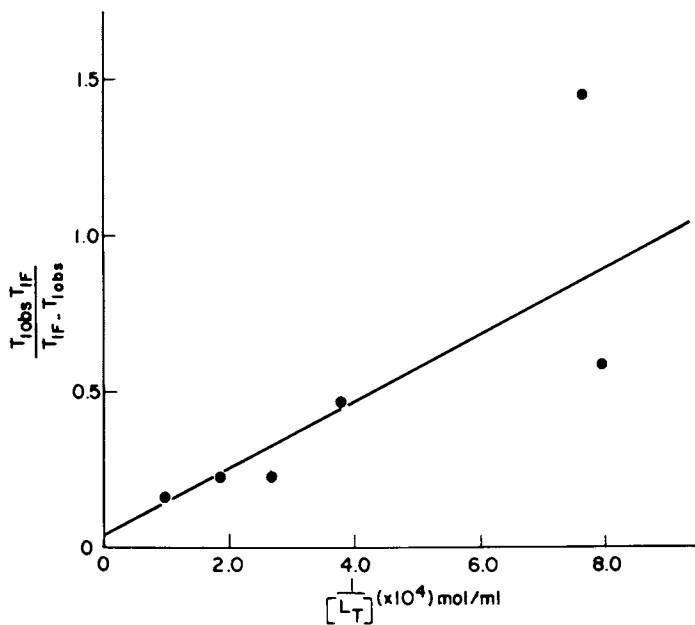
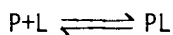


Figure 8. A plot similar to that in Figure 7 for [3-[2-C-13]glycine]methionine enkephalin

Using the $T_{1,B}$ obtained above and the pH-dependence of the $T_{1,obs}$ measured for [2-[2- ^{13}C]glycine] methionine enkephalin (see Figure 4), a pH-dependence of K_d was calculated and is shown in Figure 9. It is readily seen that K_d varies by a factor of five from 3×10^{-1} M to 1.6 M. The variation of K_d with pH may be explained by noting that equation 1 assumes that only one ionic form for both enkephalin and PS is involved in binding. However, the interaction is better represented (for $pH \geq 6.3$) by



where L^1 is PS in the NH_2 form, PL^1 is the PS-enkephalin complex involving PS in the NH_2 form, P is enkephalin in its zwitterionic form and L is PS as its NH_3^+ form. It can be shown (20) that the observed K_d is related to the true dissociation constant K_d^1 by

$$K_d = K_d^1 A$$

where A involves the pK_a 's of enkephalin, PS and the lipid-enkephalin complex. Again the pH-dependence of the observed dissociation constant indicates that the enkephalin-PS interaction is electrostatic in nature.

It is informative to compare the observed dissociation constant for the enkephalin-PS interaction with those reported for other systems. Methionine enkephalin has been shown (21) to bind to brain membrane with a high affinity binding constant of $1.7 \times 10^8 M^{-1}$ and a low affinity binding constant of $1.7 \times 10^6 M^{-1}$. PS-morphine interaction has been determined to have a K_d of $\sim 10^{-6}$ M (11). The results observed in this study suggest that the enkephalin-PS binding is weaker than the interaction of morphine with PS.

Correlation Times of the Bound Species. In order to interpret the values of $T_{1,B}$ it is essential to know the rotational correlation time for a particle bound rigidly to a vesicle. An

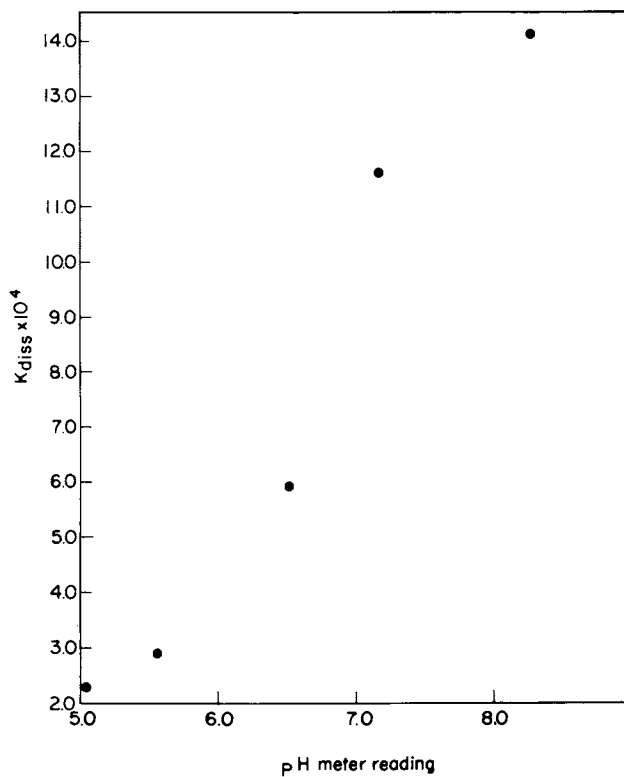


Figure 9. A plot of K_d vs. pH for [2-[2-C-13]glycine]methionine enkephalin

estimate of the rotational correlation time for a vesicle can be determined from the Stokes-Einstein relation

$$\tau = \frac{4\pi r^3 \eta}{3kT} = \frac{V\eta}{kT} \quad (7)$$

where η is the viscosity of the solution at a temperature T ($^{\circ}\text{K}$), k is the Boltzman constant, r the radius of the spherical particle, and V the volume of the particle. Vesicles of egg lecithin have been shown to have radii of 105-118 \AA (22,23). Using equation 7 with the viscosity $\eta = 0.7808$ cP of pure water, we can estimate τ , the rotational correlation time of the vesicle to be ca 1.0×10^{-6} sec rad^{-1} . A small particle rigidly bound to a vesicle should have a similar rotational correlation time. We have estimated $T_{1,B}$ to be 35 msec at 20 MHz, which corresponds to possible rotational correlation times of 7.6×10^{-10} sec rad^{-1} and 3.0×10^{-8} sec rad^{-1} (24). The corresponding linewidths for the bound species would be 9 Hz and 86 Hz. Thus, it would seem that when enkephalin binds to the lipid vesicle internal motion (possibly segmental motion) still occurs in the peptide backbone at the level of glycy1-2 and glycy1-3 residues. This is not unexpected if the binding involves mainly the NH_3^+ group of the tyrosyl unit; this may be termed a one point binding rather than the possible two point interaction which would involve both the NH_3^+ and CO_2^- group in enkephalin. It would however appear that the internal motion of the peptide is considerably decreased on binding from the value of 7.0×10^{-11} sec rad^{-1} which has been determined for the glycy1-2 residue in solution (10). This is because fast motions dominate T_1 behavior and τ_{int} values of the order of 10^{-11} sec rad^{-1} would result in considerably longer (hundreds of milliseconds) observed $T_{1,B}$ values (23). The observed $T_{1,B}$ is consistent with a correlation time for internal motion no smaller than $5.0-7.0 \times 10^{-10}$ sec rad^{-1} if the correlation time of the vesicle is 1.0×10^{-6} sec rad^{-1} (25). Thus, it is possible to estimate the rate of internal motion of a

peptide by decreasing the rate of overall motion of the particle to the extent that the internal motion dominates the spin-lattice relaxation behaviour.

Linewidths. Observed linewidths can sometimes be useful in determining which possible value of τ , determined from T_1 , corresponds to the experimental situation. In the present case, our τ values for the bound species should give linewidths of 9 or 86 Hz. Knowing K_d , and the linewidth of free enkephalin, we can calculate an expected "observed" linewidth for all of our experimental situations. Inspection of Table I reveals that no linewidth greater than 7 Hz was observed. As an example of the linewidth to be expected for conditions of maximal binding to PS, we choose the case of [2-[2- 13 C]glycine] methionine enkephalin (1.1 mg) in the presence of PS (75 mg) in D_2O (1 ml) at "pH" 5.0. The measured T_1 was 86 msec and the observed corrected linewidth was 7 Hz. Equation 8 can be derived

$$\Delta\nu_{\text{obs}} = \Delta\nu_F + \Delta\nu_B \frac{[L_T]}{K_d + [L_T] + [P_T]} \quad (8)$$

in the same manner as equation 5, where $\Delta\nu_{\text{obs}}$ is the observed linewidth, $\Delta\nu_F$ and $\Delta\nu_B$ are the linewidths of the free and bound species; $\Delta\nu_F$ is 2 Hz and $\Delta\nu_B$ corresponds to the two possible τ values for the observed $T_{1,B}$ (see above). Using a K_d of 5×10^{-1} M the linewidths predicted by equation 8 for the two possible correlation times of the bound species are 3.5 and 17 Hz. Similarly using a K_d of 3.0×10^{-1} M linewidths of 2.3 Hz and 4.4 Hz are predicted. Thus, it would seem that the shorter τ value for the bound peptide would more accurately predict the observed linewidths.

Effect of Ionic Strength. Ionic strength has been shown to be an important factor in the interaction of opiates with PS (11), high ionic strength decreasing the interaction. Under conditions of maximal interaction ("pH" 4.0), addition of 1M sodium chloride to an enkephalin-PS mixture (1.0 mg)-(20 mg) caused the spectrum

to revert to that of free enkephalin. The high salt concentration rendered formation of vesicles impossible and spectra were obtained with the lipid simply dispersed in the mixture by sonication. A lower concentration of sodium chloride (10^{-3} M) had no observable effect on the enkephalin-PS interaction.

Morphine Antagonism. In order to verify that morphine competes with enkephalin for binding to PS we added morphine sulfate pentahydrate (10 mg) to a mixture of [2-[2-¹³C]glycine]enkephalin (1.2 mg) and PS (20 mg) in D₂O at "pH" 6.3. Under this condition, the linewidth and intensity of the glycy1-2 resonance reverted to those of free enkephalin. Under similar conditions, but with PS (30 mg) and morphine sulfate pentahydrate (5.4 mg), the linewidth and intensity of the glycy1-2 resonance was again essentially that of free enkephalin.

Binding to Other Lipids. No interaction of [2-[2-¹³C]glycine]methionine enkephalin with egg lecithin was observed at pH 3.5 and 7.0 as evidenced by no significant difference between the observed linewidths and T₁'s and those of free enkephalin.

Although cerebroside sulfate has been reported to interact strongly with opiates (14-16), enkephalin has so far shown no significant binding in the present type of experiment to an egg lecithin (27 mg) - sulfatide (25 mg) mixture at pH 6.3 and 7.1. Further studies are currently underway with this system.

Conclusions

The present study has shown that enkephalin binds to PS in a pH-dependent fashion. The binding most likely involves the NH₃⁺ group of the tyrosine residue of enkephalin and the CO₂⁻ group of PS. We have measured K_d for the interaction and find it to be of the order of 5×10^{-1} M ("pH" 6.3) which is much weaker than the interaction of PS with morphine derivatives. Upon binding of enkephalin to PS the correlation time for internal motion in the backbone of the peptide increases by at least one order of magnitude (from 7.0×10^{-11} sec rad⁻¹). The T₁ of the bound peptide

has been estimated to be 35 msec (monitored for both the glycy1-2 and glycy1-3 residues). The binding exhibits the expected morphine competition. At present no evidence of binding of enkephalin to egg lecithin or cerebroside sulfate has been obtained.

We believe this study to be preliminary to the full mapping of the enkephalin interaction with PS and to the investigation of enkephalin in high affinity binding. Future studies should involve [4-[2-¹³C]phenylalanine]methionine enkephalin and [2-[2-¹³C]glycine]methionine enkephalinamide, which are in preparation. These studies will allow more accurate determination of the rates of overall and internal motion in enkephalin and help elucidate further the nature of enkephalin-lipid interactions.

References

1. Roques, B.P., Garbay-Jaureguiberry, C., Oberlin, R., Anteunis, M. and Lala, A.K., *Nature*, (1976) *262*, 778.
2. Combrisson, S., Roques, B.P. and Oberlin, R., *Tetrahedron Lett.*, (1976), 3455.
3. Bleich, H.E., Day, A.R., Freer, R.J. and Glasel, J.A., *Biochem. Biophys. Res. Commun.*, (1977) *74*, 592.
4. Fournié-Zaluski, M., Prange, T., Pascard, C. and Roques, B.P., *ibid*, (1977), *79*, 1199.
5. Gorin, F.A. and Marshall, G.R., *Proc. Natl. Acad. Sci., USA*, (1977) *74*, 5179-5183.
6. Smith, G.D. and Griffin, J.F., *Science*, (1978), *199*, 1214.
7. Baladis, Y.Y., Nikiforovich, G.V., Grinsteine, I.V., Vegner, R.E., and Chipens, G.I., *FEBS Lett.*, (1978), *86*, 239.
8. Loew, G.H. and Burt, S.K., *Proc. Natl. Acad. Sci., USA*, (1978), *75*, 7.
9. Humblet, C., DeCoen, J.L. and Koch, M.H.J., *Arch. Int. Physiol. Biochim.*, (1977), *85*, 415.

10. Tancrède, P., Deslauriers, R., McGregor, W., Ralston, E., Sarantakis, D., Somorjai, R.L. and Smith, I.C.P., *Biochem.*, (1978), *17*, 2905.
11. Abood, L.G. and Hoss, W., *Eur. J. Pharmacol.*, (1975), *32*, 66.
12. Abood, L.G. and Takeda, F., *ibid*, (1976), *39*, 71.
13. Abood, L.G., Salem, N.Jr., MacNeil, M., Bloom, L. and Abood, M.E., *Biochem. Biophys. Acta.*, (1977), *468*, 51.
14. Loh, H.H., Cho, T.M., Wu, Y.C. and Way, E.L., *Life Sci.*, (1974), *14*, 2231.
15. Cho, T.M., Cho, J.S. and Loh, H.H., *ibid*, (1976), *18*, 231.
16. Loh, H.H., Law, P.Y., Ostwald, T., Cho, T.M. and Way, E.L., *Fed. Proc.*, (1978), *37*, 147.
17. Snyder, S.H. and Simaniov, R., *J. Neurochem.*, (1977), *28*, 13.
18. Seimija, T. and Ohki, S., *Biochem. Biophys. Acta.*, (1973), *298*, 546.
19. Swift, T.J. and Connick, R.E., *J. Chem. Phys.*, (1962), *37*, 307.
20. Jarrell, H.C., Tancrède, P., Deslauriers, R., McGregor, W. and Smith, I.C.P., in preparation.
21. Birdsall, N.J.M., Hulme, E.C., Bradbury, A.F., Smyth, O.G. and Snell, C.R., *Opiates and Endogenous Opioid Peptides*, 1976, Elsevier/North-Holland Biomedical Press, Amsterdam, p. 19.
22. Newman, G.C. and Huang, C., *Biochemistry*, (1975), *14*, 3363.
23. Gent, M.P. and Prestegard, J.H., *ibid*, (1974) *13*, 4027.
24. Lyerla, Jr., J.R. and Levy, G.C., *Topics in Carbon-13 NMR Spectroscopy*, Levy, G.C., ed., Vol. 1, 1974, Wiley, New York.
25. Deslauriers, R. and Somorjai, R., *J. Amer. Chem. Soc.*, (1976), *98*, 1931.

RECEIVED March 13, 1979.

Carbon-13 Spin Relaxation Parameters of Semicrystalline Polymers

D. E. AXELSON¹ and L. MANDELKERN

Department of Chemistry and Institute of Molecular Biophysics,
Florida State University, Tallahassee, FL 32306

The overall molecular structure and morphology of a semi-crystalline polymer is generally admitted to represent a very complex situation. (1)(2)(3) Crystallinity is rarely if ever complete and, depending on molecular weight and crystallization conditions, can range from about 30 to 90% for homopolymers. (1)(4)(5) Above the level of the unit cell a lamella-like crystallite is the usual predominant feature of homopolymer crystallization, and is considered to be the primary structural entity. In addition, however, the crystallites can be organized into higher levels of morphology, or supermolecular structure such as spherulites or other geometrical forms. (6)(7)(8) There are, therefore, regions of different chain structures present in different proportions, within a semicrystalline polymer. In the most rudimentary way they can be characterized as an ordered crystalline region, a relatively diffuse interfacial region and an interzonal or amorphous region, wherein the chain units are in non-ordered conformations and connect crystallites. (1)(9) The structure and amounts of these regions determine properties. The crucial question that still needs to be resolved is the detailed structure of the non-crystalline regions, the influence of higher levels of morphology on this structure and its relation to the completely amorphous polymer at the same temperature and pressure. The thermodynamic, spectral and electromagnetic scattering properties of a large number of semicrystalline polymers have been studied for many different polymers over the last few decades. (1)(4)(5) It has been generally concluded that the degree of crystallinity is a quantitative concept. (1)(3) However, the finer details of the different structures present, and their relationship to crystallite organization, are still in need of more quantitative assessment.

Carbon-13 nuclear magnetic resonance has become an important tool with which to study the microstructure and molecular dynamics

¹Current address: DuPont of Canada Research Center, Kingston, Ontario, Canada K7L5A5

of synthetic polymers. (10) (11) The utilization of spin relaxation techniques to study the motions and to deduce the structural features of bulk synthetic polymers has been demonstrated. It is possible to observe what are essentially high resolution ^{13}C spectra of bulk amorphous polymers (11) (12) (13) (14) (15) and the non-crystalline regions of polymers (13) (16) (17) by relatively simple techniques. The observation of ^{13}C spectra for this purpose can be carried out with just complete scalar proton decoupling. The major advantages in using ^{13}C are the identification of resonant lines for the different individual carbon atoms in the chain, and despite the very often high viscosity of the medium, a lack of averaging of the relaxation parameters due to spin diffusion. To study glassy polymers or the crystalline regions, more complicated methods using dipolar decoupling, cross-polarization and magic angle spinning need to be used. (18) (19) (20)

In the present work we restrict our studies to scalar proton decoupled spectra and the determination of the spin relaxation parameters under these experimental conditions. We are thus limiting ourselves at present to probing motions, and relating them to structure, within the mobile non-ordered regions of the semicrystalline polymers at temperatures well above the glass temperature. In fact only the results at relatively high temperatures will be discussed here to avoid complications caused by transitions observed at lower temperatures. (21) Low temperature studies of linear and branched polyethylene as well as their copolymers will be reported subsequently. (22)

Experimental

Proton decoupled natural abundance ^{13}C Fourier transform NMR spectra were obtained on a Bruker HX270 at 67.9 MHz. Whenever possible the samples were studied as powders but molded films were also used. It was found that no lock material was necessary for field frequency stabilization. Drift tests showed that field fluctuations were orders of magnitude smaller (< 0.5 Hz) than the typical linewidths investigated, which were the order of several hundred Hz. Finely powdered poly(vinyl chloride) was used to suspend chunks or pellets of polyethylene for cases of sample limitation. The linewidths were found to be independent of the filler used as long as temperature variations due to decoupling were minimal. The linewidth measurements were obtained under identical experimental conditions as possible with particular attention being paid to noise modulation, bandwidth and decoupling field strength. When comparisons were important the samples were investigated consecutively, if possible. Otherwise, polymer samples of known linewidth were used as standards. With this procedure, variations were kept to a minimum and meaningful comparisons could be made. Two-level decoupling was used to minimize sample heating. (23) Quadrature detection was employed with the appropriate spectral

widths and data points to produce a well-defined resonance in the frequency domain. Linewidths were measured directly from the plotted spectra, with allowance being made for the artificial line broadening caused by applying an exponentially decaying function to the free induction decay. In some instances a computer generated least squares fitting routine was used with essentially identical results.

Spin-lattice relaxation times were measured by the fast inversion-recovery method (24) with subsequent data analysis by a non-linear three parameter least squares fitting routine. (25) Nuclear Overhauser enhancement factors were measured using a gated decoupling technique with the period between the end of the data acquisition and the next 90° pulse equal to about four times the T_1 value. Most of the data used a delay of about ten times the T_1 value. (26)

High temperature measurements were carried out using a Bruker B-ST 100/700 variable temperature unit.

Selective saturation experiments were performed with the HX270 employing the homonuclear decoupling mode available with the instrument but substituting the usual $^1\text{H} - ^1\text{H}$ situation by $^{13}\text{C} - ^{13}\text{C}$ double resonance. A Schomandl type ND100M power amplifier provided the second frequency. A spurious peak was generally observed at the irradiating frequency which could be attenuated by a slight change in delay times used. However, these have been removed from the spectra reproduced here for purely cosmetic reasons.

The properties of all but the two lowest molecular weight linear polyethylene samples have been previously described. (17) These include molecular weight, degree of crystallinity and morphological form. (8) (17) The fraction $M_n = 8.6 \times 10^3$ was obtained by column fractionation and characterized in the usual manner. (27) (28) The sample labeled 1×10^3 is also known as Polywax 1000. It is manufactured by the Petrolite Corporation. Its actual molecular weight, as obtained from gel permeation chromatography, is $M_w = 1263$; $M_n = 1136$. (29) Its melting temperature, under the conditions of the NMR measurements, was 108.8°C as determined by differential calorimetry. The low density (branched) polyethylenes studied here were commercial varieties whose molecular weights, distribution and side group concentrations have been reported. (30) The ethylene-butene-1 copolymers were a gift from the Exxon Chemical Corporation.

The four polyethylene oxide samples were obtained from the Union Carbide Corporation, and were used in the powdered form in which they were received. Their molecular weights were obtained in the conventional manner and are respectively $M_n = 3.2 \times 10^4$; $M_n = 2.7 \times 10^4$; $M_w = 6.0 \times 10^4$; $M_w = 6.67 \times 10^5$; $M_n = 6.0 \times 10^5$; $M_w = 3.3 \times 10^6$; $M_n = 1.2 \times 10^6$ for the samples studied. The enthalpy of fusion of the samples, in the original powder form, ranged from 37.8 to 39.6 cal/g, indicating that there was not very much difference in the level of crystallinity among the samples

in this form on this basis of calculation. If 45 cal/g is taken as the heat of fusion for the completely crystalline sample (4), then the degree of crystallinity is about 0.85 for these samples.

The polytrimethylene oxide sample was a gift from Professor J. E. Mark. Its viscosity average molecular weight was 100,000.

Care was taken to check periodically, by differential scanning calorimetry and small angle light scattering, whether any changes occurred in the level of crystallinity or morphology. If such changes occurred, then the samples were no longer used.

Results

Linewidths. The major interests in the experimental linewidths of semicrystalline polymers are how they depend on the morphology and level of crystallinity, their variation with temperature, and whether they are a continuous function through the melting temperature. Experiments have been designed to elucidate these factors and to supplement and make clearer those previously reported. (16) (17) The temperature dependence of the linewidth on molecular weight and level of crystallinity, of linear polyethylene, is illustrated in Figure 1. The very high molecular weight sample, $M = 2 \times 10^6$, has a degree of crystallinity, $1-\lambda$, of about 0.51, is non-spherulitic, and does not possess any well-defined supermolecular structure. (17) For this sample the linewidth is a smoothly monotonically decreasing function of the temperature. There is no indication of any discontinuity at elevated temperatures or upon melting. An asymptotic value for the linewidth of about 200 Hz is reached at 128°C. The intermediate sample of low molecular weight, $M = 8.6 \times 10^3$, with $1-\lambda$ of about 0.90 should have a rod-like morphology. (8) (31) In this case, a resolvable spectrum cannot be obtained below 90°C. Above this temperature the linewidth rapidly approaches the limiting value characteristic of the much higher molecular weight sample, $M = 2 \times 10^6$. Thus in the melt, or completely amorphous state, the linewidths for these two polymers are essentially identical despite the almost three orders of magnitude difference in molecular weight. The very lowest molecular weight sample studied, designated as 1×10^3 , with a very similar level of crystallinity and morphology as the 8.6×10^3 sample does not yield resolvable spectra until a temperature of 75-80°C is reached. Above the melting temperature for this polymer, a constant much lower limiting value of about 50 Hz is found for the linewidth. Since in this case spectra cannot be obtained below 75°C, a clear decision cannot be made as to whether a discontinuity in the linewidths occurs upon melting. The data are suggestive, however, that a discontinuity does in fact exist. In the molten state there is a very large difference in linewidth for $M = 1 \times 10^3$ as compared to the other samples. However, the asymptotic value has

already been obtained for only a very slightly higher molecular weight, $M = 8.6 \times 10^3$.

The influence of morphology on the linewidth-temperature relations is illustrated in Fig. 2 for two high molecular weight samples which have the same degree of crystallinity at room temperature. (17) The major influence of the morphology on the observed linewidths, that have been previously reported (17), manifests itself below 90°C. As the temperature is decreased the linewidth differences become continuously greater between the two samples. In the vicinity of 30-40°C the differences correspond to the values previously reported. (17) Above 90°C the linewidths of the two samples are indistinguishable from one another, and both have the same value in the melt. The continuous character of both curves indicates that either the fusion process with regard to morphology is different in the two cases; or if the morphological forms are maintained, the structures in the non-crystalline regions possess a different temperature dependence. More detailed studies which relate any changes in morphology with the course of fusion for these kinds of samples are necessary to resolve these points. For present purposes, the major, definitive conclusions are that the temperature dependence of the linewidth is continuous through the melting temperature for these two extremes in morphology. Moreover, depending on the morphology, significant differences in the magnitude of the linewidth are observed below 90°C.

Fig. 3 represents a similar phenomenon for two samples which possess spherulitic morphologies but have different levels of crystallinity. At the intermediate temperatures the increased degree of crystallinity yields only a slightly greater linewidth for the same morphology. This small difference remains constant until the melting is reached, at which point the two curves merge. At the lower temperatures, the same high value characteristic of spherulites and independent of the level of crystallinity is observed as was previously reported. (17)

The plot in Fig. 4 represents in the same diagram the influence of both the degree of crystallinity and morphology, from examples taken from the previous two figures. As would be anticipated from the previous results, the differences in the linewidths below the melting range are greatly enhanced when the data are examined in this manner. However, as is equally clear, the results would be difficult to sort out for interpretive purposes if a clear distinction was not made between the levels of crystallinity and morphology, or lack thereof, as distinctly different, independent quantities.

The linewidth-temperature relation of the polyethylene oxide samples are given in Fig. 5. Despite the large differences in molecular weight, these samples have about the same linewidth, 300-350 Hz, in the crystalline state at 25°C. They all also possess a spherulitic type of morphology. The influence on the linewidth of the different types of supermolecular structures

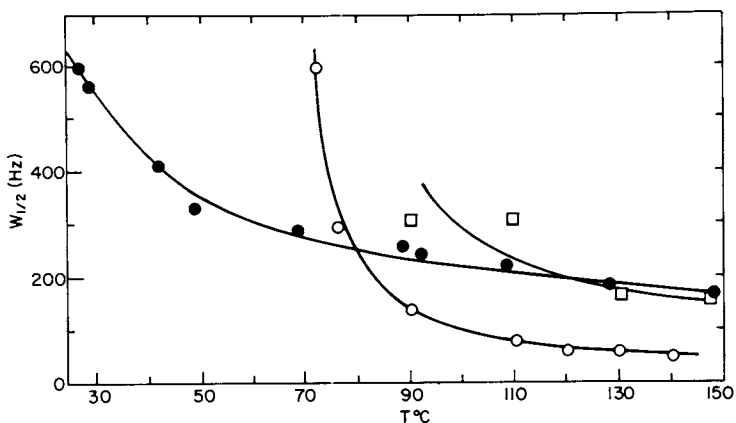


Figure 1. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for linear PE's of indicated molecular weights: (●), 2×10^6 mol wt; (□), 8.6×10^3 mol wt; and (○), 1×10^3 mol wt.

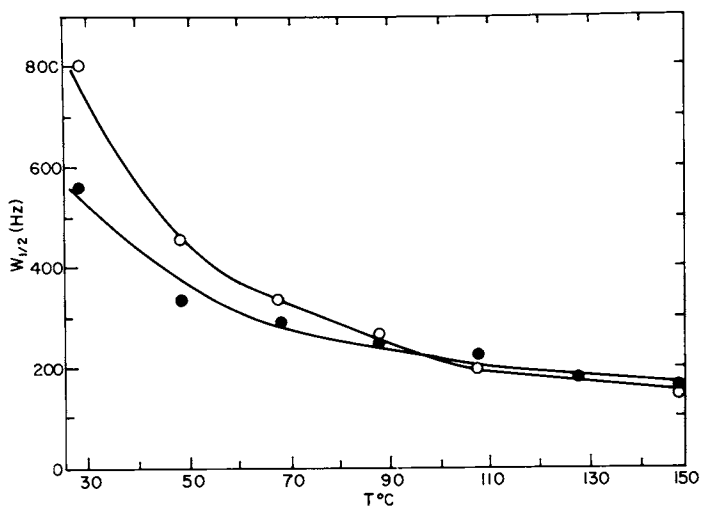


Figure 2. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for two linear PE samples of same degree of crystallinity (0.51) but differing morphologies: spherulitic, (○); no morphology (●).

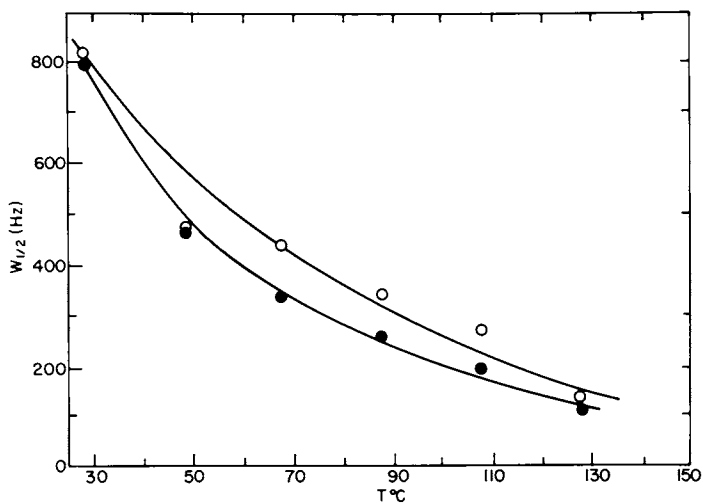


Figure 3. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for two linear PE samples having a spherulitic morphology but differing levels of crystallinity: degree of crystallinity 0.78, (○); degree of crystallinity 0.50, (●).

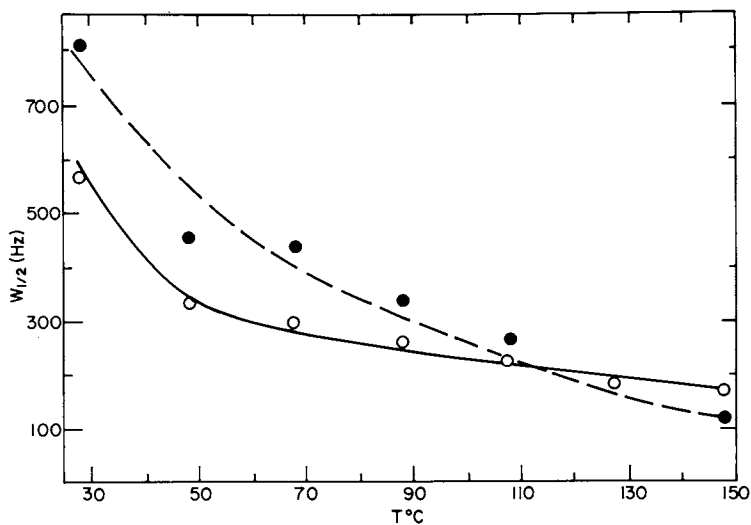


Figure 4. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for two linear PE samples of differing morphologies and levels of crystallinity. No morphology, degree of crystallinity 0.50, (O); spherulitic morphology, degree of crystallinity 0.78, (●).

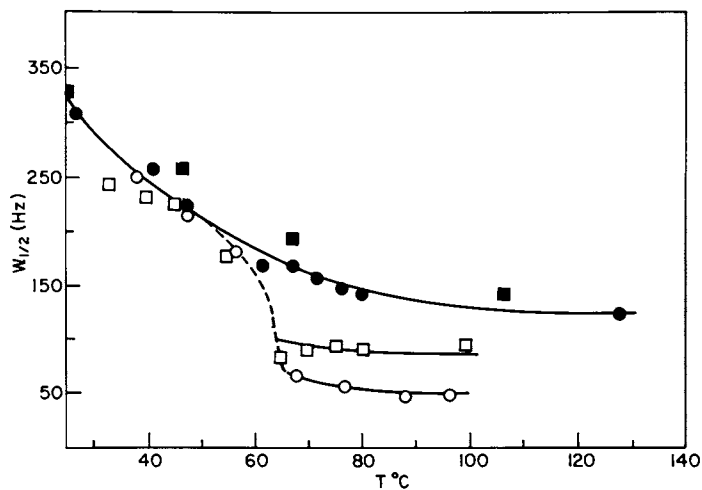


Figure 5. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for PEO samples of indicated molecular weights: (■), 3.3×10^6 mol wt; (●), 6×10^5 mol wt; (□), 6×10^4 mol wt; (○), 3×10^4 mol wt.

that can be developed in polyethylene oxide has not as yet been investigated. For the two highest molecular weight samples studied here, the linewidths are a continuously decreasing function of temperature. A limiting value of approximately 135 Hz is attained at the elevated temperatures in the melt. There is no evidence of any discontinuity in this quantity in the melting region of 60-70°C. However, although all four samples have essentially the same linewidth in the crystalline state, there is a definite discontinuity upon melting for the two lowest molecular weight samples. The change in linewidth is greatest for the lowest molecular weight sample and is independent of temperature above its melting temperature. The limiting linewidths in the molten state are 80-90 Hz for $M = 6 \times 10^4$ and are reduced to 47 Hz for $M = 3 \times 10^4$. In contrast to linear polyethylene the asymptotic linewidth for the melt of polyethylene oxide is attained at higher molecular weights. However, for both polymers the asymptotic linewidths are relatively broad.

Because of the ultimate interest in understanding the segmental motions that contribute to linewidths and the apparently broad lines that are observed for both linear polyethylene and polyethylene oxide, we have also studied a series of random ethylene-butene-1 copolymers. Because of the high co-unit content of these samples and the temperature range studied, if these polymers are crystalline at all, the level is very low. Above 50°C they are essentially completely amorphous. (32) The linewidth measurements for four such copolymers are given in Fig. 6. Above 50°C the linewidths only change slightly and are in the range of 180-200 Hz, similar to the melt of the homopolymer. As the temperature is lowered well below 35°C crystallinity begins to develop. This problem will be discussed in detail in a subsequent paper. (22) For present purposes, for these copolymers, we are mainly concerned with the linewidths in the amorphous state.

Spin-Lattice Relaxation Parameters, T_1 . In Fig. 7 the NT_1 values for polyethylene oxide samples are plotted as a function of temperature. Also plotted are the average values for the two different carbons of polytrimethylene oxide. Except for the very lowest molecular weight polyethylene oxide sample (which corresponds to a degree of polymerization of only 4.5), the spin-lattice relaxation time is the same for all the samples at a given temperature and is also a continuous function of the temperature. The respective melting temperatures of each of the polymers is indicated by the vertical arrows in the figure. It thus becomes abundantly clear from this extensive set of data that the T_1 's are continuous functions through the melting temperature. The segmental motions of the non-crystalline regions which are represented by this parameter are not dependent on the state of the polymer. The lowest molecular weight polyethylene

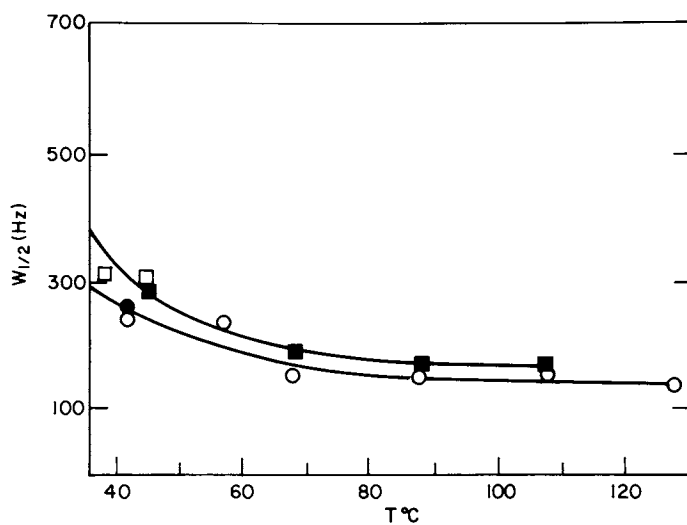


Figure 6. Plot of linewidth, $W_{1/2}$, against temperature at 67.9 MHz for ethylene-butene-1 random copolymers. Co-unit contents: (■), 10; (□), 7; (●), 21; (○), 26.

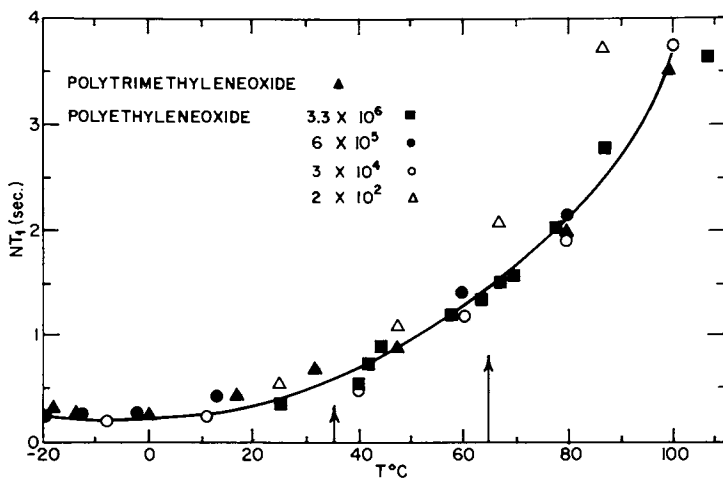


Figure 7. Plot of spin-lattice relaxation time against temperature, at 67.9 MHz, for polytrimethylene oxide and PEO.

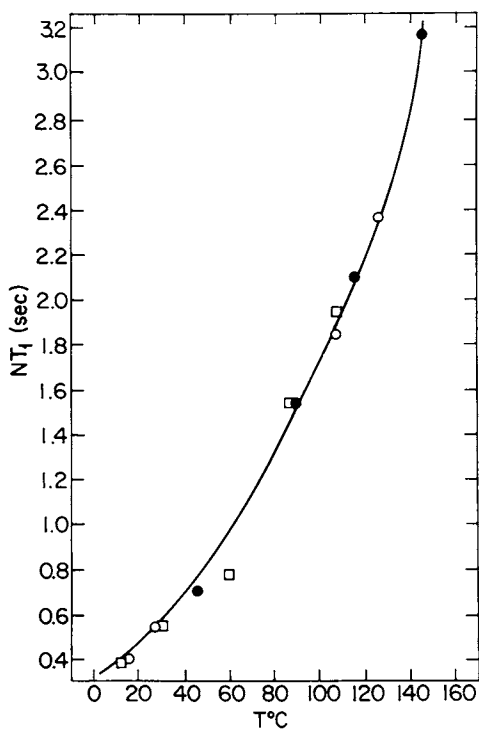


Figure 8. Plot of spin-lattice relaxation time against temperature, at 67.9 MHz, for linear and branched PE. Linear PE from Ref. 17, (●); linear PE this work, (○); branched PE this work, (□).

oxide sample gives significantly higher NT_1 values at the elevated temperatures.

A similar continuity in the ^{13}C T_1 's through the melting temperature was previously reported for linear polyethylene. (17) We have now investigated the temperature dependence of this quantity, for this polymer, in more detail and have also studied a low density (branched) polyethylene. The results for the polyethylenes are summarized in Fig. 8. The new data reported here substantiate the conclusion previously reached for linear polyethylene. A similar conclusion can now be reached for the backbone carbons of low density (branched) polyethylene. The melting temperature for this particular sample, under the crystallization conditions studied, is less than 110°C . (33) Thus, the ^{13}C spin-lattice relaxation parameters for the backbone carbons are the same for both the linear and branched polymers over the temperature range studied here. Changes that occur in T_1 as the temperature is reduced below 0°C involve other considerations and will be discussed in detail elsewhere. (22)

Nuclear Overhauser Enhancement Factor. The nuclear Overhauser enhancement factor, NOEF, has not been as extensively studied for the pure semi-crystalline polymers as have the other ^{13}C relaxation parameters. The previously reported results for linear polyethylenes (17), with varying levels of crystallinity and different supermolecular structures, are included in the summary of Table I. For this polymer at 45°C and 67.5 MHz, within the limits of the experimental results presently available, the NOEF appears to depend primarily on the level of crystallinity and not on any other morphological, structural or molecular factors. The samples with the lowest level of crystallinity that could be attained with this polymer, 0.50, possess full NOEF's which are also found in the pure melt. (34) For the higher crystallinity samples the NOEF is reduced, leading to the suggestion that the degree of crystallinity could be an important factor in determining this quantity. We have explored the temperature dependence of the sample which had an NOEF of 1.5 at 45°C . Although the NOEF increased slightly to 1.7 at 100°C , it has not yet achieved its maximum value at this point. Since the full NOEF must be attained in the pure melt, it is not as yet clear whether or not the change will be continuous.

The NOEF's for two of the polyethylene oxide samples, $M = 3.2 \times 10^4$ and 6.67×10^5 , were determined, as a function of temperature, from room temperature through the melting point. In both cases full NOEF's were observed at room temperature and in the melt. Schaefer and Natusch have reported essentially a full NOEF, 1.7 ± 0.1 , for a 1000 molecular weight polyethylene oxide which is a liquid at room temperature. (35)

It is clear that more extensive studies are needed before any detailed interpretation can be given to the NOEF's, or trends discerned, relative to the amorphous structure of the

Table I.
Carbon-13 Spin Relaxation Parameters of Linear Polyethylene (17)
at 45°C and 67.9 MHz

M_n	$(1-\lambda)$ ^d	Morph.	T_1 (msec) ^a	Linewidth $W_{1/2}$ (Hz) ^b	NOEF
8.1×10^4	0.57	Spher.	343	622	-
2.5×10^5	.51	Spher.	355	625	2.0 ^c
* 1.7×10^5	.81	Spher.	348	695	1.5 ^d
* 1.7×10^5	.68	Spher.	356	700	-
<hr/>					
* 2.0×10^6	.51	None	369	501	2.0 ^c
* 2.0×10^6	.72	None	358	496	-
6.1×10^6	.54	None	-	500 ^e	-
6.1×10^6	.70	None	-	503	-
2.75×10^4	.94	Rod	352	945	1.0 ^d

* Unfractionated sample

^a estimated accuracy $\pm 10\%$

^b estimated accuracy $\pm 5-10\%$

^c estimated accuracy ± 0.1

^d estimated accuracy ± 0.2

^e estimated

semicrystalline polymers. However, there are certain inherent restraints which make difficult the determination and collection of as much data as would be desired. There are no problems with systems which can be described by a one-correlation time model, such as linear polyethylene of low levels of crystallinity (17) and the polyethylene oxide samples just described. However, when a distribution of correlation times are necessary to describe the results, complications can exist. For example, even in the completely amorphous state *cis* polyisoprene at 40° and 0°C possesses low NOEF's which are very close to the theoretically allowed minimum. These results are a consequence of the distribution of relaxation times that are required to describe the system. (11) (13) (14) (16) Consequently, upon crystallization very little change can be expected, as is observed experimentally. (16) Thus the number of systems that can be effectively studied, to assess the influence of crystallinity on the NOEF's, are limited. These will have to be intensively studied in more detail before any general conclusions can be deduced.

Discussion

Spin-Lattice Relaxation Parameters. As is summarized in Table I, it was previously shown for linear polyethylene that at fixed temperature, the T_1 's are constant for a wide range in the level of crystallinity and molecular weight and for different morphologies or lack thereof. (17) The levels of crystallinity varied from 0.50 to 0.94 in this study. Thus the segmental motions, reflected in T_1 , are dependent only on the properties of the amorphous state. This conclusion is further substantiated by the temperature dependence of T_1 , which is illustrated in Fig. 8. These data clearly demonstrate the continuity of T_1 through the melting temperature for both linear and branched polyethylene. Consequently as far as the T_1 measurements are concerned, the segmental motions from the non-crystalline regions that contribute to this quantity are identical to those of the pure melt. The amount of crystallinity, the organization of the crystallites and the temperature, therefore, have no influence on the high frequency segmental motions in the non-crystalline regions.

These conclusions are further generalized by the more extensive data presented in Fig. 7 for polyethylene oxide and polytrimethylene oxide. The continuous nature of the T_1 function for both these polymers over a large temperature range is quite definite and is emphasized by the detailed data in the vicinity of the respective melting temperatures. This is true even for the polyethylene oxide samples where discontinuities in the linewidth are clearly indicated in Fig. 7. Obviously, the type of segmental motions which contribute to the two different relaxation parameters are influenced quite differently by the presence of crystallinity.

The T_1 values for the very low molecular weight polyethylene

oxide sample are essentially the same as the high molecular weight ones at low temperatures. However, above 50°C the NT_1 increases. Since the degree of polymerization for this sample is very low, being only a few repeating units, overall molecular reorientation can take place at these elevated temperatures. Thus individual molecular contributions are being made. A similar behavior for low molecular weight polyethylene oxide has been reported (36) as have proton T_1 measurements for low molecular weight polyethylene. (37)

The major results described could be partially anticipated from those previously reported for linear polyethylene (17) as well as those for *cis* polyisoprene. (16) For the latter polymer, by taking advantage of its crystallization kinetic characteristics, it was possible to compare the ^{13}C relaxation parameters of the completely amorphous and partially crystalline polymer (31% crystallinity) at the same temperature, 0°C. This is a unique situation and allows for some unequivocal comparisons. It was definitively observed that for all the carbons of *cis* polyisoprene the T_1 's did not change with crystallization.

Based on the results obtained to date, which have been summarized above for several different semicrystalline polymers--linear and low density (branched) polyethylene, polytrimethylene oxide, polyethylene oxide and *cis* polyisoprene--it is concluded that the relatively fast segmental motions, as manifested in T_1 , are independent of all aspects of the crystallinity and are the same as the completely amorphous polymer at the same temperature. Furthermore, it has previously been shown that for polyethylene, the motions in the non-crystalline regions are essentially the same as those in the melts of low molecular weight *n*-alkanes. (17)

Proton NMR relaxation parameters have also been determined for polyethylene (38) and polyethylene oxide (39) in the melting region. The apparent contradiction between the proton spin-lattice relaxation parameter for a high molecular weight linear polyethylene sample at its melting point, with the ^{13}C relaxation measurements, has previously been pointed out. (17) This discrepancy is still maintained with the more detailed results reported here for both types of polyethylene. For the proton relaxation a small, but distinct, discontinuity is reported at the melting temperature. (38)

On the other hand, Connor and Hartland (39) have reported results of a proton NMR study for a series of polyethylene oxide samples by rotating frame proton relaxation time $T_{1\rho}$ measurements. T_1 was also determined. For their lowest molecular weight sample, $M = 550$, the $T_{1\rho}$ values display a fairly sharp discontinuity at about 12°C. This temperature is close to the independently determined melting range of 15-25°C. However, there is no evidence for a discontinuity in the T_1 measurements for this sample. Above the melting temperature the values found for T_1 and $T_{1\rho}$ were similar. $T_{1\rho}$ exhibited a very pronounced discontinuity at about 63°C for the sample $M = 6000$. This discontinuity occurs

within the reported melting range of 60-63°C. Although there is a minimum of T_1 in this temperature region, there is no indication of any discontinuity. In contrast to these two lower molecular weight samples, for a high molecular weight sample that was also studied, $M = 2.8 \times 10^6$, no sharp discontinuity was observed in $T_{1\rho}$ in the vicinity of the melting temperature and T_1 was continuous. In summary, for the two lowest molecular weight samples, $T_{1\rho}$ displayed a discontinuity in the melting region while T_1 was continuous. For the very high molecular weight sample both of these relaxation parameters were continuous with temperature. These results are reminiscent of and very similar to those reported here for the ^{13}C relaxation parameters of polyethylene oxide as illustrated in Figs. 5 and 7. We have found that the T_1 's are continuous with temperature over a very wide molecular weight range. However, as is shown in Fig. 5, there are discontinuities in linewidths at the melting temperatures for the low molecular weight samples, but the linewidths are continuous for the higher molecular weight samples. The $T_{1\rho}$ results of Connor and Hartland (39) display virtually the same behavior. These similarities do not appear to be coincidental since the linewidth is directly related to the spin-spin relaxation parameter, T_2 , which is sensitive to low frequency motions. The $T_{1\rho}$ values are also more sensitive to the lower frequency motions and would thus be expected to behave similarly to T_2 , or linewidths, in the ^{13}C relaxation studies.

Linewidths. Ideally the spin-spin relaxation time, T_2 , can be obtained from the linewidth $W_{1/2}$ by the relation $T_2 = 1/(\pi W_{1/2})$. Although in the present work we are mainly interested in the influence of the different aspects of crystallinity on T_2 , it is informative to examine the linewidth in the completely amorphous state above the melting temperature. This state conveniently serves as a reference point for the subsequent discussion. As we shall deduce in the subsequent discussion, it is also very important in establishing an understanding of the conditions for a discontinuity to be observed in the vicinity of the melting temperature. We have previously reported (15) that for non-crystalline polyisobutylene samples, at 45°C and 67.9 MHz, the linewidths for all the carbons are independent of chain length for molecular weights greater than about $4-5 \times 10^4$. Below this molecular weight the linewidths are substantially reduced with decreasing chain length. The limiting, or leveling-off, value for the methylene carbons is about 200 Hz for this polymer. For linear polyethylene, we observed a very similar effect, as was illustrated in Fig. 1. In the pure melt above the melting temperature at 140°C, and at the same frequency, a limiting value of about 200 Hz for the linewidth is observed. The limiting value is attained at a much lower molecular weight for this polymer under these conditions. For the very low molecular weight sample, $M = 1 \times 10^3$, the linewidth is substantially reduced

to about 50 Hz. The completely amorphous ethylene-butene-1 copolymers, whose molecular weights are in the asymptotic region, have linewidths of the order of 180-200 Hz above 50°C. These are very similar to the completely amorphous homopolymer. The results for polyethylene oxide follow the same patterns. An asymptotic linewidth of 135 Hz is reached in the molten state between 6×10^4 and 6×10^5 . A monotonic decrease in linewidth is observed below this molecular weight. For high molecular weight cis polyisoprene, in the completely amorphous state at 40°C and at 67.9 MHz, the linewidths for all the carbons are only about 40 Hz. (16) Although additional type polymers should be studied before firm generalizations can be made, the data in hand indicate certain salient features relative to the amorphous states. There is a low critical molecular weight, whose exact value varies with polymer type, above which the linewidth and thus T_2 is independent of chain length. This behavior has now been observed with the three different polymers studied in detail and could be expected to be universal. Except for cis-polyisoprene, the linewidths for the other polymers are relatively broad compared to other type molecular systems. The rather narrow lines observed for the methylene carbons of cis polyisoprene, about 40 Hz at 67.9 MHz and 20 Hz at 22.9 MHz (16), are also found in the melt of other diene polymers. (22) They would appear from the data obtained so far to be atypical of polymers and a consequence of the double bond in the chain.

In contrast to the spin-lattice relaxation parameters, which remain invariant, a substantial broadening of the resonant lines occurs upon crystallization. The effect is relatively modest for cis polyisoprene at 0°C and 67.9 MHz, where comparison can be made at the same temperature. Here there is about a 50% increase in the linewidths upon the development of 30% crystallinity. Schaefer (13) reports approximately 3- to 5-fold broader lines (but they are still relatively narrow) for the crystalline trans polyisoprene relative to the completely amorphous cis polyisoprene at 40°C and 22.6 MHz. It is interesting to note in this connection that for carbon black filled cis polyisoprene the linewidths are greater by factors of 5-10 relative to the unfilled polymer.

In the present work the limiting value of the linewidths for polyethylene oxide increases from 135 Hz in the melt above 70°C, to the range 300-350 Hz in the crystalline state at room temperature. As is indicated in Table I, the resonant linewidths for linear polyethylene increase substantially upon crystallization and attain values in the range 500-900 Hz at 45°C and 67.9 MHz. As has been emphasized previously (17), the level of crystallinity is not the major determinant of the linewidth in the semicrystalline state. Rather the supermolecular structure or morphology is a major factor in governing the magnitude of the linewidth. Structural factors and crystallization conditions under which low density (branched) polyethylene forms

either spherulites or no well-defined morphology have recently been established. (33) Samples from each of these structural categories yield linewidths which are virtually identical with the corresponding values listed in Table I for the linear polymer. A morphological map, similar to those developed for linear and branched polyethylene (8) (33), has not as yet been completed for polyethylene oxide. Thus a further generalization of these important findings to this polymer cannot be made as yet.

The results that have been obtained indicate that the major influence of the crystalline regions on segmental motions, and hence to the structure of the non-crystalline regions, is in the linewidth and T_2 . The different morphologies are reflected in different values of T_2 . The segmental motions in long chain molecules which exert major influence on the spin-lattice relaxation times and the nuclear Overhauser enhancements are not in general the same motions which determine the resonant linewidth. T_1 is in general greater than T_2 . This difference can in part be a consequence of the slower modes of polymer motion, which are characterized by correlation times sufficiently long that they do not contribute significantly to T_1 but do to T_2 . It is therefore important, in terms of describing the fine structure of the non-crystalline regions, to understand the type motions which contribute to T_2 and to develop a rationale for the relatively broad lines that are observed for most crystalline polymers.

There are many possible factors that can contribute to the linewidths. It is important, therefore, that the pertinent ones be discerned and understood if the molecular interpretations are to be eventually deduced. The task is a formidable one and the complete solution of the problem is not as yet in hand. Many of the possible contributions to the linewidth, and reasons for the apparently excessive broadening, have been previously discussed. (11) (13) (17) Several mechanisms have emerged as being most likely contributors. One must, however, be willing to accept the concept that several different mechanisms can be simultaneously involved and contributing to the line broadening. Therefore it is necessary that a diverse set of experiments be designed and carried out, to substantiate or dismiss the different possibilities. A unique process is not necessarily required and should not be established as an objective. A large number of different type experiments are necessary to sort out the different possibilities. In consideration of the amount of work involved, the problem has not as yet been completely resolved.

Line broadening due to inhomogeneity in the static magnetic field, H_0 , as well as in the rf pulse H_1 , can contribute to the observed resonance. However, studies of standard samples, of known natural linewidths, enable the contributions from this source to be determined. In the present case these causes contribute only a few percent, i.e., a few Hz, to the total linewidth and are thus inconsequential to the present problem. Before discussing the different motional contributions to the linewidth,

the possibility that non-motional or static phenomena can also make a substantial contribution needs to be given serious consideration. (40) (41) Differences in bulk magnetic susceptibility within the same volume element can result in differences in nuclear screening among nuclei in different regions of the sample, resulting in a broadening of the resonance lines. (40) (41) (42) Although such broadening can occur from the irregular macroscopic sample configuration, it most likely arises from microscopic structural differences within the sample. Broadening from this cause alone will vary linearly with the applied field. (41) Thus, because of the high field used in the present work, if this process were operative, it could be quite severe, and would have escaped notice in most previous studies which have been conducted at much lower field strengths. Consequently, we have carried out a detailed study of the frequency dependence of the linewidths for the polymers studied here. Particular attention has been given to the influence of crystallinity, morphologic form and temperature. An extensive set of data has now been collected and analyzed. We shall limit ourselves here to a brief summary of the major findings as they pertain to the major themes of the present work. A more detailed report of these findings will be presented elsewhere. (43)

A substantial effect of the field on the resonant linewidths was found for the crystalline polyethylenes and polyethylene oxides. The magnitude of the changes with frequency is in qualitative accord with theoretical expectation. If other molecular and constitutional factors are held constant, then the influence of the morphology on the linewidth, which was previously observed at 67.9 MHz, is still maintained at the lower frequencies. Thus, the low frequency segmental motions of the non-crystalline regions are definitely influenced by the morphology; the previous conclusion was not a consequence of the high fields that were used. This fact thus has important molecular implications with respect to the structure. For these samples the extrapolation of the linewidth to zero frequency does not pass through the origin. Rather large residual values, about 100 Hz for the crystalline polyethylenes and about 25 Hz for crystalline polyethylene oxides, are found. This result is consistent with a residual dipolar coupling contribution to the resonant linewidth. Preliminary magic angle spinning experiments that we have performed with crystalline polyethylene oxide at low spinning frequencies substantiate that the field-dependent broadening has a major static contribution from microscopic inhomogeneities. Thus, there are at least several contributions to the resonant linewidth and its broadening.

The homopolymers of low levels of crystallinity, as well as the ethylene-butene-1 copolymers, which are either completely amorphous, or slightly crystalline at the temperatures of measurement, also display frequency-dependent linewidths. Although these effects are not nearly as severe as in the more

crystalline samples, they are not easily understood. Residual dipolar couplings could readily account for a portion of the linewidths in these cases.

The frequency dependence of the linewidth and the contributions from microscopic inhomogeneities strongly suggest significant inhomogeneous broadening. If these contributions are not completely averaged in an experiment, they will give rise to a distribution of chemical shifts and an inhomogeneous resonant line. (13) Bloembergen, Purcell and Pound (44) have shown that single frequency irradiation of a line whose width is dominated by magnetic field inhomogeneities results in the local saturation of the line. This is the so-called "hole burning" experiment and has been carried out successfully by Schaefer for several polymer systems. (13) In this experiment it is possible to determine the natural dipolar linewidth in the presence of macroscopically or microscopically inhomogeneous magnetic fields. (45) However, when attempting to saturate a resonance whose width is determined by strong spin-spin interactions, rather than field inhomogeneities, the entire line becomes saturated. In this situation the energy absorbed by the spins is no longer localized; instead, the temperature of the spin system as a whole is raised. This situation is illustrated in Fig. 9 for dioxane (15% acetone- d_6 , 85% dioxane, ambient temperature, 67.9 MHz). This homogeneous resonant line was chosen for illustration and for comparison with a linear polyethylene sample. The spectra on the right demonstrate that an increase in the saturating rf field causes a decrease in the intensity of the resonance. Simultaneously, however, the location of the line, i.e., the point of maximum intensity, as well as the linewidth remain constant, because of the homogeneity.

The results of this type experiment for a linear, non-spherulitic polyethylene sample are shown in Figs. 10 and 11. In Fig. 10 the rf irradiation was applied at approximately the location of the maximum in the line intensity. The power levels were progressively increased to saturate the major portion of the resonance. An irregularly shaped resonance is observed. Fig. 11 demonstrates more clearly the inhomogeneous nature of the initial polyethylene resonance. In this instance, the position of the irradiating rf is progressively moved upfield, as is indicated by the vertical arrows, from the position of maximum line intensity in the unperturbed spectrum. Different power levels were used concurrently. The uppermost spectrum in Fig. 11 was obtained with no selective rf irradiation being applied and is the reference against which the remaining spectra should be compared. It is again apparent from these results that the resonant line is inhomogeneous since the symmetry, linewidth at half-height, and peak maxima change with the position and strength of the irradiation.

The inability to "burn" a narrow hole in the polyethylene spectrum is an indication that the "natural homogeneous linewidth"

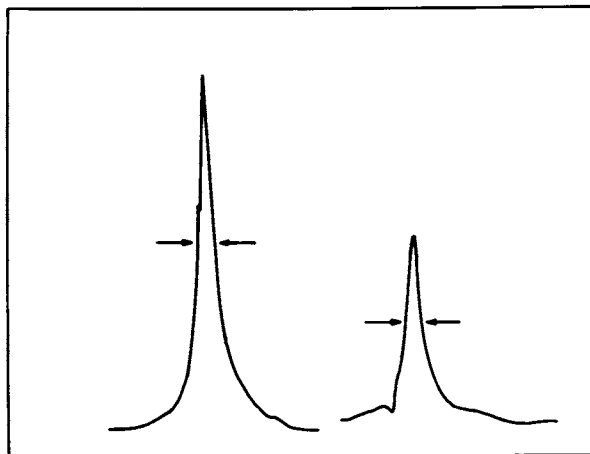


Figure 9. Selective irradiation of dioxane. Homonuclear irradiation of a homogeneously broadened resonance at 67.9 MHz. Spectral details: PW = 40 μ sec (90°C), D2 = 10 sec, 4 scans accumulated, quadrature detection; 15% acetone- d_6 , 85% dioxane mixture.

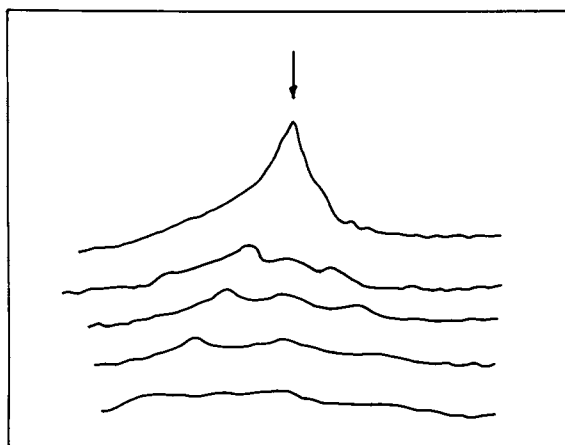


Figure 10. Selective irradiation of linear PE (2×10^6 mol wt, $1 - \lambda \sim 0.5$). Spectral details are: 35°C; 67.9 MHz; sweep width ± 5 KHz (quadrature detection); line broadening 9.7 Hz; pulse width 35 μ sec (90°C = 48 μ sec); delay = 1.0 sec, 4K data points; 1024 scans accumulated; 10-mm sample tube. Decoupling: 7W (forward), 0.4W (reflected), broad band noise modulated decoupling.

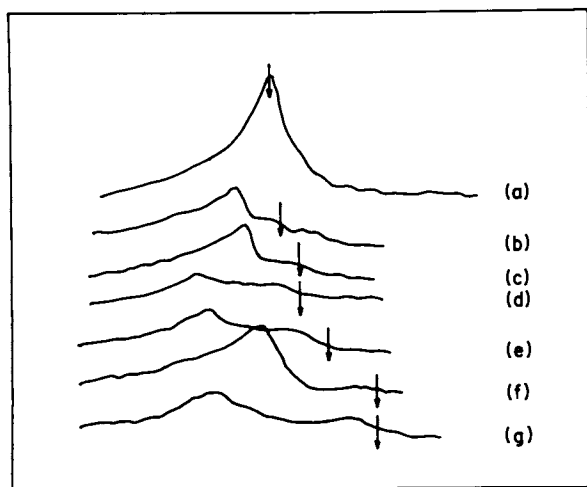


Figure 11. Selective irradiation of linear PE. Spectral details: 35°C, 67.9 MHz; variable power levels. Offsets in Hz from center frequency are: (a) 0 Hz; (b) 100 Hz; (c) 300 Hz; (d) 300 Hz; (e) 700 Hz; (f) 1300 Hz; and (g) 1300 Hz. Same conditions as in Figure 10.

for the particular sample studied is quite large. This conclusion is consistent with the large linewidth obtained by extrapolating the frequency data to zero frequency.

On the other hand, Schaefer (13) has shown from selective saturation experiments of amorphous cis polyisoprene, crystalline trans polyisoprene, as well as carbon black filled cis polyisoprene, that the resonant lines are homogeneous. The linewidths in these cases are thus not caused by inhomogeneous broadening resulting from equivalent nuclei being subject to differing local magnetic fields. The results for these systems are thus contrary in part to what has been found here.

At this point it has been established that there are at least two basic mechanisms which contribute to the broad lines that are observed for the crystalline polymers. The residual zero frequency line broadening component can be analyzed in more detail. Specific attention can be given to factors which are a consequence of the chain-like character of the molecules. The local field at a given nucleus is the sum of the individual fields contributed by the neighboring magnetic nuclei. Segmental motions will induce a time dependence to the variables so that the individual contributions can be described by the equation: (46)

$$H_{ij}(t) = \pm \mu_{ij}/r_{ij}^3 [3 \cos^2 \theta_{ij}(t) - 1] \quad (1)$$

If r_{ij} is assumed to be constant, i.e., the directly bonded protons provide the dominant contribution, and $\theta_{ij}(t)$ is only time dependent, the time averaged local field is given by

$$(\mu_{ij}/r_{ij}^3) \int_0^{T_2} (3 \cos^2 \theta_{ij}(t) - 1) dt. \quad (2)$$

Here T_2 is the order of time in which the nucleus resides in a given spin state. If there are no restrictions on the directions available to the internuclear vector, then the time average can be replaced by a space average, with the result that

$$(\mu_{ij}/r_{ij}^3) \int_0^\pi (3 \cos^2 \theta_{ij} - 1) \sin \theta d\theta = 0. \quad (3)$$

For non-viscous liquids, where the above condition will be expected to be fulfilled, narrow resonances are observed, when only this source of line broadening is involved.

Other types of restrictions can be thought of being imposed on the parameters of Eq. (1), when polymeric systems are involved. For example, the orientation angles may be unrestricted with respect to accessibility but the segmental motions may not be sufficiently rapid to average θ_{ij} over the angular range 0 to π in the time interval T_2 . Conversely the motions may be relatively rapid but the angular range may be restricted. It has been calculated that excluding θ_{ij} from but a few orientations, i.e., excluding the magnetization vectors from solid sectors of only a few degrees, is sufficient to produce broadening by about an order

of magnitude. (13) (46) This latter process has been termed incomplete motional narrowing. It was also noted (13) that in a selective saturation experiment a partially motionally-narrowed line can be expected to behave as a single dipolar-broadened NMR line.

Schaefer (13) (47) has interpreted the linewidths of cis polyisoprene in this context and concluded that the data could be explained by assuming that not all spatial orientations were accessible to the chain units as a consequence of restrictions imposed on the segmental motions. Chain entanglements were postulated to be the major source of these restrictions for this amorphous polymer. In the carbon black filled cis polyisoprene, the filler itself was considered to be an additional source of entanglements. For semicrystalline polymers, the presence of crystallites and their relative arrangement could play a similar role as well as introducing inhomogeneities into the system which can serve as another source of broadening.

The line broadening caused by partial motional narrowing can be distinguished from that due to isotropic reorientation at a reduced rate by appropriate magic angle spinning experiments. (13) (47) Random isotropic motion at reduced rates covers frequencies of the order of the inverse of the correlation time, i.e. of the order of 10^5 - 10^7 Hz. Hence, sample rotation at the usually accessible rates of 10^2 - 10^4 Hz at the magic angle will have no influence since the linewidths are determined by frequencies several orders of magnitude greater. Partial motional narrowing, however, results in the linewidth being determined in part by very low or zero frequency components. These are affected by fast magic angle spinning. The extent of line narrowing that is obtained depends on the distribution of the frequencies generated by the residual dipolar interaction relative to the spinning frequency.

The concept of chain entanglements influencing the linewidths, or T_2 's, can be examined more directly by studying the influence of molecular weight. It is well established that the zero shear bulk viscosity of all amorphous polymers is directly proportional to the molecular weight below a critical low molecular weight, M_c , and above this molecular weight increases as the 3.5 power of M . (48) (49) (50) M_c represents approximately twice the molecular weight between chain entanglements. The molecular weight dependence of the viscosity results from the frictional loss in the two different molecular weight regions. (48)

This molecular weight dependence manifests itself quite distinctly in the ^{13}C linewidths in the completely amorphous state and is consistent with certain proton NMR T_2 studies that have been reported. For polyisobutylene the ^{13}C linewidths are invariant at 45°C for each of the carbons for molecular weights greater than 4.5×10^4 over a 60-fold change in molecular weight. (15) This range corresponds to a change of eight orders of

magnitude in the bulk viscosity. There is a decrease in the linewidth below this molecular weight. From bulk viscosity measurements, M_c is found to be 1.5×10^4 (51) (52), which is the same order of magnitude that was found for the invariant linewidth. The polyethylene linewidths, as illustrated in Fig. 1, clearly indicate that a constant value is attained in the melt between $1-8 \times 10^3$. Proton T_2 measurements at 150°C indicate that there is a change in slope in this quantity as a function of molecular weight at about $M_n = 6 \times 10^3$. (37) The critical molecular weight as determined from bulk viscosity measurements for this polymer has been given as 2×10^3 (53) and 3.8×10^3 . (52) These values are very close to those which would be deduced from the linewidth measurements. For polydimethyl siloxane the break in the proton T_2 -molecular weight curve occurs at about $M = 5 \times 10^4$. (37) M_c is about 2.5×10^4 from viscosity measurements. (52) Fig. 5 indicates that for polyethylene oxide at 90°C the linewidth becomes constant between 6×10^4 and 6×10^5 . The viscosity data indicates that the critical molecular weight is about 10^4 . (54) (55) (56) For all the cases cited above, which represent those data for which a comparison can be presently made, there is a direct connection between the critical molecular weight representing the influence of entanglements on the bulk viscosity and other properties, and the NMR linewidths, or spin-spin relaxation parameters of the amorphous polymers. Thus the entanglements must modulate the segmental motions so that even in the amorphous state they are a major reason for the incomplete motional narrowing, as has been postulated by Schaefer. (13) This effect would then be further accentuated with crystallization.

From the above, the observations that in some cases a discontinuity at the melting temperature is observed in the ^{13}C linewidths, while in other situations the linewidth is continuous, can be readily explained. For polyethylene oxide the linewidths in the crystalline state, for the samples studied to date, are all about the same, presumably due to the similarity in morphology and level of crystallinity. However, due to the differences in the linewidth in the amorphous state, the lower molecular weight samples must exhibit a discontinuity at the melting temperature, while the higher ones will be continuous. A similar situation will exist for polyethylene. In this case it has been seen in Fig. 1 that the major influence of morphological form on the linewidth at lower temperatures has disappeared for the high molecular weight sample. Thus it is continuous upon melting. The continuity of the ^{13}C linewidth over a large temperature range, above room temperature, that has previously been reported (17) and is further detailed here does not reveal any change in the vicinity of the α transition region ($80-100^\circ\text{C}$). Major changes are found in broad line proton NMR experiments, which measure linewidths or second moments. (57) There is, however, no discrepancy between these results since the α transition is a property of the crystalline regions. The proton measurements

examine the complete sample, while the proton decoupled ^{13}C studies reported here are restricted to motions within the non-crystalline regions. These are unaffected by the α transition.

The proton spin-spin relaxation decay of unfractionated polyethylene melts have been studied by Folland and Charlesby (58) who interpreted their data in terms of the broad molecular weight distribution. The system was postulated to consist of high molecular weight entangled molecules which coexisted with lower molecular weight species. Thus they argue for the presence of a more mobile component as well as one which is subject to motional constraints due to the entanglements. Other processes could be involved, but very likely for the same molecular reasons, such as a distribution of correlation times (59) and incomplete motional averaging of the dipolar interactions. (60) It does not appear necessary to require the existence of discretely different structural entities, as has been argued. (61)

Further evidence for the direct influence of chain entanglements on the linewidth in the proton spectra for some completely amorphous polymers has been demonstrated by Cohen-Addad and collaborators. (60) (62) (63) Spectral narrowing, as a consequence of residual dipolar broadening, was observed in magic angle spinning experiments, for polyisobutylene, polydimethyl siloxane and *cis* 1,4 polybutadiene. The significant result here is not simply that the resonant linewidth was narrowed upon magic angle spinning but that this effect was only observed over the concentration and molecular weight range where the chains were known to be entangled. For diluted systems, or for the pure polymers whose molecular weights were lower than the critical value for chain entanglement, no influence of the magic angle spinning was observed.

In the amorphous state, therefore, at sufficiently high molecular weights, polymer chains exhibit both liquid-like and solid-like properties from the point of view of NMR measurements. The liquid-like properties are manifested in the high frequency segmental motions, and reflected in the spin-lattice relaxation measurements. The solid-like properties, as indicated by the line broadening, are a result of residual dipolar couplings caused by incomplete motional narrowing. This latter effect is removed as soon as the molecular basis for the entangled system does not exist.

Conclusion

The results discussed above indicate that the further study of the ^{13}C spin relaxation parameters possess the potential to develop our understanding of the structure of the non-crystalline regions of semicrystalline polymers. Significant progress has already been made in relating the spin-lattice relaxation parameters with that of the pure melt. The linewidths, or spin-spin relaxation parameters, of semicrystalline polymers have been

shown to contain contributions from several major sources. The pathways and methods by which to sort these out have been set forth. Of particular importance is the influence of morphology, or crystallite arrangement, rather than the degree of crystallinity on the linewidths, which should be reflected in other properties. An inhomogeneous resonant line is typical of most semicrystalline polymers. A more detailed analysis of the line shapes that have been observed will be discussed elsewhere together with a more detailed discussion of the influence of field strength. (43) A clear picture has not as yet appeared on the influence of crystallinity and morphology on the nuclear Overhauser enhancement factor. More detailed work remains to be done in this area. However, preliminary results indicate that there is a major influence of the level of crystallinity.

Acknowledgement

This work was supported by the National Science Foundation under Grant No. DMR 76-21925.

Literature Cited

1. L. Mandelkern, Morphology of Semicrystalline Polymers, in Characterization of Materials in Research: Ceramics and Polymers, Syracuse University Press, p. 369 (1975).
2. E. W. Fischer, Prog. Colloid and Polymer Sci. 57, 149 (1975).
3. J. H. Magill in Treatise in Material Science, V. 10, Part A, Academic Press, p. 1 (1977).
4. L. Mandelkern, Crystallization of Polymers, McGraw-Hill (1964).
5. L. Mandelkern, Acc. Chem. Res. 9, 81 (1976).
6. S. Go, R. Prud'homme, R. Stein and L. Mandelkern, J. Polym. Sci., Polym. Phys. Ed. 12, 1185 (1974).
7. L. Mandelkern, S. Go, D. Peiffer and R. S. Stein, J. Polym. Sci., Polym. Phys. Ed. 15, 1189 (1977).
8. J. Maxfield and L. Mandelkern, Macromolecules 10, 1141 (1977)
9. L. Mandelkern, J. Phys. Chem. 75, 3909 (1971).
10. V. D. Mochel, J. Macromol. Sci.-Revs. Macromol. Chem. C8, 289 (1972).
11. J. Schaefer, in Topics in Carbon-13 NMR Spectroscopy, Vol. 1, G. C. Levy, ed., Wiley-Interscience, New York, p. 149 (1974).
12. M. W. Duch and D. M. Grant, Macromolecules 3, 165 (1970).

13. J. Schaefer, *Macromolecules* 5, 427 (1972).
14. J. Schaefer, *Macromolecules* 6, 882 (1973).
15. R. A. Komoroski and L. Mandelkern, *J. Poly. Sci., Polym. Symp.* C54, 201 (1976).
16. R. A. Komoroski, J. Maxfield and L. Mandelkern, *Macromolecules* 10, 545 (1977).
17. R. A. Komoroski, J. Maxfield, F. Sakaguchi and L. Mandelkern, *Macromolecules* 10, 550 (1977).
18. J. Schaefer, E. O. Stejskal and R. Buchdahl, *Macromolecules* 8, 291 (1975).
19. J. Schaefer and E. O. Stejskal, *J. Amer. Chem. Soc.* 98, 2035 (1976).
20. H. A. Resing and W. B. Moniz, *Macromolecules* 8, 560 (1975).
21. D. E. Axelson and L. Mandelkern, *J. Polym. Sci., Polym. Phys. Ed.* 16, 1135 (1978).
22. D. E. Axelson and L. Mandelkern, to be published.
23. G. C. Levy, I. R. Peat, R. Rosanske and S. Parks, *J. Magn. Reson.* 18, 205 (1975).
24. D. Canet, G. C. Levy and I. R. Peat, *J. Magn. Reson.* 18, 199 (1975).
25. J. Kowalewski, G. C. Levy, L. F. Johnson and L. Palmer, *J. Magn. Reson.* 26, 533 (1977).
26. S. J. Opella, D. J. Nelson and O. Jardetzky, *J. Chem. Phys.* 64, 2533 (1976).
27. J. G. Fatou and L. Mandelkern, *J. Phys. Chem.* 69, 71 (1965).
28. E. Ergöz, J. G. Fatou and L. Mandelkern, *Macromolecules* 5, 147 (1974).
29. S. Go, F. Kloos and L. Mandelkern, to be published.
30. D. E. Axelson, G. C. Levy and L. Mandelkern, *Macromolecules*, in press.
31. J. D. Hoffman, L. J. Frolen, G. S. Ross and J. I. Lauritzen, Jr., *J. Res. Natl. Bur. Stand., Sect. A* 79, 671 (1975).

32. F. Sakaguchi and L. Mandelkern, unpublished observations.
33. J. Maxfield and L. Mandelkern, *J. Polym. Sci., Polym. Phys. Ed.*, in press.
34. Y. Inoué, A. Nishioka, and R. Chujo, *Makromol. Chem.* 168, 163 (1973).
35. J. Schaefer and D. F. S. Natusch, *Macromolecules* 5, 416 (1972).
36. J. J. Lindberg, I. Sirén, E. Rahkamaa and P. Tormala, *Die Angewandte Makromolekulare Chemie* 50, 187 (1976).
37. D. W. McCall, D. C. Douglass and E. W. Anderson, *J. Polym. Sci.* 59, 301 (1962).
38. C. L. Beatty and M. F. Froix, *Polym. Prepr. Am. Chem. Soc. Div. Polym. Chem.* 16, 628 (1975).
39. T. M. Connor and A. Hartland, *J. Polym. Sci., Polym. Phys. Ed.* 7, 1005 (1969).
40. J. A. Pople, W. G. Schneider and A. Bernstein, High Resolution Nuclear Magnetic Resonance, McGraw-Hill, p. 80 (1959).
41. J. K. Becconsall, P. A. Curnuck and M. C. McIvor, *Appl. Spec. Rev.* 4, 307 (1971).
42. C. P. Poole and H. A. Farrach, Relaxation in Magnetic Resonance, Academic Press (1971).
43. D. E. Axelson, R. A. Komoroski and L. Mandelkern, to be published.
44. N. Bloembergen, E. M. Purcell and R. V. Pound, *Phys. Rev.* 73, 679 (1948).
45. J. Schaefer, *J. Magn. Resonance* 6, 670 (1972).
46. S. Kaufman, W. P. Slichter and D. D. Davis, *J. Polym. Sci., Polym. Phys. Ed.* 9, 829 (1971).
47. J. Schaefer, S. H. Chin and S. I. Weissman, *Macromolecules* 5, 798 (1972).
48. F. Bueche, Physical Properties of Polymers, Interscience, p. 65ff (1962).
49. F. Bueche, *J. Chem. Phys.* 20, 1959 (1952); *ibid.*, 25, 599 (1956).

50. W. W. Graessley, *Adv. Polym. Sci.* 16, 1 (1974).
51. J. D. Ferry, *Viscoelastic Properties of Polymers*, 2nd edition, Wiley (1970).
52. G. C. Berry and T. G. Fox, *Adv. Polym. Sci.* 5, 261 (1968).
53. R. S. Porter and J. F. Johnson, *J. Appl. Polym. Sci.* 3, 194 (1960).
54. R. S. Porter and J. F. Johnson, *Trans. Soc. Rheol.* 6, 107 (1962); *Soc. Plast. Eng. Trans.* 3, 18 (1963).
55. H. Markovitz, T. G. Fox and J. D. Ferry, *J. Phys. Chem.* 66, 1567 (1962).
56. T. P. Yin, S. E. Lovell, and J. D. Ferry, *J. Phys. Chem.* 65, 534 (1961).
57. H. G. Olf and A. Peterlin, *J. Polym. Sci. A-2* 8, 753, 771 (1970).
58. R. Folland and A. Charlesby, *J. Polym. Sci., Polymer Lett. Ed.* 16, 339 (1978).
59. F. Horii, R. Kitamaru and T. Suzuki, *J. Polym. Sci., Polym. Lett. Ed.* 15, 65 (1977).
60. J. P. Cohen-Addad, M. Domard and J. Herz, *J. Chem. Phys.* 68, 1194 (1978).
61. W. L. F. Götz, H. G. Zachmann, *Kolloid-Z.Z. Polym.* 247, 814 (1971).
62. J. P. Cohen-Addad and C. Roby, *J. Chem. Phys.* 63, 3095 (1975).
63. J. P. Cohen-Addad and J. P. Faure, *J. Chem. Phys.* 61, 1571 (1974).

Discussion

J. Guillet, University of Toronto, Ontario: Just one comment about your glass transition temperatures. Surely because the frequency with which you are working is so high, the glass transition temperature will occur at a much lower temperature rather than at a much higher one. Would this not be correct?

D. Axelson: The glass temperature usually increases with increasing frequency. However, in the present problem our conclusions are based on the correlation time, which is a frequency-independent quantity.

J. Guillet: What is the essential frequency of your measurement? Presumably it is not the frequency of the radiation.

D. Axelson: These spectra were obtained at 67.9 MHz, but that's not the problem. We can measure correlation times regardless of the frequency. The correlation time at the glass temperature is very long. From a measurement of the correlation time we should be able to tell whether it is a true glass. In all these cases the correlation times are six to nine orders of magnitude lower than can possibly exist in a glass. For this reason I think the correlation between the NMR measurement and dielectric relaxation and dynamic mechanical do not relate one to one because of the frequency effects in the other measurements.

J. Guillet: But you would expect a frequency effect in this one as well, if only the frequency of the motion itself.

D. Axelson: The relaxation parameters are frequency dependent but not the correlation time.

J. C. Randall, Phillips Petroleum, Oklahoma: I was interested in your linewidth curves vs temperature in which you essentially had the melting point curves. Did you do any freezing point curves?

D. Axelson: These were obtained for branched polyethylene and were similar to the melting point curves. The conditions were such that either direction gave identical linewidths.

J. C. Randall: Yes, I could see that in the low density polyethylene. It would be interesting to compare a system of spherulites vs known morphology for essentially the same crystallinity. What result would those conditions give?

D. Axelson: This would be a very interesting experiment to carry out but technically somewhat difficult for us at present. We have plans to carry out such measurements in the near future.

C. J. Carman, B. F. Goodrich, Ohio: Since T_g is a zero frequency measurement and since the NMR experiment is at a higher frequency, I think T_g would go to a higher value. In other words, your apparent T_g with an NMR measurement would be higher than a T_g as measured with a zero frequency measurement (DSC). Therefore I don't think the numbers you presented are too surprising in view of the fact that you are at a higher frequency. Your T_c should be a pseudofunction of T_g at higher temperatures than of a T_g measured by a thermal measurement.

D. Axelson: Based on correlation time measurements the upper limit T_g 's that we report are frequency-independent. The results would only be surprising to those who argue for a much higher T_g for the polyethylenes.

A. Jones, Clark University, Massachusetts: Is the dominant effect on T_2 morphological and the dynamics of low frequency motions somewhat difficult to extract from T_2 ?

D. Axelson: For the polyethylenes, at least, there is a major effect of morphology on linewidth. This is going to make more difficult a detailed description of the dynamics of the low frequency motion relative to a completely amorphous polymer.

J. Guillet: I thank Dr. Carman for pointing out my error caused by the early hour of the morning, I think. It is quite correct that T_g should be higher and, as I recall from the work on dielectric and other measurements of the glass transition, a

ten degree rise in the glass transition might be expected for every log hertz. This would correspond to a glass transition at about ten to the fourth or fifth cycles per second, which sounds about the right range for the NMR. Fifty degrees' difference would be expected, ten degrees for each order of magnitude change in the frequency. It could correlate quite well with the glass transition temperature.

D. Axelson: As we have already pointed out, the correlation time is frequency-independent. The longest correlation time that we have measured is about 10^{-6} s. Whether the results correlate well with the glass temperature depends on the value one accepts for linear and branched polyethylene. Those values have been a controversial matter.

J. Guillet: It seems to me interest should be focused on only one kind of correlation time and that is the one that relates to the motions of long segments in the polymer. Obviously, the glass transitions are not going to be affected by rotations of methyl groups and phenyl groups. The motion that really relates to the glass transition is whether or not a complete reordering of segments of ten- to fifty-monomer units takes place. This correlates with the glass transition.

D. Axelson: Carbon-13 NMR allows for the measurement of the average correlation time for each individual carbon atom. For the glass temperature problem we are obviously only concerned with the correlation time of the backbone carbons.

C. J. Carman: Earlier in your talk you showed the carbon T_1 data and NOEF for partially crystalline and amorphous polyisoprenes. Was this a natural rubber which had been allowed to crystallize to different degrees or was this a synthetic rubber?

D. Axelson: The sample studied was a synthetic cis-polyisoprene. Its cis-1,4 content was greater than 99%.

C. J. Carman: How was the crystallinity controlled and how was the crystallinity ascertained? As I recall the data, the T_1 's were not affected; neither were the NOEF's. In examining the amorphous region how can one be certain the crystalline region is participating in the data anyway?

D. Axelson: Detailed answers to these questions can be found in *Macromolecules* 10, 545 (1977); *ibid.*, 10, 55 (1977).

P. Sipos, Dupont, Ontario: In the case of polyethylene what was the origin of the sample? Because it makes a difference as far as being catalytic or free radical.

D. Axelson: The low density (branched) polyethylenes were free radical initiated; the linear polymers were derived from commercial sources and purified and characterized as described (*Macromolecules* 10, 550(1977)).

RECEIVED March 13, 1979.

Use of Carbon-13 NMR Analysis in the Characterization of Alternating Copolymers Prepared by Chemical Modifications of 1,4-Polydienes

R. LACAS, G. MAURICE, and J. PRUD'HOMME

Department of Chemistry, University of Montreal, Montreal, Quebec, Canada H3C 3V1

Chemical modifications of unsaturated polymers, especially polydienes, have been investigated for many years in order to derive novel polymers having specific physical and mechanical properties (1). More recently, interest in elastomers with higher stability to oxidative degradation has intensified research into hydrogenation of polydienes (2,3), including polydiene moieties in thermoplastic block and graft copolymers (4). Another feature of chemical modifications of polydienes is their ability to provide a route for preparing novel polymers of particular structures such as head-to-head polymers or alternating copolymers which cannot be prepared by conventional polymerization or copolymerization processes. For example, equimolar alternating copolymers of ethylene with propylene can be obtained by hydrogenating 1,4-polyisoprene (3,5), while head-to-head polypropylene can be obtained by hydrogenating 1,4-poly(2,3-dimethyl-1,3-butadiene) (1,4-polydimethylbutadiene) (5). Hydrohalogenation reactions carried out on 1,4-polydienes may also result in equimolar alternating copolymers of well defined structures (6).

In the present paper we wish to report both ^1H and ^{13}C NMR studies of such alternating copolymers obtained by hydrogenation and hydrohalogenation reactions carried out on 1,4-polydienes prepared with butyllithium in nonpolar solvents. Available for this work were hydrogenated 1,4-polyisoprene and 1,4-polydimethylbutadiene, hydrochlorinated 1,4-polyisoprene and 1,4-polydimethylbutadiene, and hydrobrominated 1,4-polyisoprene. Unlike 1,4-polyisoprene, 1,4-polydimethylbutadiene has symmetrically tetrasubstituted double bonds in its backbone. This situation gives rise to the possibility of threo and erythro diastereoisomerism in the repeating units of the saturated materials. Also of interest, is the possibility of head-to-head and tail-to-tail additions of hydrogen halide in 1,4-polydimethylbutadiene. Both these features were investigated on the basis of the ^{13}C chemical shift substituent effects derived from spectra measured on suitable model compounds, including hydrohalogenated 1,4-polyisoprene.

0-8412-0505-1/79/47-103-215\$05.25/0
© 1979 American Chemical Society

Preparation and 220MHz ^1H NMR Characterization of the Materials

Starting Materials. The polydienes submitted to the chemical modifications were polyisoprene and polydimethylbutadiene samples prepared by anionic polymerization using *sec*-butyllithium as initiator and hydrocarbons as solvents (5,6). The polyisoprene sample was prepared at 25°C using benzene as solvent (6). Its microstructure determined by ^1H NMR spectroscopy was 71% *cis*-1,4, 22% *trans*-1,4, and 7% 3,4. Its number average molecular weight was 8.6×10^4 . The polydimethylbutadiene sample was prepared at 60°C using cyclohexane as solvent (5). Its microstructure determined by ^1H NMR spectroscopy was 74% *trans*-1,4, 23% *cis*-1,4, and 3% 1,2. Its number average molecular weight was 4.3×10^4 .

Hydrogenation Reactions. The hydrogenation reactions were carried out on 0.5% polymer solutions in cyclohexane by using coordination catalysts made by the reaction of triethylaluminum with the cobalt (II) salt of 4-cyclohexylbutanoic acid. For that purpose, catalyst solutions having an aluminum/cobalt molar ratio of 4 were prepared under a nitrogen atmosphere by slowly adding a molar solution of triethylaluminum to a 2×10^{-2} M solution of the cobalt salt, both in cyclohexane. The catalyst solutions were added to the polydiene solutions under a nitrogen atmosphere following which hydrogen was bubbled through the solutions at a constant pressure of 4 atm for a period of 2h, at 50°C. Under these experimental conditions, quantitative hydrogenation of the 1,4-polyisoprene sample was obtained using 5 mol % of catalyst based on unsaturated monomer units, whereas nearly quantitative hydrogenation of the 1,4-polydimethylbutadiene sample required 30 mol % of catalyst. Completion of the reactions is demonstrated in Figures 1 and 2 where the upfield regions of the 220 MHz ^1H NMR spectra measured before and after hydrogenation are presented for the 1,4-polyisoprene and the 1,4-polydimethylbutadiene samples, respectively.

From Figure 1, one can see that no trace of residual unsaturated units is detectable in the ^1H spectrum of the hydrogenated 1,4-polyisoprene sample. On the other hand, the spectrum exhibits a well resolved methyl doublet ($J = 6.5$ Hz) centered at 0.92 ppm. From Figure 2, one can see that the resonances of the unsaturated units are not completely absent in the ^1H spectrum of the hydrogenated 1,4-polydimethylbutadiene sample. However relative intensity measurements made on the spectrum indicate a degree of saturation higher than 98% for this polymer. Interestingly, the methyl resonance of the hydrogenated 1,4-polydimethylbutadiene sample appears as two doublets centered at 0.90 ppm indicating that two different configurations can be distinguished for this polymer. In fact, assuming the same coupling constant, $J = 6.5$ Hz, as that observed in the spectrum of hydrogenated 1,4-polyisoprene, one can interpret the methyl resonances of hydrogenated 1,4-polydimethylbutadiene as the

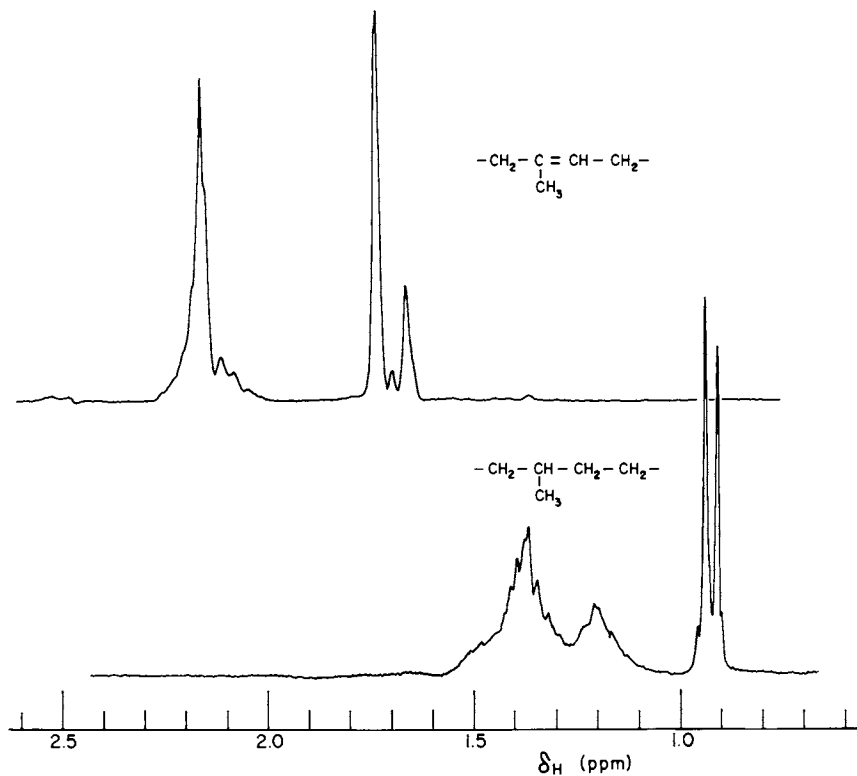


Figure 1. Upfield region of the 220 MHz ^1H NMR spectra made before and after the hydrogenation of the 1,4-polyisoprene sample. Chlorobenzene solutions at 100°C with TMS as reference.

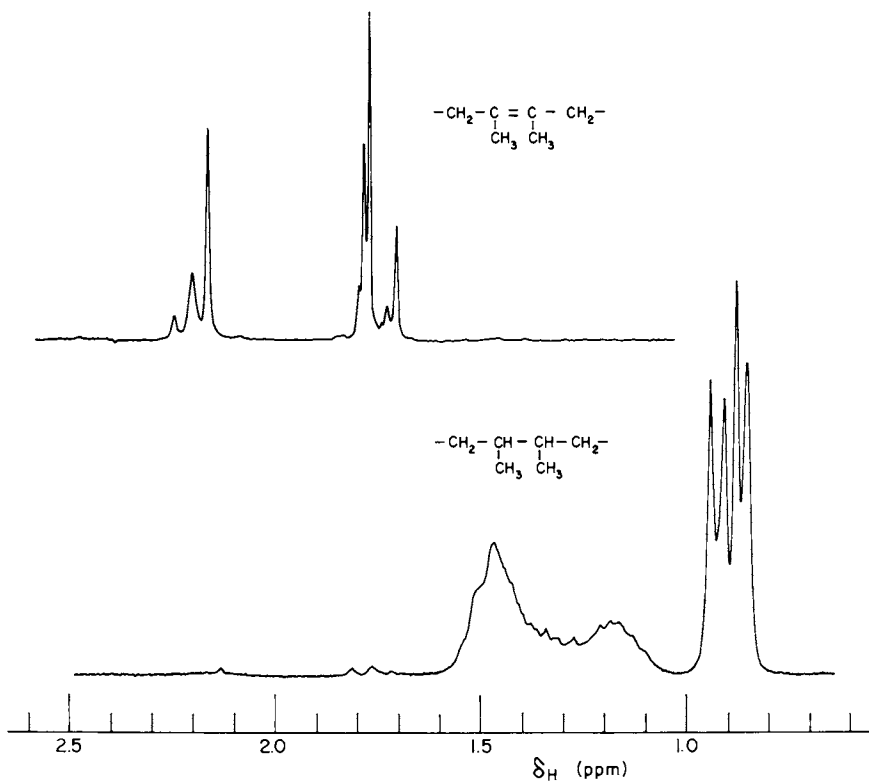


Figure 2. Upfield region of the 220 MHz ^1H NMR spectra made before and after the hydrogenation of the 1,4-polydimethylbutadiene sample. Chlorobenzene solutions at 100°C with TMS as reference.

juxtaposition of two doublets with a chemical shift difference of 13.5 Hz. One of these doublets would correspond to the threo configuration and the other to the erythro configuration of the saturated repeating units.

Hydrohalogenation Reactions. Both the hydrochlorination and hydrobromination of the 1,4-polyisoprene sample were conducted at 25°C on 1% polymer solutions in toluene. The solutions were first purged with dry nitrogen after which the dry hydrogen halide was bubbled through the reaction system at a pressure slightly above atmospheric for a period of 24 h. Figure 3 shows the 220 MHz ¹H NMR spectra measured on the two hydrohalogenated products. In either ¹H spectrum quantitative hydrohalogenation is indicated by the loss of the isoprene unit methylene resonances at 2.2 ppm. On the other hand, the fact that the methyl resonances appear as sharp singlets at 1.53 ppm for the hydrochlorinated product and at 1.69 ppm for the hydrobrominated product indicates the exclusiveness of Markownikoff's rule for the addition of either hydrogen halide to the repeating units of 1,4-polyisoprene.

The hydrochlorination of the 1,4-polydimethylbutadiene sample was conducted using the same procedure as for 1,4-polyisoprene, except that it was necessary to increase the hydrogen chloride pressure to 4 atm in order to obtain a quantitative saturation of this polymer. Figure 4 shows the 220 MHz ¹H NMR spectrum measured on the product. Owing to the possible superposition of the ¹H resonances of the hydrochlorinated units with those of the unsaturated units (see Figure 2), estimation of the degree of conversion from the ¹H spectrum alone is difficult. However, elemental analysis of the product indicated a degree of hydrochlorination close to 99%. From Figure 4, one can see that the two methyl resonances expected for hydrochlorinated 1,4-polydimethylbutadiene appear as pairs of complex signals centered at 1.1 and 1.5 ppm, respectively. The signals centered at 1.1 ppm arise from the methyl groups on the tertiary carbons. As for hydrogenated 1,4-polydimethylbutadiene, it is reasonable to interpret these signals as the juxtaposition of two doublets corresponding to threo and erythro configurations of the repeating units although, in the present case, the resolution of the two doublets is not as good as for the hydrogenated polymer. The other two signals centered at 1.5 ppm arise from the methyl groups on the quaternary carbons. Apparent in either signal is a fine structure which suggests sensitivity to the position of the chlorine atoms in the neighbouring units. The same effect may also explain the lack of resolution observed for the methyl resonances centered at 1.1 ppm. This point is clarified in the next section where the same material is characterized by ¹³C NMR spectroscopy.

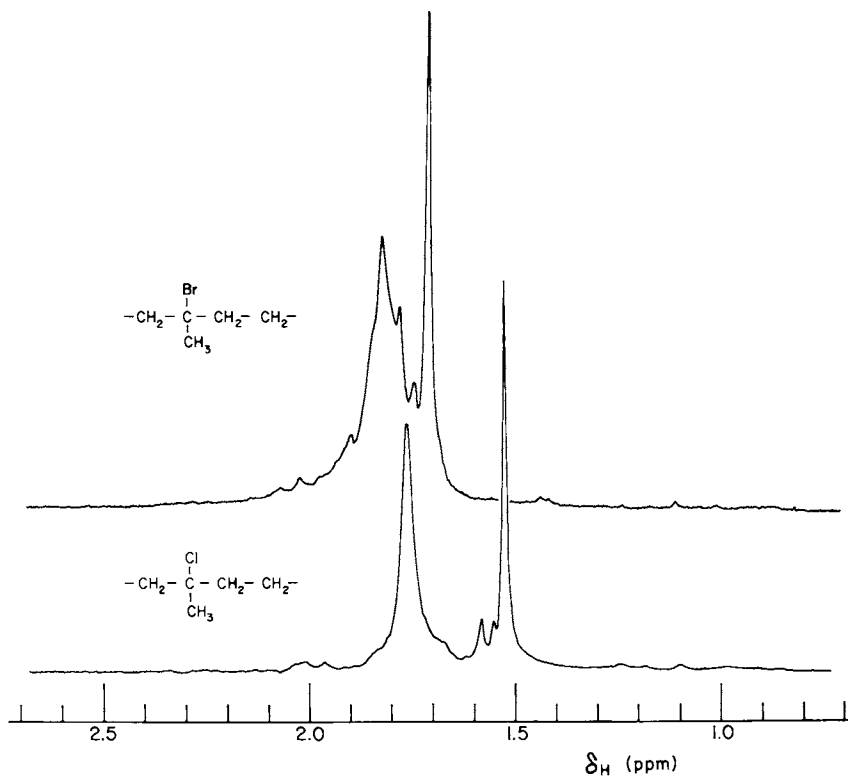


Figure 3. A 220-MHz ^1H NMR spectra of the hydrohalogenated 1,4-polyisoprene samples. Chlorobenzene solutions at 100°C with TMS as reference.

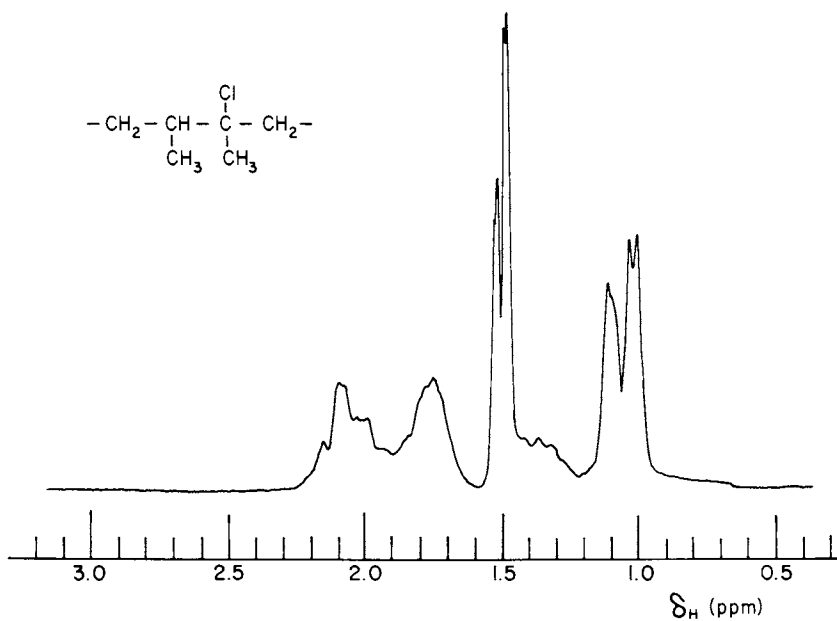


Figure 4. A 220-MHz ^1H NMR spectrum of the hydrochlorinated 1,4-polydimethylbutadiene sample. Chlorobenzene solution at 100°C with TMS as reference.

^{13}C NMR Characterization of the Materials

Hydrogenated 1,4-polyisoprene. Figure 5 shows the 22.6 MHz ^{13}C NMR spectrum of the hydrogenated 1,4-polyisoprene sample. The spectrum is identical to those already reported in the literature for hydrogenated natural rubber (7) as well as for hydrogenated synthetic *cis*-1,4-polyisoprene (8). It consists of four main signals which are identified in Figure 5. The spectrum also exhibits much weaker signals which arise from the hydrogenated 3,4 units present in the polymer. As shown in Table I, the chemical shifts of the four main resonances observed in the spectrum of Figure 5 can be satisfactorily predicted by the empirical equation derived by Lindeman and Adams (9) for low molecular weight linear and branched alkanes. Also listed in Table I are the chemical shifts measured by Carman et al. (8) for the equivalent carbon atoms in the model compound 2,6,10,14-tetramethylpentadecane. Interestingly, the largest difference between the chemical shifts measured for hydrogenated 1,4-polyisoprene and those predicted from the model compound resonances does not exceed 0.2 ppm.

Hydrogenated 1,4-polydimethylbutadiene. Figure 6 shows the 22.6 MHz ^{13}C NMR spectrum of the hydrogenated 1,4-polydimethylbutadiene sample. The identification of methyl, methylene, and methine carbon resonances was made by off-resonance decoupling experiments. Two distinct signals occur for each type of carbon suggesting discrimination between *threo* and *erythro* configurations for the repeating units. Such a discrimination was also apparent in the 220 MHz ^1H NMR spectrum (Figure 2) which exhibited two distinct methyl doublets centered at 0.90 ppm. This interpretation is substantiated by comparing the ^{13}C NMR spectrum of Figure 6 with that measured by Lindeman and Adams (9) for a mixture of the two diastereoisomers of 3,4-dimethylhexane. The latter spectrum (not shown) also exhibits two distinct resonances for all carbons but the terminal methyl carbons which are separated from the tertiary carbons by one methylene unit. More interesting is the fact that very similar chemical shift differences are observed for each pair of signals in either case: 2 ppm for both the methyl and methylene resonances and 1 ppm for the methine resonances.

Hydrogenated 1,4-polydimethylbutadiene may be considered as either an alternating copolymer of ethylene and 2-butene or as a head-to-head polypropylene. Such alternating structures of ethylene and 2-butene monomer units can be obtained directly by copolymerizing the two olefins with Ziegler-Natta catalysts. For instance, Natta et al. (10) reported that *cis*-2-butene can be copolymerized with ethylene using $\text{VC1}_4/\text{AlR}_3$ catalyst systems to yield crystalline alternating copolymers of the *erythro*-diisotactic structure. The ^{13}C NMR spectrum of this latter copolymer has been reported by Zambelli et al. (11). As shown

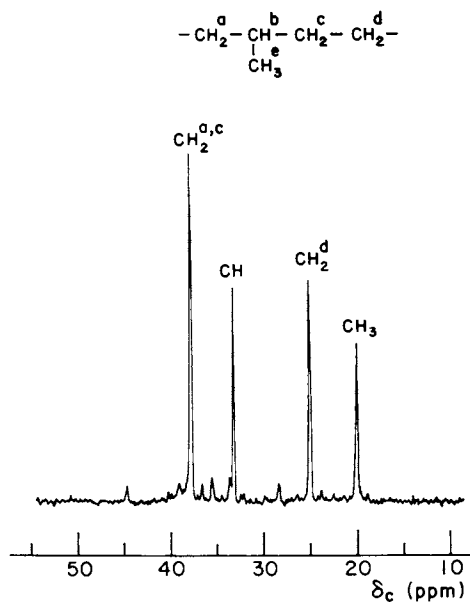


Figure 5. Proton noise-decoupled 22.6-MHz C-13 NMR spectrum of the hydrogenated 1,4-polyisoprene sample. Perdeuteriobenzene solution at 25°C with TMS as internal reference. Approximately 5000 pulses with an acquisition time of 0.7 sec and a flip angle of 30°.

Table I.

Observed and Predicted ^{13}C Chemical Shifts*
for Hydrogenated 1,4-Polyisoprene

Carbon**	Chemical Shifts (ppm)		
	Observed	Predicted	
		Lindeman-Adams	Model compound***
CH_3	20.1	19.6	20.2
CH_2^{d}	25.1	24.6	25.3
CH	33.4	32.5	33.3
$\text{CH}_2^{\text{a,c}}$	38.1	36.9	38.0

* All chemical shifts are in ppm downfield from TMS.

** Identified in Figure 5.

*** Measured by Carman et al. (8) on 2,6,10,14-tetramethylpentadecane.

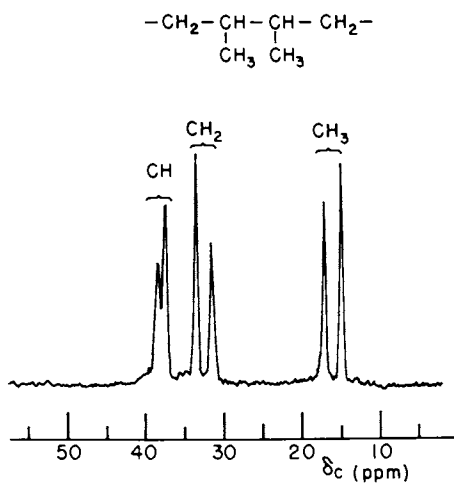


Figure 6. Proton noise-decoupled 22.6-MHz C-13 NMR spectrum of the hydrogenated 1,4-polydimethylbutadiene sample. Perdeuteriobenzene solution at 25°C with TMS as internal reference. Approximately 9000 pulses with an acquisition time of 0.7 sec and a flip angle of 30°.

in Table II, comparison of the chemical shifts measured by Zambelli et al. with those observed for the hydrogenated 1,4-polydimethylbutadiene sample allows the assignments of the erythro signals in the latter. The results are indicated in Table II beside the chemical shifts of the observed signals. One can see that the largest difference in chemical shift between the two sets of matched signals does not exceed 0.2 ppm. Note that an additional methylene carbon resonance was observed at 30.3 ppm (downfield from TMS) in the spectrum of the copolymer studied by Zambelli et al. They attributed this resonance to internal methylene carbons in sequences of more than one ethylene unit because the copolymer contained more than 50 mol % of ethylene units.

On the basis of the assignments given in Table II, one may conclude that the threo structure is slightly predominant in the hydrogenated 1,4-polydimethylbutadiene sample. This result taken in conjunction with the microstructure of the starting material (74% trans-1,4, 23% cis-1,4, and 3% 1,2) indicates a nonstereospecific addition of hydrogen to the unsaturated 1,4 units of polydimethylbutadiene.

Hydrochlorinated and Hydrobrominated 1,4-Polyisoprene.

Figure 7 shows the 22.6 MHz ^{13}C NMR spectra of both the hydrochlorinated and hydrobrominated 1,4-polyisoprene samples. Like the spectrum of hydrogenated 1,4-polyisoprene, either spectrum in Figure 7 exhibits four major signals only, the assignments of which are indicated in the Figure. Either spectrum also shows much weaker signals which arise from the hydrohalogenated 3,4 units present in the material. The fact that only four major carbon resonances are observed in the spectra of Figure 7 results from the exclusiveness of Markownikoff's rule for the hydrogen halide addition to 1,4-polyisoprene, a characteristic which was previously evidenced from the 220 MHz ^1H NMR spectra (Figure 3). This situation is of interest, since such halogen atoms regularly introduced in a polyolefinic chain, particularly bromine atoms, might be used as intermediates for subsequent chemical modifications based upon substitution reactions.

Comparing the ^{13}C NMR spectrum data of the hydrohalogenated 1,4-polyisoprene samples with those of the hydrogenated sample allows one to evaluate the shielding contributions produced by the halogen substituents in the surrounding α , β , and γ positions in the polymer chains. The results of this comparison are summarized in Table III (upper part) where the substituent effects upon replacement of the hydrogen atom on the tertiary carbon by either a chlorine or a bromine atom are listed for the 1,4-polyisoprene derivatives. One can see that the differences between the chlorine and the bromine substituent effects range from 1.0 ppm for the γ effect to 1.6 ppm for the α effect. Interestingly, the set of shift parameters in the upper part of Table III is very close to that which one can compute from

Table II.

¹³C Chemical Shifts* of Hydrogenated 1,4-Polydimethylbutadiene
Compared to those of cis-2-Butene-Ethylene
Alternating Erythrodiisotactic Copolymer

Carbon	Hydrogenated 1,4-Polydimethylbutadiene	cis-2-Butene-Ethylene Alternating Copolymer**
CH ₃	14.7 (t)*** 17.0 (e)	16.8
CH ₂	31.4 (e) 33.5 (t)	30.3 31.5
CH	37.5 (t) 38.5 (e)	38.6

* All chemical shifts are in ppm downfield from TMS.

** Data from Zambelli et al. (11). Original chemical shifts were converted downfield from HMDS to downfield from TMS using $\delta(\text{HMDS}) = 2.0$ ppm.

*** Assignment of threo (t) and erythro (e) signals.

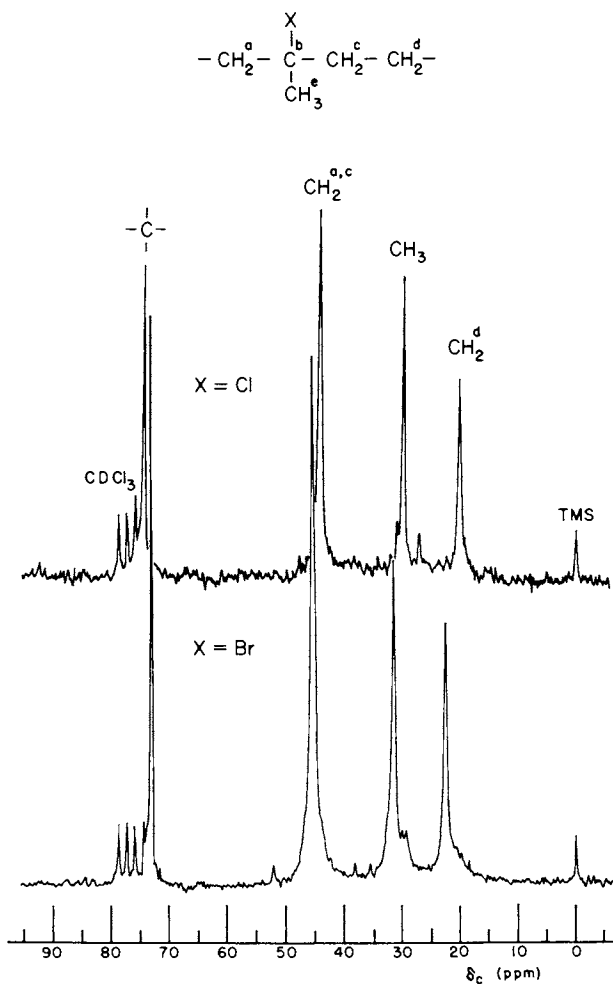


Figure 7. Proton noise-decoupled 22.6-MHz C-13 NMR spectra of the hydrohalogenated 1,4-polyisoprene samples. Deuteriochloroform solutions at 25°C with TMS as internal reference. Approximately 5000 pulses with an acquisition time of 0.7 sec and a flip angle of 30°.

Table III.

Substituent Effects upon Replacement of Hydrogen by Halogen

in		$\begin{array}{cccc} & & X & & \\ & \beta' & \alpha & \beta' & \gamma \\ - & \text{CH}_2 & - \text{C} & - \text{CH}_2 & - \text{CH}_2 - \\ & & & & \\ & & \beta & & \\ & & \text{CH}_3 & & \end{array}$		
X	α	β	β'	γ
Cl	+ 40.9	+ 9.7	+ 6.1	- 2.4
Br	+ 39.3	+ 11.3	+ 7.2	- 1.4

in		$\begin{array}{cccc} & & X & & \\ & \beta & \alpha & \beta' & \gamma \\ \beta & \text{CH}_3 & - \text{C} & - \text{CH}_2 & - \text{CH}_3 \\ & & & & \\ & & \beta & & \\ & & \text{CH}_3 & & \end{array}$		
X	α	β	β'	γ
Cl	+ 40.8	+ 9.4	+ 7.0	- 2.3
Br	+ 37.9	+ 11.6	+ 8.4	- 1.1

standard source spectra for 2-methylbutane and its 2-chloro and 2-bromo derivatives. For the sake of comparison, this latter set of shift parameters is listed in the lower part of Table III.

From Table III, one can see that a common feature in both sets of shift parameters (that for 1,4-polyisoprene derivatives and that for 2-methylbutane derivatives) is a higher β effect for a methyl carbon (denoted β in Table III) than for a methylene carbon (denoted β' in Table III). The difference between β and β' effects is close to 4 ppm for the polymers whereas it is close to 3 ppm for the model compounds. On the other hand, one can see that there is no significant difference in the γ effects whether the carbon under consideration is a methyl or methylene carbon. The empirical shift parameters calculated from the ^{13}C NMR data of the 1,4-polyisoprene derivatives will provide, hereafter, a basis for assigning the carbon resonances observed in the spectrum of hydrochlorinated 1,4-polydimethylbutadiene, which like hydrochlorinated 1,4-polyisoprene has quaternary carbons substituted by one chlorine atom.

Hydrochlorinated 1,4-polydimethylbutadiene. Figure 8 shows the 22.6 MHz ^{13}C NMR spectrum of the hydrochlorinated 1,4-polydimethylbutadiene sample. Identification of methyl, methylene, methine, and quaternary carbon resonances was made by off-resonance decoupling experiments. It can be seen that all resonances but the methyl resonance at 15.1 ppm, and the methine resonance at 44.3 ppm, occur as pairs of signals. On the other hand, the off-resonance spectrum (not shown) revealed the presence of a methylene resonance overlapping the pair of methyl signals centered at 27.0 ppm. In fact, the spectrum in Figure 8 contains four distinct methylene resonances centered at 27.1, 31.1, 37.0, and 41.4 ppm. This feature arises because there is no selective induction mechanism for controlling the position of the chlorine atom in the addition of hydrogen chloride to symmetrically substituted double bonds such as those in 1,4-polydimethylbutadiene. This situation leads to the possibility of head-to-tail, head-to-head, and tail-to-tail arrangements of two consecutive hydrochlorinated 1,4-units. These three arrangements are depicted in Figure 8. They show four chemically nonequivalent methylene carbons which are denoted as CH_2^{C} , CH_2^{d} , $\text{CH}_2^{\text{C}'}$, and $\text{CH}_2^{\text{d}'}$.

The chemical shifts of the four methylene carbons depicted in Figure 8 have been calculated using the empirical shift parameters derived from the preceding study on hydrochlorinated 1,4-polyisoprene. They are listed in Table IV together with the chemical shifts predicted for the other carbon atoms in hydrochlorinated 1,4-polydimethylbutadiene. All the calculations were based on the mean values of the chemical shifts observed for the threo and erythro diastereoisomers in hydrogenated 1,4-polydimethylbutadiene, i.e. 15.9 ppm for CH_3 , 32.5 ppm for CH_2 , and 38.0 ppm for CH. Also listed in Table IV are the experimental chemical shifts. The complete assignments given in both Table IV and Figure 8 were made by fitting the observed methyl and methylene

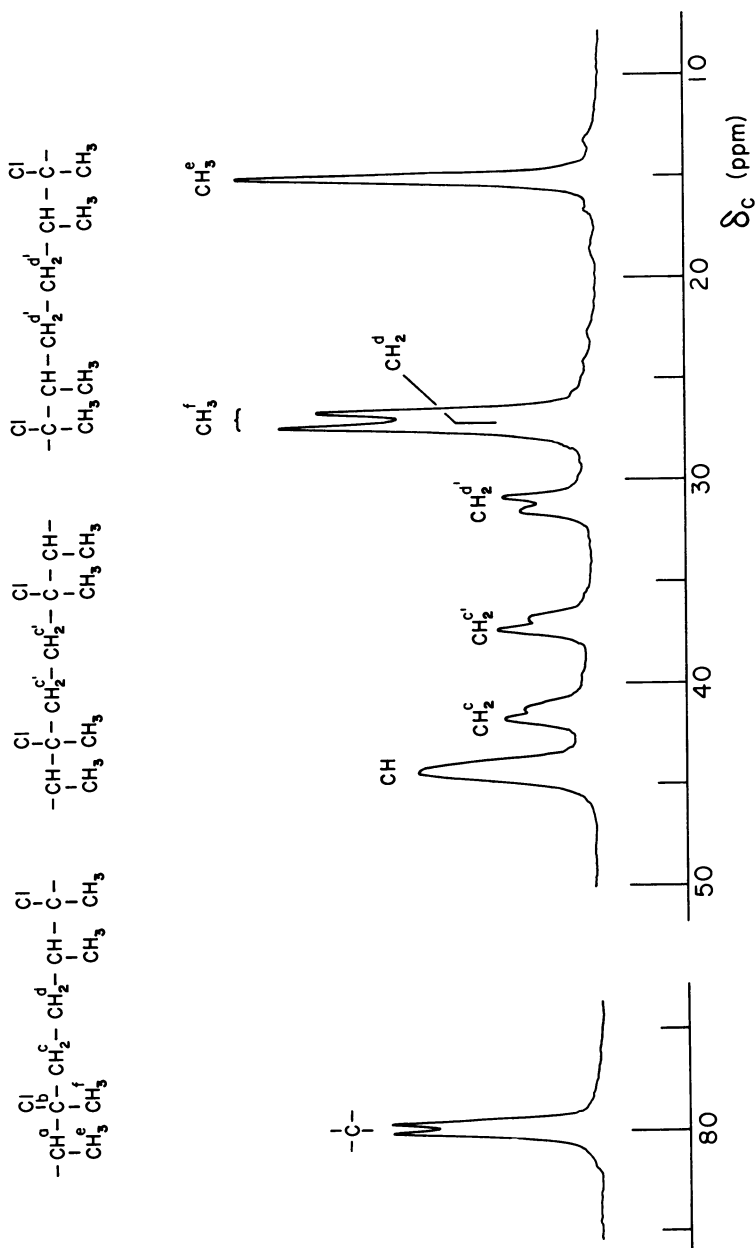


Figure 8. Proton noise-decoupled 22.6-MHz C-13 NMR spectrum of the hydrochlorinated 1,4-polydimethylbutadiene sample. CD_2Cl_2 solution at 25°C with TMS as internal reference. Approximately 15000 pulses with an acquisition time of 1.6 sec and a flip angle of 90°.

Table IV.

Observed and Predicted ^{13}C Chemical Shifts* for
Hydrochlorinated 1,4-Polydimethylbutadiene

Carbon**	Chemical Shifts (ppm)	
	Observed	Predicted
CH_3^e	15.1	13.4
CH_3^f	26.6 27.3	25.5
CH_2^d	27.1	27.6
$\text{CH}_2^{d'}$	30.8 31.5	30.1
$\text{CH}_2^{c'}$	36.7 37.3	36.2
CH_2^c	41.2 41.7	38.6
CH	44.3	44.1
$\begin{array}{c} \\ -\text{C}- \\ \end{array}$	79.6 80.1	78.9

* All chemical shifts are in ppm downfield from TMS.

** Identified in Figure 8.

carbon chemical shifts to the predicted shifts. From Table IV, one can see that a fairly good fitting was obtained for all resonances except the methyl resonance observed at 15.1 ppm and the methylene resonance centered at 41.5 ppm. The difference between the predicted and observed shifts is 1.7 ppm for the former and 3 ppm for the latter.

Several carbon resonances occur as pairs of signals in the spectrum of Figure 8. As for hydrogenated 1,4-polydimethylbutadiene, this may be attributed to the presence of repeating units having threo and erythro configurations in hydrochlorinated 1,4-polydimethylbutadiene. Note that a similar splitting into pairs of signals was also observed for the two methyl resonances in the 220 MHz ^1H NMR spectrum of the same material (Figure 4). In contrast with the ^1H spectrum, only the methyl carbon on the quaternary carbon gives rise to a pair of signals in the ^{13}C spectrum. The other methyl resonance which arises from the methyl group on the tertiary carbon appears as a single signal at 15.1 ppm in the ^{13}C spectrum. The fact that this latter ^{13}C resonance does not exhibit threo and erythro configurational sensitivity might be explained by a fortuitous magnetic equivalence due to steric interactions of the methyl group with the three γ substituents on the quaternary carbon.

Quantitative analysis of the 22.6 MHz ^{13}C NMR spectrum in Figure 8 yields the following information concerning the microstructure of the hydrochlorinated 1,4-polydimethylbutadiene sample. First, the population of one of the diastereoisomers, threo or erythro, is slightly predominant in the material. This is substantiated by the relative intensities of the signals which occur as pairs in the spectrum of Figure 8. This is also supported by the relative intensities of the two methyl doublets observed at 1.1 ppm in the 220 MHz ^1H NMR spectrum of Figure 4. Second, a random placement of the chlorine atoms occurred in the course of the hydrochlorination reaction. This latter result is based on the nearly equal intensities measured for the three methylene carbon resonances directly observable in the spectrum of Figure 8. Note that a fourth methylene carbon resonance was detected at 27.1 ppm by off-resonance decoupling experiments. As shown previously, this methylene resonance at 27.1 ppm and that observed at 41.4 ppm each arise from one of the two chemically nonequivalent methylene carbons in head-to-tail arrangements of two consecutive hydrochlorinated units. On the other hand, the methylene resonances observed at 31.1 and 37.0 ppm each arise from the two chemically equivalent methylene carbons in tail-to-tail and head-to-head arrangements, respectively. Therefore the equal intensities measured for the three methylene resonances observed at 31.1, 37.0, and 41.4 ppm indicate the 1:2:1 proportions of head-to-head, head-to-tail, and tail-to-tail arrangements expected for a random placement of the chlorine atoms in the course of the hydrochlorination reaction.

Acknowledgments

This work was supported by the National Research Council of Canada and the Quebec Ministry of Education. The authors wish to thank the Canadian 220 MHz NMR Centre for making the proton NMR measurements and Mr. R. Mayer for the carbon NMR spectra.

Literature Cited

1. E.M. Fettes, "Chemical Reactions of Polymers", Interscience Publ., New York, 1964, Chap. II.
2. J.C. Falk, J. Polym. Sci., Part A-1, 9, 2617 (1971).
3. L.A. Mango, and R.W. Lenz, Makromol. Chem., 163, 13 (1973).
4. R.C. Jones, U.S. Pat. 3,431,323 (March 4, 1969).
5. D. Khlok, Y. Deslandes, and J. Prud'homme, Macromolecules, 9, 809 (1976).
6. A. Tran and J. Prud'homme, Macromolecules, 10, 149 (1977).
7. Y. Tanaka, H. Sato, A. Ogura, and I. Nagoya, J. Polym. Sci., Polym. Chem. Ed., 14, 73 (1976).
8. C.J. Carman, A.R. Tarpley, and J.H. Goldstein, Macromolecules, 6, 719 (1973).
9. L.P. Lindeman, and J.Q. Adams, Anal. Chem., 43, 1245 (1971).
10. G. Natta, G. Dall'Asta, G. Mazzanti, I. Pasquon, A. Valvassori, and A. Zambelli, J. Am. Chem. Soc., 83, 3343 (1961).
11. A. Zambelli, G. Gatti, S. Sacchi, W.O. Crain, and J.D. Roberts, Macromolecules, 4, 475 (1971).

Discussion

J. Randall, Phillips Petroleum, Oklahoma: I was interested in the β and β' effects for the halogen substituents. Have you looked at the Grant and Paul approach in which they introduced corrective terms? The change from a tertiary to a quaternary carbon would cause conformational changes. Grant and Paul faced the same problem when looking at methyl group substitutions. I was wondering if the differences in the β and β' terms might disappear if you did this. Any conformational contribution might disappear. It would be interesting to see how close the two β terms come. I found this to work on the substituent effect for an aromatic ring. I introduced corrective terms and got the same β or α . It may not happen here because the conformational change may not be predicted by the simple alkyl shifts.

J. Prud'homme: We did not make the corrections. This is a good suggestion.

G. Babbitt, Allied Chemical Corp., N.J.: How do you know that a methylene resonance was under the methyl signals in the ^{13}C spectrum of hydrochlorinated 1,4-polydimethylbutadiene?

J. Prud'homme: Through the use of an off-resonance measurement. The methyl signals appeared as two juxtaposed quartets in the middle of which we could see a methylene resonance.

G. Babbitt. In other words the methyl signals opened by going to quartets and revealed the triplet underneath.

J. Prud'homme: Exactly.

G. Babbitt: In the same spectrum, you showed pairs of methylene signals and attributed the doubling to erythro and threo structures of the H-Cl units. Three units were pictured: an ethylene unit in the center with H-Cl containing units on either end. It is possible that both ends can be erythro, or both ends can be threo, or be mixed. Yet, only doublets are obtained. It seems to me there should be more multiplicity in the methylene signals.

J. Prud'homme: I think here the same situation arises as with hydrogenated 1,4-polydimethylbutadiene. It would appear that when two methylene units separate two chiral carbons, each methylene unit shows little sensitivity to the meso and racemic configurations of the two adjacent chiral carbons. Carman et al. have reported spectra measured on alkanes which show that when two tertiary chiral carbons are separated by two methylene units, the difference in the chemical shifts of the methylene units in the meso and racemic configurations is close to 0.3 ppm. It was not possible to observe this kind of resolution in the spectra of the present polymers. Only a broadening effect occurred.

RECEIVED March 13, 1979.

Carbon-13 NMR Studies on the Cationic Polymerization of Cyclic Ethers

G. PRUCKMAYR and T. K. WU

E. I. du Pont de Nemours & Co., Inc., Experimental Station, Wilmington, DE 19898

The cationic ring-opening polymerization of cyclic ethers has been the subject of many recent investigations (1,2,3,4). Nuclear magnetic resonance (NMR) methods, particularly carbon-13 techniques, have been found most useful in studying the mechanism of these polymerizations (5). In the present review we would like to report some of our recent work in this field.

The first part of this report will illustrate how ^{13}C -NMR has been utilized in the elucidation of the polymerization mechanisms of cyclic ethers. In the second part, quantitative applications of ^{13}C -NMR for determinations of thermodynamic and kinetic constants will be discussed. The last section deals with possible applications of quantitative ^{13}C -NMR analysis in copolymerization of cyclic ethers.

EXPERIMENTAL

Tetrahydrofuran (THF) and oxepane (OXE) were distilled from CaH_2 prior to use. All other reagents and solvents are commercially available in reagent grade purity and were used without further purification.

The proton noise-decoupled ^{13}C -NMR spectra were obtained on a Bruker WH-90 Fourier transform spectrometer operating at 22.63 MHz. The other spectrometer systems used were a Bruker Model HFX-90 and a Varian XL-100. Tetramethylsilane (TMS) was used as internal reference, and all chemical shifts are reported downfield from TMS. Field-frequency stabilization was maintained by deuterium lock on external or internal perdeuterated nitromethane. Quantitative spectral intensities were obtained by gated decoupling and a pulse delay of 10 seconds. Accumulation of 1000 pulses with phase alternating pulse sequence was generally used. For "relative" spectral intensities no pulse delay was used, and accumulation of 200 pulses was found to give adequate signal-to-noise ratios for quantitative data collection.

A calibration curve was obtained from ^{13}C -NMR spectra of a series of polytetramethylene ether (PTME)-THF/ CH_3NO_2 solutions at

different concentrations and temperatures. The PTME was obtained by polymerization of THF with $\text{Me}_3\text{O}^+\text{BF}_4^-$ in CH_3NO_2 (molar ratio $\text{THF}:\text{Me}_3\text{OBF}_4:\text{CH}_3\text{NO}_2 = 1.36:0.1:1.07$) under conditions similar to the subsequent kinetic study. The reaction mixture was quenched with MeONa/MeOH , and the polymer isolated by removal of unreacted monomer and solvent under vacuum, and extraction of the residue with ether. After isolation, the resulting dimethoxypolytetramethylene ether $\text{MeO}-\left[\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{O}\right]_n-\text{Me}$ ($M_n \approx 600$) was used directly in the calibration mixtures.

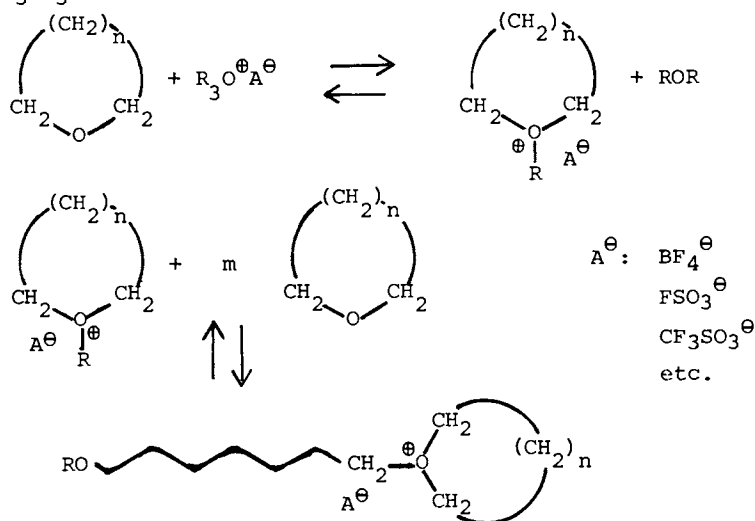
RESULTS AND DISCUSSION

I. Polymerization of Cyclic Ethers

General Mechanism and Spectra. The cationic ring-opening polymerization of cyclic ethers has long been known to involve oxonium ions (6). For THF it is well recognized that under certain conditions all the reactions are reversible and that limiting conversions are reached at given temperatures. The polymerization of THF has therefore been frequently characterized as a "living" polymerization (7).

In the initial step of the polymerization, a cyclic oxonium ion is formed by transfer of an alkyl group from the initiator to the cyclic ether. Propagation occurs by $\text{S}_\text{N}2$ attack of a monomer molecule at a ring α -methylene position of the cyclic tertiary oxonium ion, followed by opening of the oxonium ring and formation of a new cyclic oxonium ion.

The initiator may be a Lewis acid, an oxonium salt or precursor (8), or an ester of a strong acid (9). The anion A^- in the formula scheme below may designate, e.g., tetrafluoroborate (BF_4^-), fluorosulfonate (FSO_3^-), trifluoromethyl sulfonate (CF_3SO_3^-), etc.

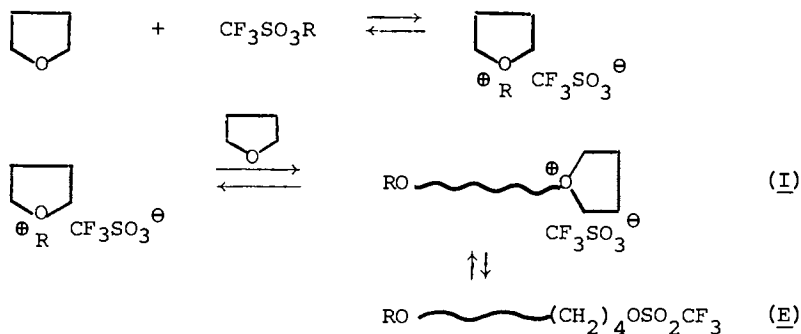


Kinetic study of this reaction usually requires sampling the polymerizing mixture and analyzing for the concentrations of the various reaction species at different polymerization times. Vofsi and Tobolsky in 1965 reported the use of radioactively tagged initiator (10), while Saegusa and coworkers in 1968 developed a "phenoxy end-capping" method in which the oxonium ion is trapped with sodium phenoxide and the derived phenyl ether at the polymer chain end quantitatively determined by UV spectrophotometry (11).

We have been investigating similar model polymerizations. In 1973 we reported the use of ^1H -NMR spectroscopy for the identification of the various species in such a polymerization (12). We found this method to be extremely useful for kinetic in-situ study of polymerizations without disturbing the system. Subsequently we applied ^{19}F -NMR to follow the polymerization initiated with catalysts containing fluorine atoms (13). At the same time the superior resolution of ^{13}C -NMR was exploited to investigate the various proposed equilibria in the polymerization of cyclic ethers (5).

Figure 1 shows the proton noise-decoupled ^{13}C -NMR spectrum of a polytetrahydrofuran (polytetramethylene ether glycol, PTMEG) dissolved in THF. In this spectrum the carbons numbered 1, 2 and 3 which are α to the oxygen appear at lower field than the β -carbons labeled as 4, 5 and 6. The carbon atoms in the polymer are clearly resolved from the corresponding carbons of the THF monomer. The fact that carbons 3 and 4 near the hydroxyl end-groups can be easily identified shows the excellent resolution of this technique.

Polymerization Equilibria. As mentioned earlier, esters of strong acids, e.g. trifluoromethane sulfonic acid ("triflates"), are excellent initiators for the polymerization of THF. With such initiators, however, a complication arises. In addition to the normal propagation \rightleftharpoons depropagation equilibria of oxonium ions, Smith and Hubin postulated that the macroion (I) may also convert into a corresponding nonpolar macroester (E) by attack of the anion (14).



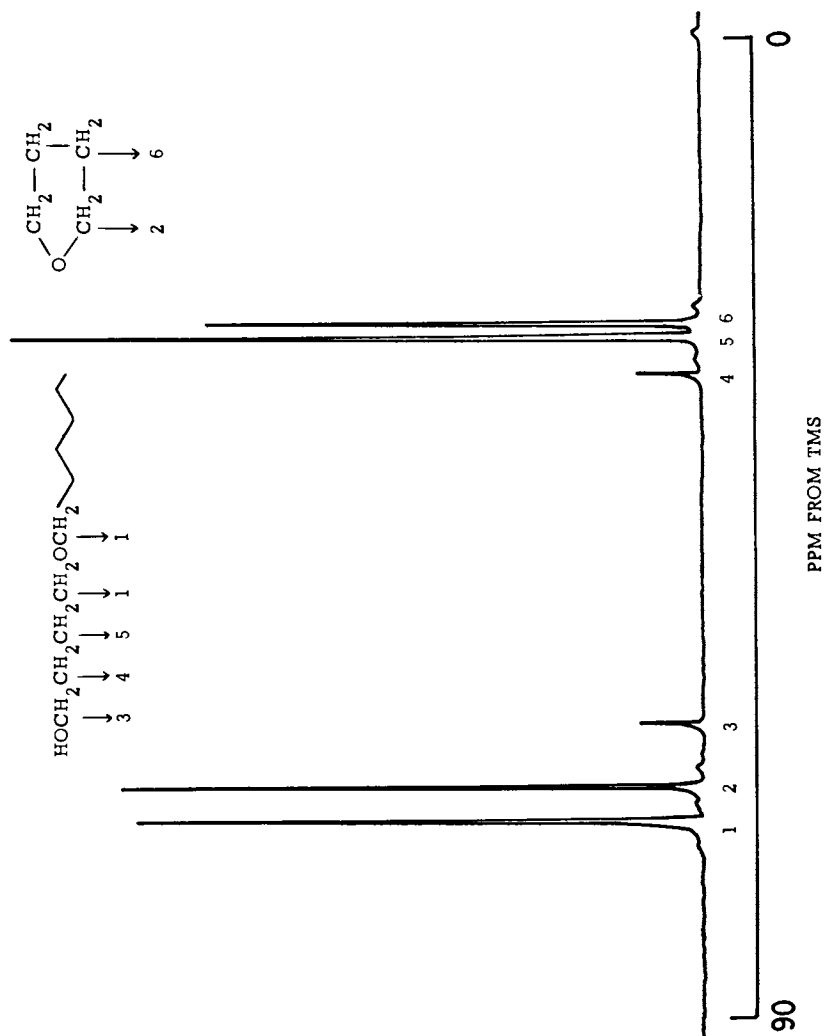


Figure 1. ^{13}C NMR spectrum of PTMEG in THF (1:1)

Subsequently, Penczek further expanded this concept and concluded that the extent of macroion or macroester formation was dependent on the polarity of the polymerization medium (15). Initial efforts to substantiate this theory using proton NMR did not lead to unambiguous spectral assignments (11). By recognizing the large chemical shift difference between fluorine in an anion and in a neutral species, Saegusa's group and we independently obtained ^{19}F -NMR data to support this mechanism (13,16). However, the ^{19}F technique is limited to examination of fluorine-containing initiators, and we decided to use ^{13}C -NMR to shed further light into the nature of this problem.

Figure 2 shows the complete ^{13}C spectra of two THF/methyltriflate polymerization mixtures, one in a nonpolar solvent (CCl_4 , Figure 2,B) and the other one in a strongly polar solvent (CH_3NO_2 , Figure 2,A). The chemical shifts of the α - and β -methylene carbon peaks of the polymer and those of the monomer closely correspond to those shown in the spectrum of polytetrahydrofuran (Figure 1). It is noteworthy that at the low field side of this polymer peak two strong signals appear at 87.5 ppm in CH_3NO_2 solution, while a single resonance peak is observed at 79.1 ppm in CCl_4 solution. We assigned the two low-field signals in CH_3NO_2 to the endo- and exo-cyclic α -methylene carbons at the oxonium ion, respectively, and the single low-field signal in CCl_4 to the α -methylene carbon of the corresponding macrofluorosulfate (5). (Some of the spectral assignments were confirmed by off-resonance decoupling.) The ^{13}C -NMR spectra therefore supported the macroion-macroester equilibration proposed by Smith and Hubin (14), and Penczek (15).

Further detail can be seen in Figure 3, which is a horizontal expansion of the oxonium region from about 85 ppm to 95 ppm, illustrating the spectra of reaction mixtures of methyltriflate with 5-, 6-, and 7-membered ring compounds. The α -methylene-carbon peaks of the methyl oxonium ions of the 5- and 7-membered cyclic ethers are found at the low field side of the spectra. A consistent ring-size effect is evident resulting in a downfield shift of about 1.7 ppm per ring expansion by one CH_2 unit. In compounds which undergo ring-opening polymerization the chemical shift of the open chain or "exo-cyclic" methylene carbons of the polymeric oxonium ions is different from the chemical shift of the ring, or "endo-cyclic" methylene carbons. Tetrahydropyran (THP), the strainless 6-membered ring, forms a tertiary oxonium ion, but does not subsequently ring open.

Based on ^1H -, ^{19}F -, and ^{13}C -NMR results we have schematically represented the equilibrium polymerization of THF with esters of trifluoromethane sulfonic acid as shown in Scheme I. Initiation occurs when the alkyl (R) group of the ester is transferred to THF to form an oxonium ion. In polar media, the oxygen atom of another THF molecule will add to the α -methylene position of the oxonium ion leading to ring-opening propagation. Because the charged species are stabilized in polar medium, the

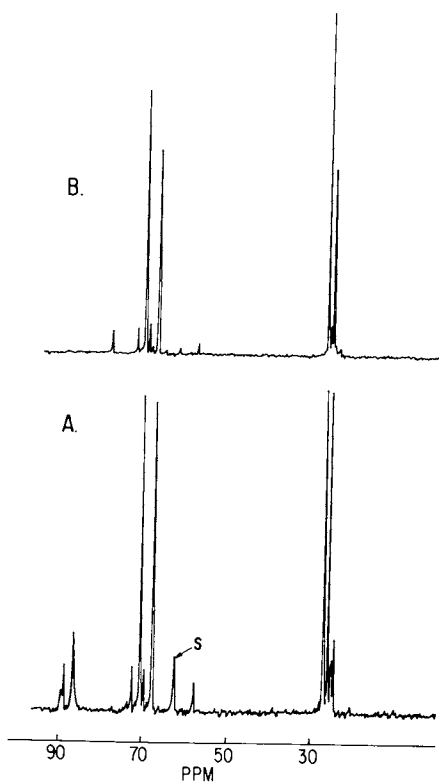


Figure 2. $C-13$ NMR spectra (22.63 MHz) of the polymerization mixture of THF- CH_3OSO_2F (6:1): (A) 64% in CH_3NO_2 , after a polymerization time of 20 min. (S indicates the solvent peak); (B) 64% in CCl_4 , after a polymerization time of 20 min.

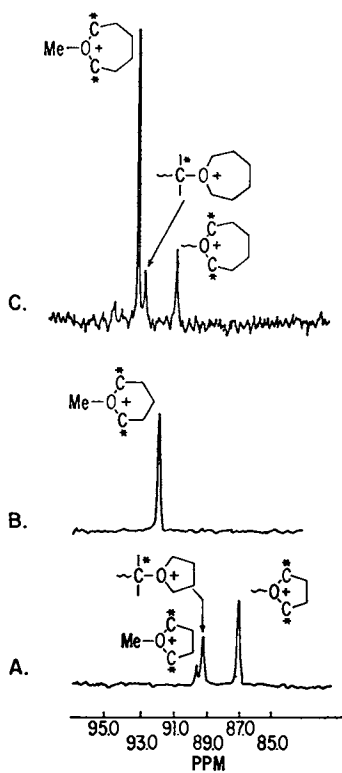
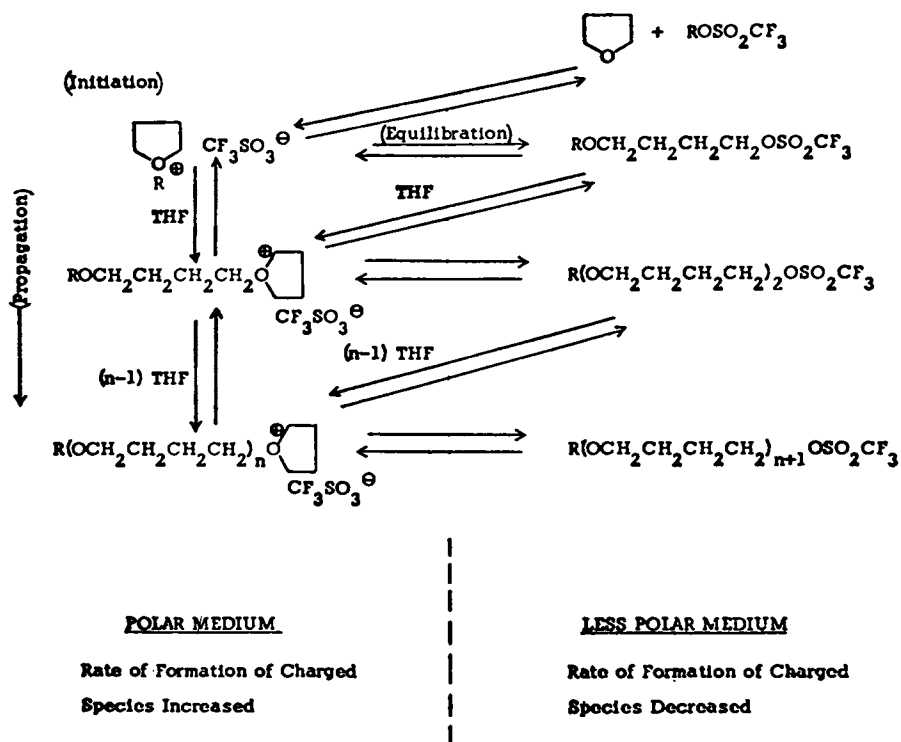


Figure 3. $C-13$ NMR spectra (22.63 MHz) of the oxonium ion region: (A) THF- $CH_3OSO_2CF_3$ (6:1) in CH_3NO_2 (64%), after 15 min; (B) THP- $CH_3OSO_2CF_3$ (6:2) in CH_3NO_2 (67%), after 60 min; (C) OXP- $CH_3OSO_2CF_3$ (2:6) in CH_3NO_2 (67%), after 30 min.



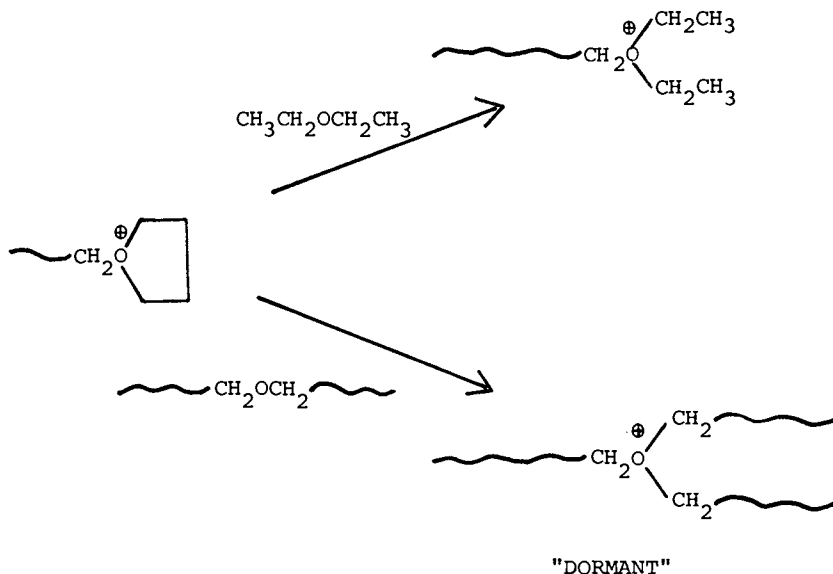
Reaction Scheme I. Polymerization of THF with esters of fluorosulfonic acids

equilibrium is shifted to favor the macroion. Therefore, the polymerization proceeds largely by following the steps from top to bottom shown on the left side of the outline.

In nonpolar media, on the other hand, the newly formed oxonium ion will either quickly convert to the corresponding soluble ester, or it will precipitate, since monomeric or short-chain oligomeric oxonium salts have low solubility in such media. The soluble ester is structurally similar to the initiator and may add another THF molecule. The resulting oxonium ion will again revert to the ester or precipitate. In fact, precipitates are generally observed during the early stages of polymerization in media of low polarity. They have been isolated and characterized as monomeric or short chain oligomeric oxonium salts (17).

As the polymer chains increase in length (at longer polymerization times or very low initiator concentrations), they will tend to stabilize the ionic ends in solution. Although the concentration of ionic species under these conditions will still be very low, both types of end-groups may participate in chain propagation (18), since the propagation rate of oxonium ions was found to be much higher than that of the corresponding macroester (19).

Chain Transfer. Dreyfuss and Dreyfuss discussed the possibility of chain transfer during cationic polymerization of cyclic ethers (20). This can occur when the cyclic oxonium ion is attacked by an oxygen of a polymer molecule rather than by monomer. The oxonium ion formed in this case is a branched site, an open-chain tertiary oxonium ion, which has been called a "dormant" ion because of its lack of ring strain (21).

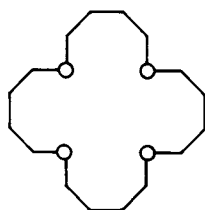


Dreyfuss and Dreyfuss reasoned that a similar chain transfer should also occur with added small acyclic ethers. Indeed in the presence of diethyl ether they found that the ultimate conversion of THF to polymer was not affected but that the intrinsic viscosity of the polymer decreased with time (20).

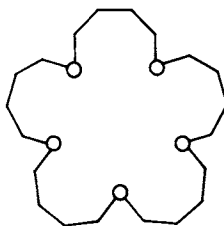
Investigation of this chain transfer reaction is greatly facilitated by using ^{13}C -NMR. In Figure 4 the low field region of a polymerization mixture of THF/ MeOSO_2F /diethyl ether is shown. We observe the α -methylene carbons of the methyl tetrahydrofuranium ion, the α -carbons of the two types of propagating chain heads, the macroion and the macroester (17). The observation of the α -methylene carbon resonances of the acyclic tertiary oxonium ion provides a direct proof of chain transfer reaction in THF polymerization.

Formation of Cyclic Oligomers. Chain transfer reactions occur by intermolecular attack of oxygen from another polyether chain on the α -methylene carbons of the oxonium ion. In an intramolecular attack a distant oxygen of the growing polymer chain itself attacks the α -methylene position of its oxonium center. In this case a macrocyclic oxonium ion is formed. Subsequent exocyclic attack by a monomer molecule will yield a macrocyclic compound containing more than one monomer units (Scheme II).

We first confirmed the formation of these macrocycles in the polymerization of THF by using coupled gas chromatography/mass spectrometry (22). Macrocyclic ethers containing up to 8 THF units could be separated and identified by this method (23). The two predominant macrocyclic species found in THF polymerization mixtures are a cyclic tetramer and a cyclic pentamer. In analogy to the "crown ether" nomenclature, we proposed the name 20-crown-4 for the cyclic tetramer and 25-crown-5 for the cyclic pentamer (22).



20-CROWN-4



25-CROWN-5

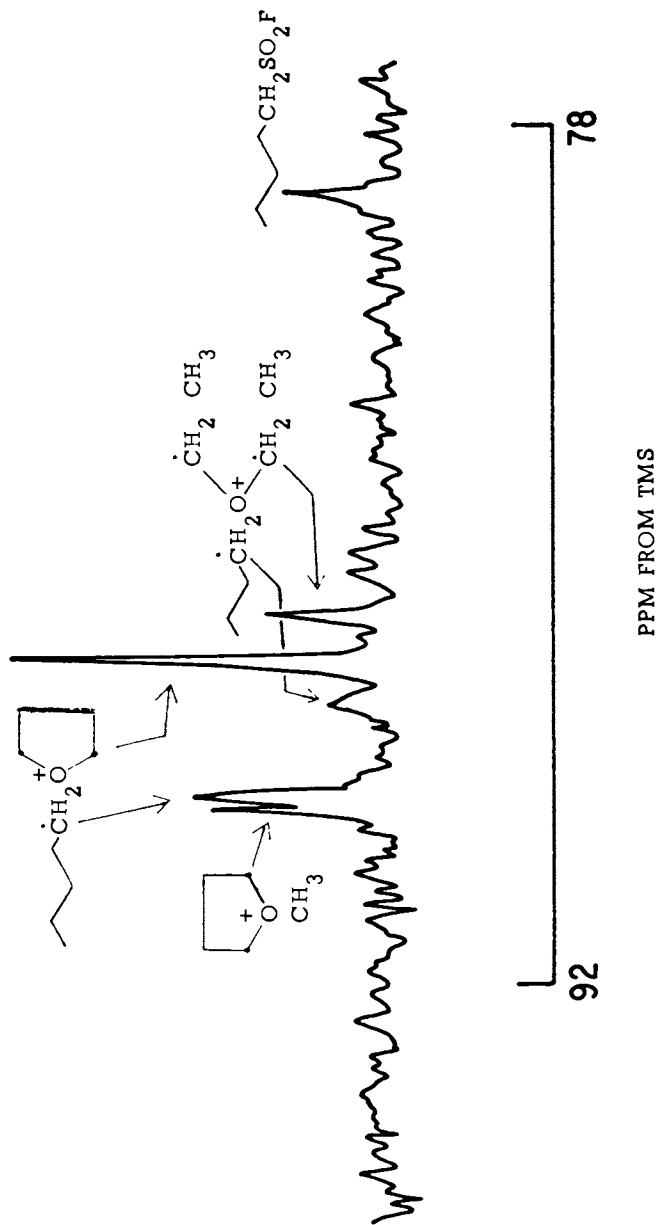
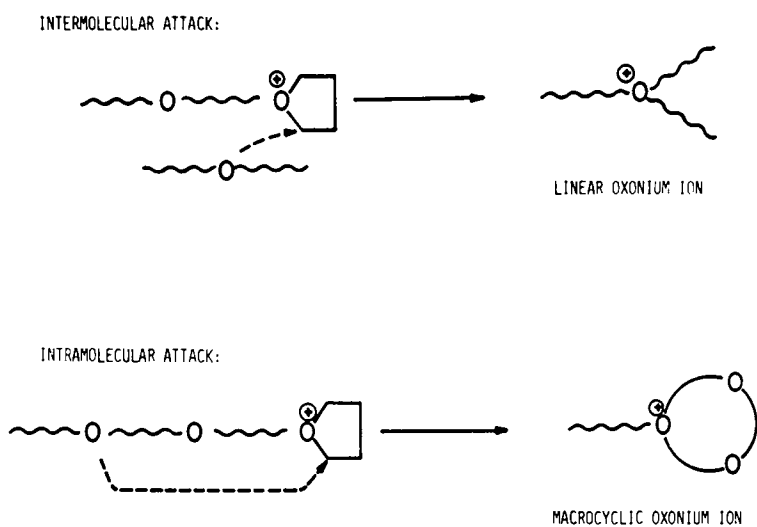


Figure 4. Partial ^{13}C NMR spectrum of a TFE- $\text{CH}_3\text{OSO}_2\text{F}$ (2.5:1) polymerization mixture in CH_3NO_2 (45.5%) after addition of diethyl ether (2.5)

**American Chemical
Society Library**

1155 16th St. N. W.

Washington, D. C. 20036



Reaction Scheme II.

Identification of these macrocycles was also facilitated by examining their ^{13}C -NMR spectra. Figure 5 shows a spectrum of the GC fraction of 20-crown-4. Due to the symmetry of this molecule there are only two distinguishable carbons: They are those α and β to the oxygen atoms at 70.4 ppm and 26.5 ppm, respectively. (The triplet at 77 ppm is due to the solvent, CDCl_3 .)

Comparison of the chemical shift data (Table 1) reveals that the peak positions of α and β carbons of 20-crown-4 are quite different from the corresponding carbons of THF or the polymeric PTME. Small but distinct chemical shift differences were also found for macrocyclic oligomers of other ring sizes.

II. Quantitative Applications of ^{13}C -NMR

In view of the excellent resolution of ^{13}C spectra it would be of interest to use these data for quantitative correlations. However, quantitative analysis by proton noise-decoupled Fourier transform ^{13}C -NMR is complicated by the fact that different carbon nuclei may have different spin relaxation times and nuclear Overhauser enhancement (NOE) factors. Therefore, the observed peak areas in the spectra are not necessarily proportional to the number of carbon atoms involved.

Schaefer and Natusch have shown that for many synthetic high polymers in solution the NOE factors and relaxation times of carbon atoms in or near the main chains are similar (24). In such cases the relative peak areas in the spectra obtained by the noise-decoupled and fast pulsing technique can be used as a good approximation for quantitative microstructure analysis. However for our investigation of the polymerization of cyclic ethers we are frequently interested in the quantitative measurements of monomers and oligomers as well as the concentrations of the continuously growing polymeric species. Therefore, the assumption of Schaefer and Natusch is not applicable.

The standard method of obtaining quantitative spectra involves the use of gated decoupling and long pulse delay, both of which require very long data collection times. Figure 6 depicts the partial ^{13}C spectrum of the α -carbon region of an equilibrated polymerization mixture of THF/ Me_3OBF_4 in CD_3NO_2 . Gated decoupling and a long pulse delay time of 10 seconds were employed to obtain the spectrum. From the monomer and polymer peak areas, the extent of polymerization at equilibrium can be determined. Measurements of chain end and the polymer peaks provide information on number-average degree of polymerization. The data collection time required to obtain this spectrum was almost three hours.

Since we were also interested in obtaining quantitative kinetic data for which the long data collection time technique cannot be used, we devised a second approach using "relative" peak intensities in the spectra obtained by fast pulsing. The two approaches are summarized as follows:

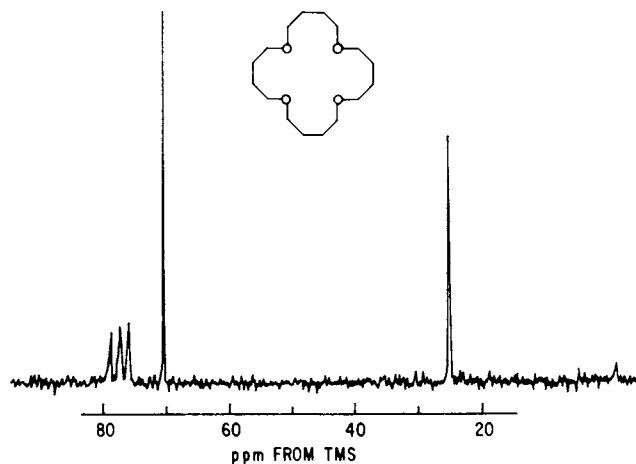


Figure 5. ^{13}C -NMR spectrum of the cyclic THF tetramer 20-crown-4 in CDCl_3 .

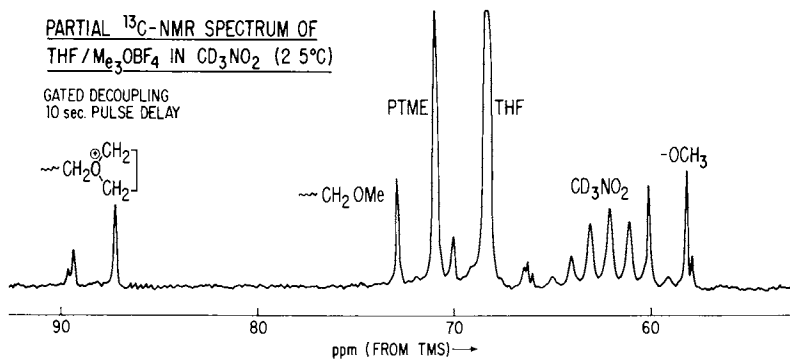


Figure 6. Polymerization mixture $\text{THF-CD}_3\text{NO}_2\text{-Me}_3\text{OBF}_4$ (mol ratios = 1:0.75:0.08) after equilibration

Table I. C-13 NMR Chemical Shift of Tetramethylene Ethers


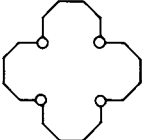
	<u>α-CARBONS</u>	<u>β-CARBONS</u>
 THF	68.2	26.2
 20-CROWN-4	70.4	26.5
$\sim\text{CH}_2\text{OCH}_2(\text{CH}_2)_2\text{CH}_2\text{OCH}_2\sim$	71.1	27.4

Table II. Quantitative C-13 NMR Spectroscopy

- A. "Absolute" Signal Intensities (Long Data Collection Times)
 - Gated Decoupling to Suppress NOE
 - Pulse Delay (>10 seconds)

- B. "Relative" Signal Intensities (Short Data Collection Times)
 - Internal Standard
 - Assumptions:
 - No Change in Relaxation Time
 - No Change in NOE
 - Viscosity Effects Negligible

In the first and obvious approach, "absolute" signal intensities are measured. Since very long data collection times are required, this method is only useful in studying equilibrated, i.e. nonchanging, systems.

In the second approach, "relative" signal intensities are compared, and data collection times of the order of 5 to 8 minutes per scan were found to be sufficient. In this approach, an internal standard peak, such as a solvent peak (e.g. CH_3NO_2), is used as the reference and compared with the peak intensity of a carbon of interest, e.g. of monomer. The underlying assumptions are that the relaxation time and NOE ratios of the internal standard and the carbon of interest remain unchanged during the course of polymerization, and that viscosity effects are negligible. Since we are dealing with relatively low conversions and low molecular weight polymers in solution, this assumption is not unreasonable.

In order to verify the validity of these assumptions we prepared several calibration samples containing different ratios of THF to CH_3NO_2 . Different amounts of polymer were added to these samples to simulate the viscous properties of the polymerization mixture. We found that the peak intensity ratio of THF to CH_3NO_2 obtained by the fast pulsing technique can indeed be linearly correlated with the corresponding weight ratios of these two compounds. Moreover, change of temperature from 0 to 35° introduced no appreciable deviations. The calibration curve is shown in Figure 7. The composition range of interest in our polymerization study is indicated by the bracket.

Thermodynamic Data. For an equilibrium polymerization, the equilibrium constant K_e is equal to the ratio of rate of

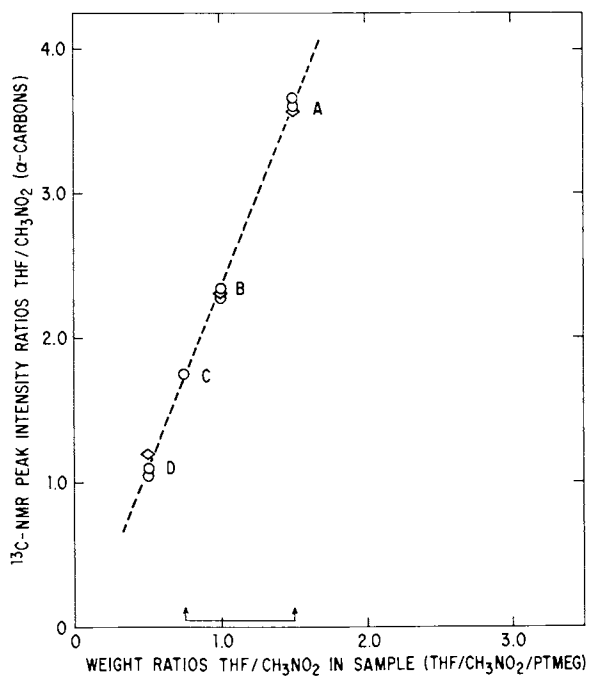
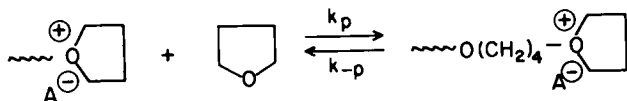


Figure 7. C -13 NMR calibration curve: (A), no PTME; (B), 20% PTME; (C), 30% PTME; (D), 40% PTME; (\diamond), 0°C; (\circ), 35°C.

propagation, k_p , to that of depropagation, k_{-p} . This constant is also related to monomer and polymer concentrations by the law of mass action. To a good approximation K_e is equal to $1/[M]_e$, where $[M]_e$ is the monomer concentration at equilibrium. From the free energy relationship one may rearrange the terms and make appropriate substitution to obtain the expression as shown (Eq. 1). Measurements of $[M]_e$ at different polymerization temperatures should yield the enthalpy and entropy of polymerization.



$$K_e = \frac{k_p}{k_{-p}} = \frac{1}{[M]_e}$$

$$\begin{aligned} \Delta F_p &= -RT \ln K_e \\ &= \Delta H_p - T\Delta S_p \end{aligned}$$

$$\boxed{\ln [M]_e = \frac{1}{T} \frac{\Delta H_p}{R} - \frac{\Delta S_p}{R}} \quad (1)$$

In Figure 8 the log of $[M]_e$ is plotted against the reciprocal of polymerization temperature. Three types of NMR data are shown. The filled circles are from the fast pulsing technique, the open circle from the gated and delayed pulsing technique and the open squares from proton NMR. From the slope and intercept of the least-square fitted line the enthalpy and entropy of polymerization were obtained, respectively.

The thermodynamic constants of THF polymerization have been investigated by a number of authors. A variety of experimental techniques have been utilized including determinations of conversion to polymer, combustion, heat capacities and vapor pressure. Comparison of our results with some previously published data shows that our results are within the range of the values reported (Table 3).

Kinetic Data. Let us now consider the kinetics of a reversible cyclic ether polymerization. For such a polymerization in progress, the kinetic expression is

$$-\frac{d[M]}{dt} = k_p [P^*][M] - k_{-p} [P^*] \quad (2)$$

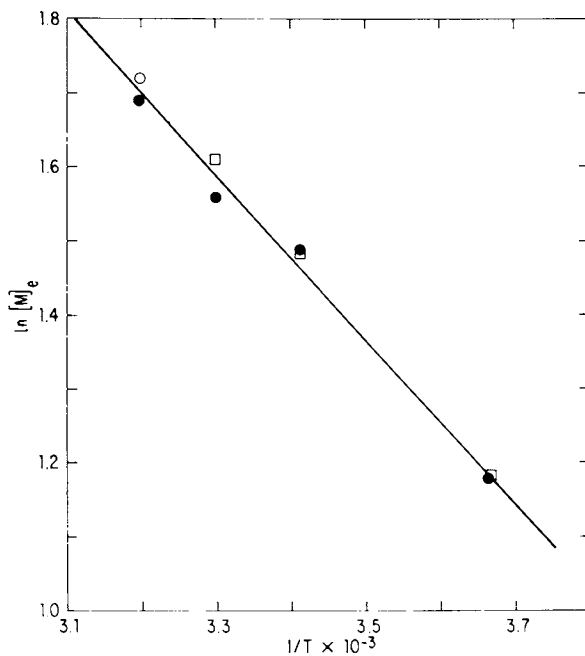


Figure 8. Determination of thermodynamic constants of THF polymerization (plot of Equation 1): (●), C-13 NMR (decoupled and fast pulsing); (○), C-13 NMR (gated and delayed pulsing); (□), ^1H NMR. $\Delta H_p = -2.2 \text{ kcal mol}^{-1}$; $\Delta S_p = -10.4 \text{ cal deg}^{-1} \text{ mol}^{-1}$.

where $[M]$ and $[P^*]$ are the molar concentrations of the monomer and growing polymer respectively, and k_p and k_{-p} are the rate constants defined earlier.

At equilibrium $d[M]/dt = 0$ and

$$k_p [M]_e = k_{-p} \quad (3)$$

By proper substitution, Eq. (2) is simplified to

$$-\frac{d[M]}{dt} = k_p [P^*] \{ [M] - [M]_e \} \quad (4)$$

Integration of Eq. (4) leads to

$$\ln \frac{[M]_{t_1} - [M]_e}{[M]_{t_2} - [M]_e} = k_p \int_{t_1}^{t_2} [P^*] dt \quad (5)$$

If the instantaneous monomer concentrations $[M]_{t_1}$ and $[M]_{t_2}$ can be continuously monitored during polymerization, and $[P^*]$ is also known, k_p can then be calculated with Eq. (5). This approach was used by Saegusa and others to study the polymerization of THF (25,26).

However, this relationship is not applicable when data from ^{13}C spectra are used. ^{13}C spectra are obtained via Fourier transform computation of data accumulated over a definite time interval, and instantaneous concentration measurements are not possible.

We have therefore modified the kinetic expression to handle the ^{13}C -NMR data. For the special case where the concentration of active chain ends $[P^*]$ is constant, the derived kinetic expression is reduced to a very simple form:

$$\ln([M]_t - [M]_e) \cong -k_p [P^*] t + \text{CONSTANT} \quad (6)$$

$[M]_t$ represents the monomer concentration at time t , as obtained from Fourier transform ^{13}C -NMR data. The rate constant of propagation k_p can now be determined by measuring $[M]_t$ as a function of polymerization time, t . (For the derivation of this expression, see the Appendix.)

In a kinetic study, we carried out a polymerization reaction of THF in CH_3NO_2 with $(CH_3)_3OBF_4^+$ at $40^\circ C$. to equilibrium and then quickly chilled the reaction mixture to $0^\circ C$. to follow further polymerization at this temperature. The kinetic data obtained are shown in Table 4. A ^{13}C scan was obtained once every 8 minutes which was found to be the optimal spectral accumulation time.

The data of Table 4 were now plotted in terms of $\ln([M]_t - [M]_e)$ versus polymerization time t , counting from the reference

Table III. Summary of Published Data of Enthalpy and Entropy of Polymerization of THF

$-\Delta H_p$ KCal / mol	$-\Delta S_p$ Cal / deg / mol	METHOD	REFERENCE
2.2	10.4	EQUILIBRIUM (^{13}C -NMR)	THIS WORK
4.6	17.7	EQUILIBRIUM (CONV. to POL.)	IVIN et al (1958)
9.1	—	COMBUSTION	CASS (1958)
4.3	17.0	EQUILIBRIUM (CONV. to POL.)	SIMS (1964)
3.3	10.7	EQUILIBRIUM WITH SOLVENT CORRECTION	IVIN et al (1965)
—	14.8	HEAT CAPACITIES	GLEGG et al (1968)
1.8	3.9	EQUILIBRIUM VAPOR PRESSURE	BUSFIELD et al (1972)

Table IV. Polymerization of Tetrahydrofuran¹ in Nitromethane at 0°C

t (min)	$[\bar{M}]_t^2$	$\ln([\bar{M}]_t - [M]_e)$
4	2.57	0.25
12	2.09	-0.21
20	1.99	-0.34
28	1.74	-0.77
42	1.50	-1.51
58	1.36	-2.53
EQUILIBRIUM	1.28	—

¹ THF: 7.38 mol l⁻¹, (CH₃)₃OBf₄: 0.59 mol l⁻¹² INTEGRATED MONOMER CONCENTRATION IN RELATIVE UNITS

time t_0 (Figure 9). These data were then least-square fitted to a straight line. The slope of this line is equal to the negative value of product $\frac{[P^*]}{k_p}$ and k_p . The propagation constant k_p of THF polymerization in CH_3NO_2 at 0°C . was found to be $1.5 \times 10^{-3} \text{ l. mol}^{-1} \text{ sec}^{-1}$. This is in good agreement with the propagation constant of a similar polymerization mixture at this temperature calculated from ^{19}F -NMR data (18).

III. Application to Copolymerizations

^{13}C -NMR kinetic analysis would appear to be most useful for studying polymerization systems which cannot be adequately characterized by proton or fluorine NMR methods. Examples of such systems are e.g. copolymerizations of cyclic ethers, and in the last part of this review we would like to discuss briefly some preliminary results on THF copolymerizations.

Figure 10 presents a summary of the α carbon chemical shifts of oxonium ions and esters of some of the compounds discussed earlier. The carbon atoms α to an oxonium center cover a range of about 25 ppm. The peaks due to all the different oxonium ions and esters can be clearly distinguished, and ^{13}C -NMR therefore appeared to be an excellent technique for studying such copolymerizations.

As an example of a cyclic ether copolymerization, we will briefly discuss the polymerization of THF with OXP initiated with methyltriflate. The homopolymerizations of both cyclic monomers follow a similar mechanism, and both were found to proceed via macrooxonium ion and/or the macroester mechanism depending on the polarity of the polymerization medium. There should then be 8 possible end-groups, i.e. two types of methoxy tails having a penultimate THF or OXP unit, respectively, two covalent macroesters, and four different oxonium ion propagating chain heads: two from a THF oxonium center attached to penultimate THF or OXP units, and two from an OXP oxonium center attached to THF and OXP penultimate units (Scheme III).

Figure 11 shows the α carbon resonance region of such a THF/OXP copolymerization in CH_3NO_2 . At about 55 ppm we observe the peak due to the methoxy methyl carbons of the chain ends, and further downfield a solvent peak and then the methylene carbons of the unreacted monomers, THF and OXP. There are two peaks attributable to the polymeric methylene carbons. The higher field one is due to THF and the other one to OXP. Similarly, two peaks are observed for the methylene carbons attached to the methoxy chain ends. The fact that the intensities of these two peaks are similar indicates that both THF and OXP participate in the initiation step.

In the macroester region, we have a small signal due to OXP-ester only, no THF macroester was observed under these conditions. On the other hand, in the oxonium region there are resonances due to the THF-macroions only but not to OXP-macroions. Therefore, in this particular polymerization system we have identified 4 out

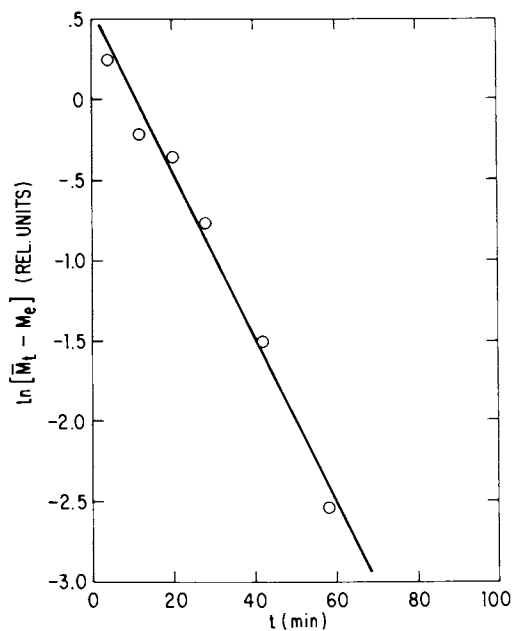


Figure 9. Determination of kinetic constants (plot of Equation 6). Polymerization of THF in CH_3NO_2 at 0°C : $k_p = 1.5 \times 10^{-3} \text{ L mol}^{-1} \text{ sec}^{-1}$.

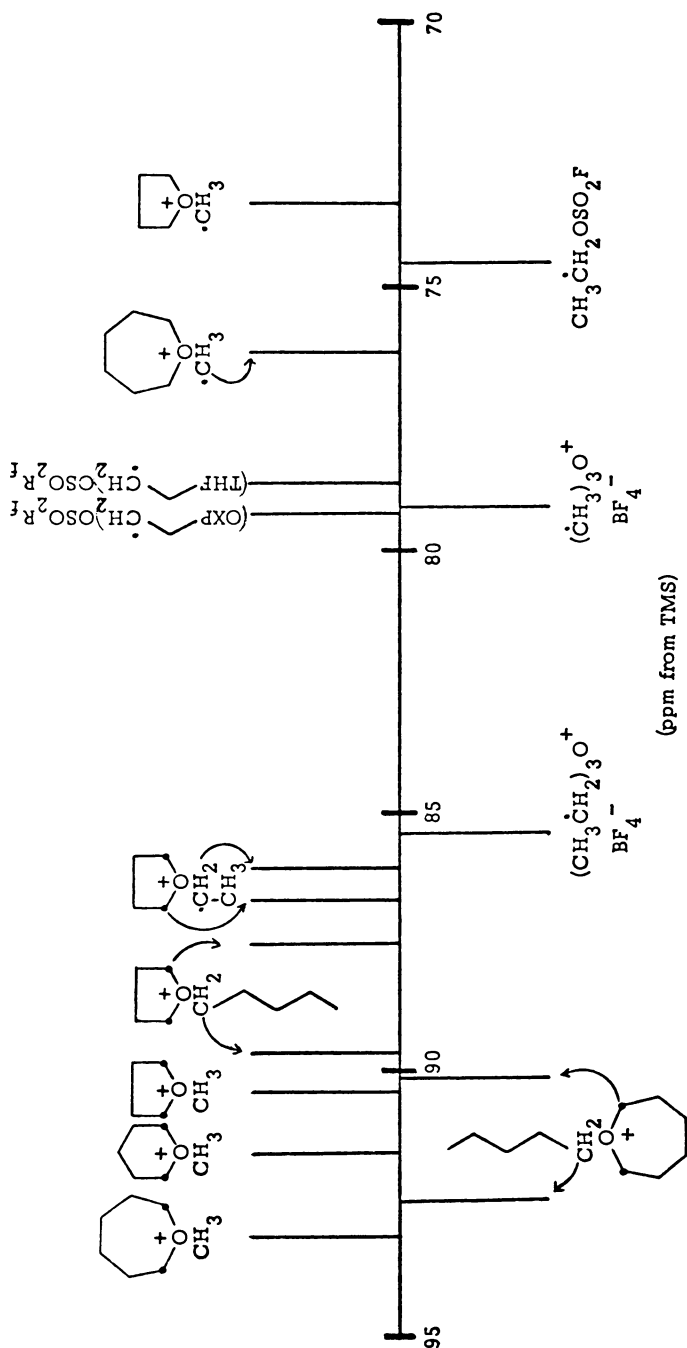
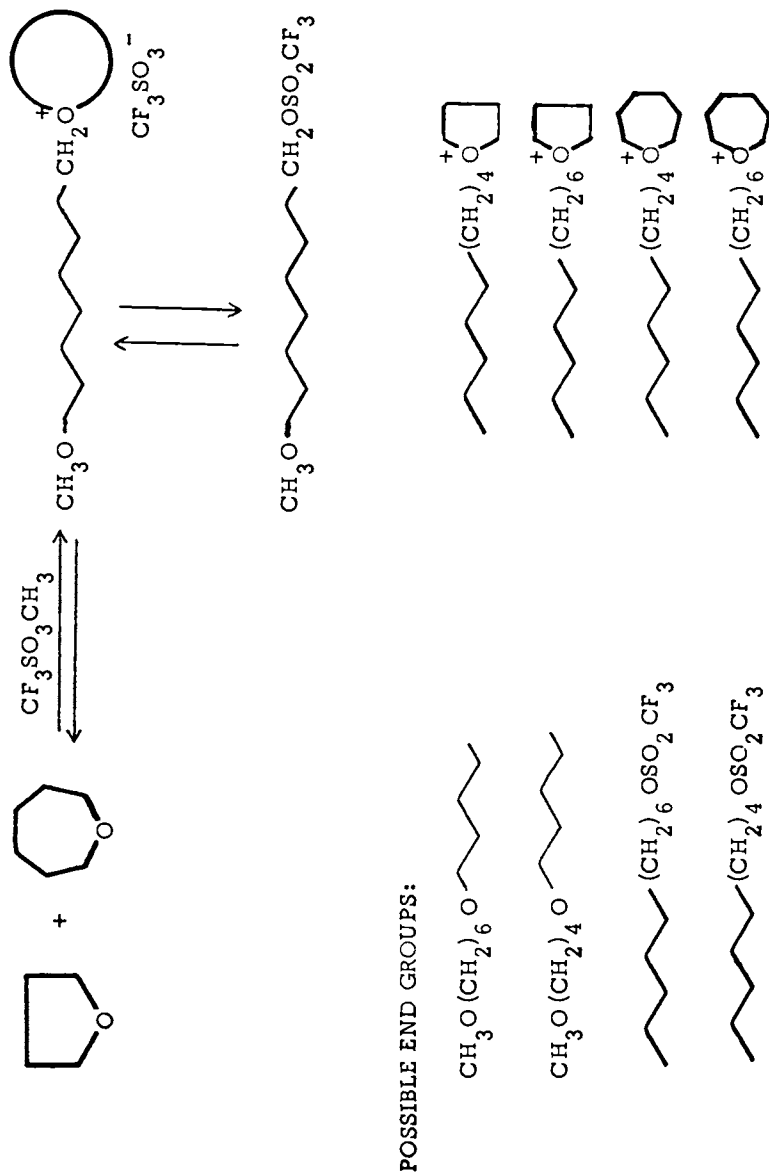


Figure 10. $C-13$ NMR chemical shift assignments of some oxonium ions and fluorosulfonate esters



Reaction Scheme III. Copolymerization

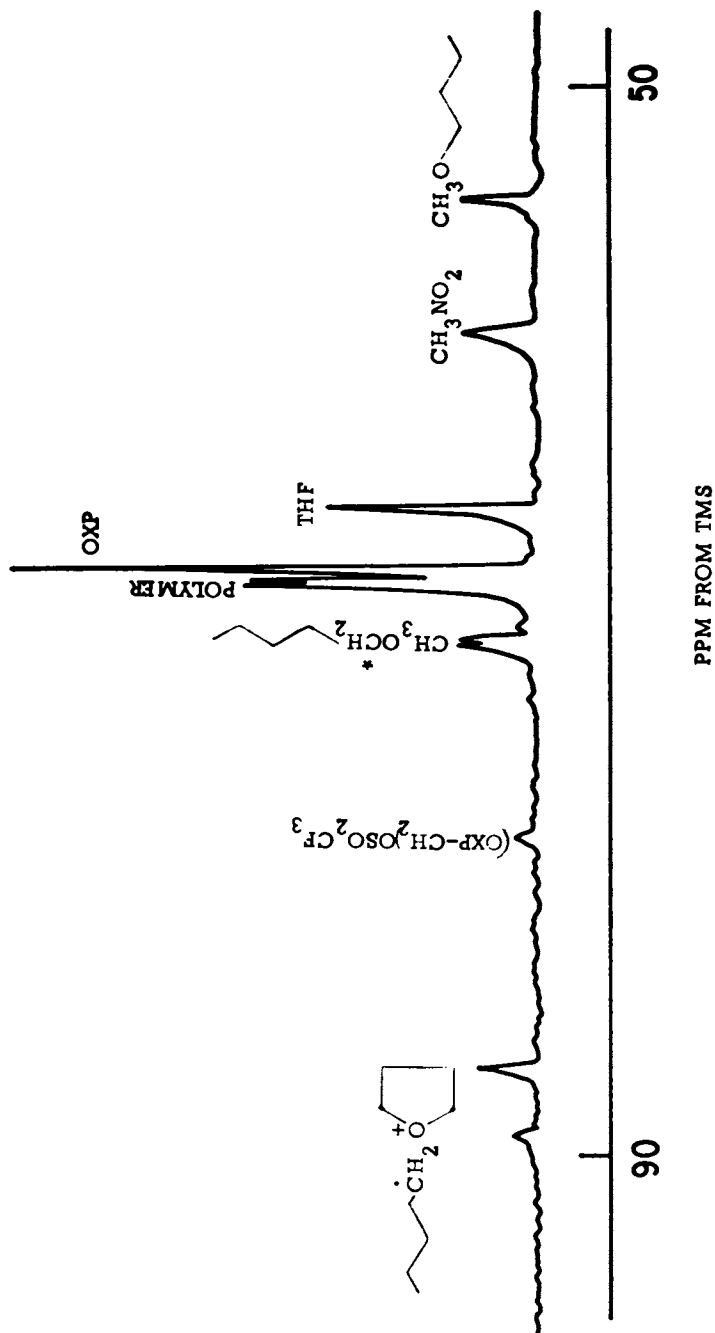


Figure 11. Partial ^{13}C NMR spectrum of a THF-OXP- $\text{CF}_3\text{SO}_3\text{CH}_3$ (1.4:2.8:1) polymerization mixture in CH_3NO_2 (28%) after 30 min at 25°C .

of the 8 possible chain ends. The results of a quantitative ^{13}C analysis of a similar THF/XP copolymerization system are summarized in Table 5 in terms of total % conversion to polymer, kinetic degree of copolymerization, copolymer composition and feed composition. The kinetic degree of copolymerization of 6.2 is, within experimental error, the same as the theoretically calculated value. Therefore, our results indicate that each initiator molecule initiates one copolymer chain as in homopolymerizations. Furthermore the copolymer composition was found to be very similar to feed composition. This suggests that cationic copolymerization of the two cyclic ethers may be statistically random.

Table V. Copolymerization of THF-XP at 20°C

Moles of THF/XP/ CD_3NO_2 / $\text{CH}_3\text{SO}_3\text{CF}_3$ = 1.04/.75/1.56/.14

Total Conversion, %	44
Kinetic Degree of Copolymerization	6.2
Copolymer Composition	
Mol % THF	57
Mol % XP	43
Feed Composition	
Mol % THF	58
Mol % XP	42

We are currently extending this approach to investigate the copolymerization of THF with other cyclic ethers.

CONCLUSION

NMR methods are ideally suited to study polymerization reactions of cyclic ethers in situ, without disturbing the polymerization equilibria. ^{13}C -NMR methods generally afford much more detailed information than either ^1H - or ^{19}F -NMR methods, but quantitative evaluation of data is not as straightforward, due to the necessity for Fourier transform and noise decoupling techniques. A ^{13}C -NMR method based on relative signal intensities has now been developed for obtaining quantitative information of homopolymerization and copolymerization systems, which may otherwise not be easily accessible.

ACKNOWLEDGMENT

We would like to thank Dr. W. W. Yau for his assistance in modifying the kinetic theory, Dr. J. J. Chang for technical assistance, and Dr. G. E. Heinsohn for a critical review of the manuscript.

APPENDIX

Kinetic Analysis of Cyclic Ether Polymerization
by Fourier Transform NMR

The kinetic expression generally applicable to reversible equilibrium polymerizations is:

$$\ln \frac{[M]_{t_1} - [M]_e}{[M]_{t_2} - [M]_e} = k_p \int_{t_1}^{t_2} [P^*] dt \quad (1)$$

$[M]_t$ is the monomer concentration at time t , $[M]_e$ is the equilibrium monomer concentration, and $[P^*]$ is the concentration of active polymer chain sites. Application of this formula requires instantaneous concentration measurements at times t_1 and t_2 . This relationship is not applicable when the data are accumulated over a definite time interval, such as by multiple pulsing in ^{13}C -NMR. In order to handle this type of data, we have modified this expression, introducing an arbitrary reference time, t_0 . $[M]_t$ and $[M]_0$ are the monomer concentrations at times t and t_0 .

$$-\ln \frac{[M] - [M]_e}{[M]_0 - [M]_e} = k_p \int_{\tau=t_0}^t [P^*] d\tau = k_p P(t) \quad (2)$$

$P(t)$ is used to designate the integral of $[P^*]$. Eq. (2) is then converted into an exponential form and integrated through Δt , the time interval required for spectral accumulation.

$$\int_{\tau=t-\frac{1}{2}\Delta t}^{t+\frac{1}{2}\Delta t} [M] d\tau - [M]_e \Delta t = ([M]_0 - [M]_e) \int_{\tau=t-\frac{1}{2}\Delta t}^{t+\frac{1}{2}\Delta t} e^{-k_p P(\tau)} d\tau \quad (3)$$

Rearrangement of terms of Eq. (3) leads to

$$\frac{[M]_t - [M]_e}{[M]_0 - [M]_e} = \frac{1}{\Delta t} \int_{\tau=t-\frac{1}{2}\Delta t}^{t+\frac{1}{2}\Delta t} e^{-k_p P(\tau)} d\tau \quad (4)$$

The $[M]_t$ represents the integral and is the monomer concentration measured from the ^{13}C spectrum at time t .

Since the integral of the right-hand term of Eq. (4) cannot be analytically integrated, we took the following procedure:

Let the integrand of that integral be X and the integrated X be Y . If each Y is expanded in Taylor's series, we obtain two series of terms in Eq. (5).

$$\begin{aligned}
 \int_{\tau = t - \frac{1}{2}\Delta t}^{t + \frac{1}{2}\Delta t} X d\tau &= Y\left(t + \frac{1}{2}\Delta t\right) - Y\left(t - \frac{1}{2}\Delta t\right) \\
 &= Y(t) + \left(\frac{1}{2}\Delta t\right)Y' + \frac{\left(\frac{1}{2}\Delta t\right)^2}{2!}Y'' + \frac{\left(\frac{1}{2}\Delta t\right)^3}{3!}Y''' + \dots \\
 &\quad - \left[Y(t) - \left(\frac{1}{2}\Delta t\right)Y' + \frac{\left(\frac{1}{2}\Delta t\right)^2}{2!}Y'' - \frac{\left(\frac{1}{2}\Delta t\right)^3}{3!}Y''' + \dots \right] \\
 &= (\Delta t)Y' + \frac{(\Delta t)^3}{24}Y''' + \frac{(\Delta t)^5}{16 \times 5!}Y^{(5)} + \dots \\
 &= (\Delta t) \left[X + \frac{(\Delta t)^2}{24}X'' + \dots \right] \quad (5)
 \end{aligned}$$

where Y' , Y'' and Y''' represent the first, second and third derivatives of Y , respectively. Since every other term of these two series cancels, we collect the remaining terms in Y 's and then in X 's. The X 's and Y 's are related by the expressions shown below:

$$X = Y' = e^{-k_p P(t)} \quad (6)$$

$$X' = Y'' = \frac{dX}{dt} = -k_p [P^*] X \quad (7)$$

$$X'' = Y''' = \left(k_p^2 [P^*]^2 - k_p \frac{d[P^*]}{dt} \right) X \quad (8)$$

In Eq. (9) we write the kinetic equation first in terms of X and then in terms of the pertinent kinetic constants.

$$\begin{aligned}
 \frac{[\bar{M}]_t - [M]_e}{[M]_0 - [M]_e} &= X + \frac{(\Delta t)^2}{24} X'' + \dots \\
 &= e^{-k_p P(t)} \left\{ 1 + \frac{(\Delta t)^2}{24} \left(k_p^2 [P^*]^2 - k_p \frac{d[P^*]}{dt} \right) \right\} + \dots \quad (9)
 \end{aligned}$$

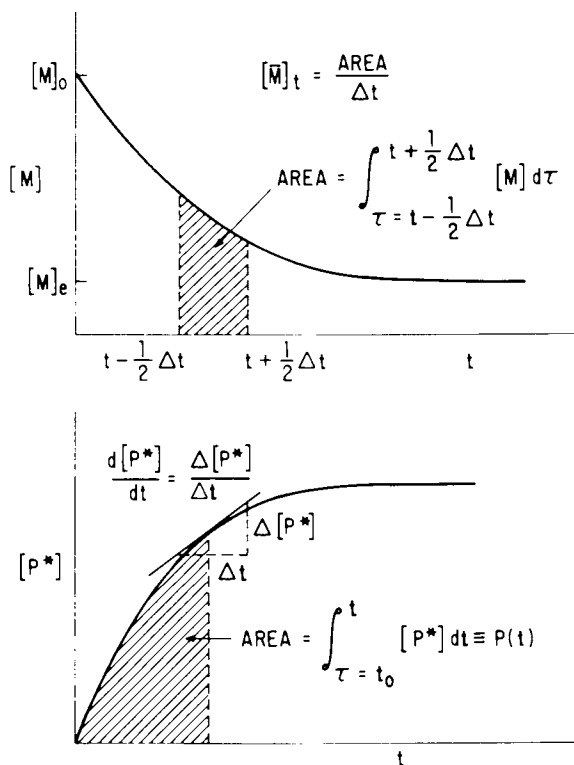


Figure 12. Model graphs for a kinetic analysis of an equilibrium polymerization (definition of variables of Equation 11)

Next, Eq. (9) is converted into the logarithmic form

$$\ln \frac{[M]_0 - [M]_e}{[\bar{M}]_t - [M]_e} = k_p P(t) - \ln \left\{ 1 - \frac{(\Delta t)^2}{24} k_p \frac{d[P^*]}{dt} + \frac{(\Delta t)^2}{24} k_p^2 [P^*]^2 + \dots \right\} \quad (10)$$

The higher order terms in Δt of Eq. (10) are very small and can be neglected, and we arrive at the final expression.

$$\ln \frac{[M]_0 - [M]_e}{[\bar{M}]_t - [M]_e} \cong k_p \left[P(t) + \frac{(\Delta t)^2}{24} \frac{d[P^*]}{dt} \right] - \frac{(\Delta t)^2}{24} k_p^2 [P^*]^2 \quad (11)$$

Examination of Eq. (11) reveals that it is very similar to the well known equation used with instantaneous data collection, but it contains two additional correction terms. These correction terms will vanish if Δt , the accumulation time, becomes infinitesimally small. Then $[\bar{M}]_t$ becomes the instantaneous monomer concentration.

Figure 12 illustrates the experimental data required for a kinetic analysis. We need $[M]_0$, the monomer concentration at a reference time t_0 , the equilibrium concentration $[M]_e$, $[\bar{M}]_t$, which is this area divided by Δt . Also needed are $P(t)$, the integrated area under the $[P^*]$ versus t curve and the slope, $d[P^*]/dt$.

For polymerization systems in which the concentration of active chain sites $[P^*]$ is constant, the kinetic expression derived in Eq. (11) can be further simplified. Since $d[P^*]/dt = 0$, the second term of Eq. (11) vanishes and the third term becomes a constant. By taking the constant $[P^*]$ outside the integral and integration, the kinetic expression is reduced to the simple form shown in Eqs. (12) and (13) and in the discussion section.

$$\begin{aligned} \ln \frac{[M]_0 - [M]_e}{[\bar{M}]_t - [M]_e} &\cong k_p \int_{\tau=t_1}^t [P^*] d\tau + \text{CONSTANT} \\ &\cong k_p [P^*] (t - t_1) + \text{CONSTANT} \end{aligned} \quad (12)$$

$$\ln ([\bar{M}]_t - [M]_e) \cong -k_p [P^*] t + \text{CONSTANT} \quad (13)$$

Eq. (13) shows that k_p can be determined by measuring $[\bar{M}]_t$ as a function of polymerization of time, t .

Literature Cited

1. Saegusa, T., Kimura, Y., Fujii, H., Kobayashi, S., *Macromolecules*, (1973), 6, 657.
2. Penczek, St., Matyjaszewski, K., *J. Polym. Sci., Symposium 56*, (1976), 255.
3. Hoene, R., Reichert, K. W., *Makromol. Chem.*, (1976), 177, 3545.
4. Pruckmayr, G., Wu, T. K., *Macromolecules*, (1978), 11, 662.
5. Pruckmayr, G., Wu, T. K., *Macromolecules*, (1975), 8, 954.
6. Meerwein, H., Delfs, D., Morschel, H., *Angew. Chem.*, (1960), 24, 927.
7. Dreyfuss, P., Dreyfuss, M. P., *Advan. Chem. Ser.*, (1969), 91, 335.
8. Dreyfuss, P., Dreyfuss, M. P., "Ring-Opening Polymerizations", K. C. Frisch, S. L. Reegen, eds., Marcel Dekker, N.Y., (1969).
9. Saegusa, T., Kobayashi, S., *Polyethers, ACS Symposium 6*, (1975), 150.
10. Vofsi, D., Tobolsky, A. V., *J. Polym. Sci.*, (1965), A3, 3261.
11. Kobayashi, S., Danda, H., Saegusa, T., *Bull. Chem. Soc. Japan*, (1973), 46, 3214.
12. Pruckmayr, G., Wu, T. K., *Macromolecules*, (1973), 6, 33.
13. Wu, T. K., Pruckmayr, G., *Macromolecules*, (1975), 8, 77.
14. Smith, S., Hubin, A. J., *J. Macromol. Sci., Chem.*, (1973), 7, 1399.
15. Matyjaszewski, K., Penczek, St., *J. Polym. Sci., Chem.*, (1974), 12, 1905.
16. Kobayashi, S., Danda, H., Saegusa, T., *Macromolecules*, (1974), 7, 415.

17. Pruckmayr, G., Wu, T. K., ACS Central Regional Meeting, Akron, Ohio, (May 1976).
18. Kobayashi, S., Morikawa, K., Saegusa, T., *Macromolecules*, (1975), 8, 386.
19. Buyle, A. M., Matyjaszewski, K., Penczek, St., *Macromolecules*, (1977), 10, 269.
20. Dreyfuss, M. P., Dreyfuss, P., *J. Polym. Sci.*, (1966), (A-1) 4, 2179.
21. Saegusa, T., Kobayashi, S., *Progress in Polymer Sci. (Japan)*, (1973), 6, 107.
22. McKenna, J. M., Wu, T. K., Pruckmayr, G., *Macromolecules*, (1977), 10, 877.
23. Pruckmayr, G., Wu, T. K., *Macromolecules*, (1978), 11, 265.
24. Schaefer, J., Natusch, D. F. S., *Macromolecules*, (1972), 5, 416.
25. Saegusa, T., Kobayashi, S., *J. Polym. Sci., Polymer Symposia*, (1976), 56, 241.
26. Dreyfuss, P., Dreyfuss, M. P., "Comprehensive Chemical Kinetics", C. H. Bamford, C. H. Tipper, eds., 259, Elsevier, 15, (1976).

Discussion

W. Pasika, Laurentian Univ., Ont.: I would like to refer back to the fact that the compositional ratio is the same as the feed ratio. You indicated it was a random process. If the mechanism is an S_N2 type mechanism then the compositional ratio will depend very much on the character of the monomer. The character of the copolymerization could be other than random. In such a system the compositional ratio will not necessarily be the same as the feed ratio.

T. K. Wu: True. This is a preliminary report, and we discussed only one data point from one particular feed ratio. The result may be fortuitous.

W. Pasika: On the other hand it may well be that these particular two monomers have in fact the same reactivity when it comes to the particular type of mechanism.

D. J. Worsfold, NRC, Ont.: I was glad to see that you were able to identify and measure the amount of dormant polymer present in the THF polymerization. The amount of the dormant chain end

would increase during the course of the polymerization as the amount of polymer produced increases and one would think this would have some effect on the first order kinetics of the reaction. Yet it has always been successful to use first order kinetics to describe the disappearance of the monomer. Is it because the amount of dormant chain end is very small or is it that there is some compensatory effect which still gives the first order kinetics in monomer disappearance?

T. K. Wu: We tried to determine the amount of dormant species in homopolymerizations. We can measure the ratio of "exocyclic" vs. "endocyclic" methylene carbon resonance, and if the ratio is not exactly 1 to 2 we can get an estimate of the concentration of dormant ion.

G. Pruckmayr, Du Pont, Delaware: The concentration of dormant ion is low. We can estimate the concentration of macrocyclic plus linear dormant ions from the intensity ratio of the different α -methylene groups. The total is quite small in homopolymerizations, particularly at short reaction times.

P. Sipos, Du Pont, Ont.: In connection with your copolymerization have you seen a difference in the composition of the polymer and the feed ratio when larger oxy rings are used?

T. K. Wu: The largest ring used so far was oxepane, but we are planning to go in the other direction, using four membered rings. However, in this case complications arise because the strained oxetane ring does not undergo equilibrium polymerization.

RECEIVED March 13, 1979.

A Comparison of Models and Model Parameters for the Interpretation of Carbon-13 Relaxation in Common Polymers

ALAN ANTHONY JONES, GARY L. ROBINSON, and FREDRIC E. GERR

Jeppson Laboratory, Department of Chemistry, Clark University, Worcester, MA 01610

Carbon-13 spin relaxation in polymers is now a common probe of chain dynamics resulting in a proliferation of polymers studied and models used to relate spin relaxation to polymer motion (1). Presently it appears useful to draw comparisons both among the dynamics of various polymers and among the interpretational models. For these comparisons, it is necessary to center attention on polymers which have been thoroughly investigated experimentally. By this we mean it is desirable to have measurements of several different relaxation parameters including the spin-lattice relaxation time T_1 , the nuclear Overhauser enhancement NOE, and to a limited extent the spin-spin relaxation time T_2 . Although carbon-13 NMR is a most useful probe of chain dynamics, it is also important to measure relaxation parameters of other nuclei in the same polymer. Both proton and fluorine nuclei are excellent candidates which add information on motions at other frequencies not available from carbon-13 relaxation alone (2). An approach somewhat similar to observing two types of nuclei is the observation of one nucleus at several magnetic field strengths since this also provides information on dynamics in other frequency domains. Lastly it is informative to vary such non-spectroscopic parameters as polymer molecular weight, concentration, temperature, and solvent. If a model can consistently interpret a large amount of spin relaxation data under a variety of experimental conditions, then it is worthwhile to consider in some detail the significance of the model parameters.

Survey of Interpretational Models

Relaxation parameters are usually written in terms of transition probabilities, W , and spectral densities, J . For carbon-13 nuclei under condition of proton decoupling (3)

$$1/T_1 = W_0 + 2W_{1I} + W_2$$

0-8412-0505-1/79/47-103-271\$05.00/0
© 1979 American Chemical Society

$$\begin{aligned}
 (\text{NOE}) - 1 &= \gamma_S(W_2 - W_0)/\gamma_I(W_0 + 2W_{1I} + W_2) \\
 &= T_1\gamma_S(W_2 - W_0)/\gamma_I \\
 W_0 &= \gamma_I^2\gamma_S^2\hbar^2J_0(\omega_0)/20r^6 \\
 W_{1I} &= 3\gamma_I^2\gamma_S^2\hbar^2J_1(\omega_I)/40r^6 \\
 W_2 &= 3\gamma_I^2\gamma_S^2\hbar^2J_2(\omega_2)/10r^6 \\
 \omega_0 &= \omega_S - \omega_I; \omega_2 = \omega_S + \omega_I.
 \end{aligned}
 \tag{1}$$

The carbon nuclei are to be identified with I, the proton nuclei with S, and the carbon-proton internuclear distance with r. The spectral density is the Fourier transform of a correlation function which is usually based on a probabilistic description of the motion modulating the dipole-dipole interactions. The spin-spin relaxation time, T_2 , is usually written directly as a function of spectral densities (3).

$$\begin{aligned}
 1/T_2 &= (1/40)\hbar^2\gamma_S^2\gamma_I^2(J_0(\omega_0) + \\
 &3J_1(\omega_I) + 6J_2(\omega_S + \omega_I) + \\
 &4J_0(0) + 6J_1(\omega_S)).
 \end{aligned}
 \tag{1a}$$

The simplest motional description is isotropic tumbling characterized by a single exponential correlation time (4). This model has been successfully employed to interpret carbon-13 relaxation in a few cases, notably the methylene carbons in polyisobutylene among the well studied systems (5). However, this model is unable to account for relaxation in many macromolecular systems, for instance polystyrene (6) and poly(phenylene oxide) (7, 8). In the latter case, the estimate of the correlation time varies by an order of magnitude between an interpretation based on T_1 and an interpretation based on the NOE (7).

The failure of this model led to the application of motional descriptions involving several correlation times. The simplest of these, a two correlation time model, was developed by Woessner (9) and suggested for macromolecular systems by Allerhand, Dodrell, and Glushko (3). The model considers two motions modulating the dipole-dipole interaction: anisotropic internal rotation about an axis which also undergoes overall rotatory diffusion. This model can successfully account for the carbon-13 T_1 and NOE values observed for the methyl carbons in PIB (5). The methyl group is

pictured as rotating about the three-fold symmetry axis which is rigidly attached to a backbone bond undergoing isotropic tumbling caused by backbone rearrangements. On the other hand, this model failed to account for relaxation of nuclei in the phenyl group of polystyrene (6).

It was soon realized that a distribution of exponential correlation times is required to characterize backbone motion for a successful interpretation of both carbon-13 T_1 and NOE values in many polymers (1, 10). A correlation function corresponding to a distribution of exponential correlation times can be generated in two ways. First, a convenient mathematical form can serve as the basis for generating and adjusting a distribution of correlation times. Functions used earlier for the analysis of dielectric relaxation such as the Cole-Cole (11) and Fuoss-Kirkwood (12) descriptions can be applied to the interpretation of carbon-13 relaxation. Probably the most proficient of the mathematical form models is the $\log\text{-}\chi^2$ distribution introduced by Schaefer (10). These models are able to account for carbon-13 T_1 and NOE data although some authors have questioned the physical insight provided by the fitting parameters (11, 13).

The second method used to generate correlation functions which result in a distribution of exponential correlation times is to start with a lattice model and consider rearrangements caused by a crankshaft motion, the three-bond jump. There are now at least three modifications of this model all based on the approach introduced by Valeur, Jarry, Geny and Monnerie (VJGM) (14, 15, 16). The fundamental correlation function is identical with one developed by Glarum (17), and Hunt and Powles (18) from a different physical picture. This second type of model involving a distribution of correlation times has been successful in interpreting carbon-13 relaxation data (11), and it has also been successful in interpreting proton relaxation data or proton and carbon relaxation data which were not interpreted as well by the distributions of correlation times generated from mathematical forms (2, 8). In addition to these interpretational advantages, the lattice models may provide a better basis for physical insight since they are based on a specific motion possible in linear polymers (13) and since the dependence of model parameters on concentration and temperature seems reasonable (11). Because of these potential advantages, we shall turn our attention to a more detailed consideration of the lattice models.

Lattice Models

The starting point of all three models is the equation

$$dP_a^n/dt = w(-2P_a^n + P_a^{n-2} + P_a^{n+2}) \quad [2]$$

which expresses the time dependence of the probability P of bond n in the a direction. The probability per unit time that any particular three-bond segment with the proper gauche conformation undergoes rearrangement is w . For a very long chain, a continuous solution was produced by VJGM (14, 15) which served as a basis for a correlation function and, by Fourier transformation, a spectral density. However, this result alone did not provide a consistent interpretation of spin relaxation until the correlation function was modified by the inclusion of an additional exponential decay (16, 19). The modified spectral density, most simply written as

$$J(\omega) = \frac{\tau_0 \tau_D (\tau_0 - \tau_D)}{(\tau_0 - \tau_D)^2 + \omega^2 \tau_0^2 \tau_D^2} \left\{ \left(\frac{\tau_0}{2\tau_D} \right)^{1/2} \right.$$

$$\times \left[\frac{(1 + \omega^2 \tau_0^2)^{1/2} + 1}{1 + \omega^2 \tau_0^2} \right]^{1/2} + \left(\frac{\tau_0}{2\tau_D} \right)^{1/2} \quad [3]$$

$$\times \frac{\omega \tau_0 \tau_D}{(\tau_0 - \tau_D)} \left[\frac{(1 + \omega^2 \tau_0^2)^{1/2} - 1}{1 + \omega^2 \tau_0^2} \right]^{1/2} - 1 \left. \right\}$$

has two adjustable parameters. The rate of occurrence of the three-bond jump is governed by the choice of τ_D , and the added exponential decay is governed by τ_0 . The physical significance of τ_0 is of some interest since it was not introduced from the fundamental lattice equation, Eq. 2.

In interpretational applications it is found that τ_D and τ_0 have different apparent activation energies (2, 20). The activation energy for τ_0 is lower than for τ_D and has values nearly equal to the activation energy derived from solvent viscosity (2, 20). The larger activation energy of τ_D has been associated with backbone rearrangements while that of τ_0 has been associated with long range tumbling. Going back to the lattice equations, it is not easy to support such a distinction, but on the other hand τ_0 does enter the correlation function in the same manner as an entirely independent overall rotatory diffusion correlation time.

A second solution to Eq. 2 is derived from a different physical picture. Jones and Stockmayer (13) solved the lattice equation for a finite segment length. Rearrangements caused by the three-bond jump are considered in a segment containing $2m-1$ bonds with complete neglect of directional correlations of bonds outside the segment. This yields a dynamic description for the central bond in the segment which has been carried through to produce the spectral density (13),

$$J(\omega) = 2 \sum_{k=1}^s \frac{G_k \tau_k}{1 + \omega^2 \tau_k^2} \quad [4]$$

$$\tau_k^{-1} = w \lambda_k \quad s = \frac{m+1}{2}$$

$$\lambda_k = 4 \sin^2((2k-1)\pi/2(m+1))$$

$$G_k = 1/s + (2/s) \sum_{q=1}^{s-1} \exp(-\gamma q) \cos((2k-1) \pi q/2s)$$

$$\gamma = \ln 9.$$

In this model the parameters are the segment length governed by the choice of $2m - 1$, and the rate of occurrence of the three-bond jump governed by the choice of w but usually expressed by τ_h which equals $(2w)^{-1}$. For the first time, the form of the model given here allows for any choice of m . Applications of this model have yielded sensible values for an apparent activation energy for backbone rearrangements based on τ_h (6, 8, 21). Over temperature intervals of 50° to 100°C , the segment length remains constant with values of the order of 5 to 15 bonds for polymers in solution, and segment length has been considered as a measure of length of chain involved in cooperative or coupled motions (6, 8, 13, 21). The Jones and Stockmayer model is not a continuous solution and becomes cumbersome for segment lengths of the order of hundreds which is encountered in some but not all solid rubbers.

The third lattice solution presented by Bendler and Yaris (22) allows the number of bonds to become a continuous variable with the parameters of the model being both a short and long range cutoff of motions expressed as an upper and lower frequency, W_A and W_B , respectively. The spectral density expression is

$$J(\omega) = \left(\frac{2}{\omega^{1/2} (W_B^{1/2} - W_A^{1/2})} \right) \int_{(W_A/\omega)^{1/2}}^{(W_B/\omega)^{1/2}} \frac{X^2}{(1 + X^4)} dX$$

$$X = k(W_2/\omega)^{1/2} \quad [5]$$

$$W_2 = 2w/5$$

w = three-bond jump rate.

When this model is applied to the interpretation of spin relaxation of polymers in solution, the extent of cooperative motion can be measured by a parameter $R = (W_A/W_B)^{1/2}$ which is found to take on values from 1 to 50. If a bond is the smallest moving unit, then R , called the "range", corresponds approximately to the number of bonds involved in cooperative or coupled motion (22). Both W_B and W_A are strongly temperature dependent and vary non-monotonically with temperature which appears to complicate this simple identification of R .

Table I.

COMPARISONS OF INTERPRETATIONS OF METHINE CARBON
RELAXATION IN DISSOLVED POLYSTYRENE

	T_1 (ms)	T_2 (ms)	NOE	Model Parameters
Experiment (10)	65	26	1.8	
VJGM (22)	66	25	1.8	$\tau_0 = 3.5 \times 10^{-8}$ s $\tau_D = 1.9 \times 10^{-9}$ s
Jones and Stockmayer	65	28	2.07	$\tau_h = 1.0 \times 10^{-9}$ s $(2m - 1) = 17$
Bendler and Yaris (22)	65	26	2.1	$W_A = 6.3 \times 10^6$ Hz $W_B = 4 \times 10^9$ Hz $R = 16.4$

Following Bendler and Yaris (22), it seems fruitful to compare the results of fitting the three lattice models to a common data set. A standard choice would be the experimental values obtained by Schaefer (10) on isotactic polystyrene in *o*-dichlorobenzene at 35°C. Table I contains the fitting parameters for the VJGM and Bendler and Yaris models (22) combined with the Jones and Stockmayer approach. All three models can account for the data within experimental error, and the only immediate comparison of model parameters is between the range of the Bendler and Yaris model and the segment length of the Jones and Stockmayer model. Given the rather different mathematical approaches of the two models, the range and segment length are surprisingly similar.

To compare the time scales of the dynamics characterization produced by each model, the spectral density or correlation function can be written as a distribution of exponential correlation times. For a correlation function, $\Phi(t)$, the general expression is

$$\Phi(t) = \int_0^{\infty} G(\tau) \exp(-t/\tau) d\tau \quad [6]$$

except in the Jones and Stockmayer model where the integral is replaced by a sum. With expressions given by the authors of each model, one can calculate the weighted inverse first moment from

$$(\tau_h^*)^{-1} = \int_0^{\infty} \tau^{-1} G(\tau) d\tau \quad [7]$$

which corresponds to the weighted harmonic average correlation time τ_h^* . Previously in the Jones and Stockmayer model, the unweighted harmonic average correlation time was labeled τ_h . For the VJGM model, the inverse first moment is infinite which indicates the presence of very many short correlation times produced by the continuous solution. For the Jones and Stockmayer model, a summation corresponding to Eq. 7 yields a value of 0.68 ns for τ_h^* based on the polystyrene interpretation. In the case of the Bendler and Yaris model, Eq. 7 reduces the simple expression

$$\tau_h^* = 3 / (W_A + W_B + (W_B W_A)^{1/2}). \quad [8]$$

The value of τ_h^* calculated from W_B and W_A corresponding to the polystyrene interpretation is 0.72 ns which is in good agreement with the time scale of the Jones and Stockmayer model.

One can also compare the spectral densities for the three models as was done by Bendler and Yaris for their model relative to the VJGM model (22). The comparison of the spectral densities produced by the three models using the parameters given in Table I

is shown in Fig. 1. All three spectral densities are of rather similar shape in the region of common NMR measurements which leads to the conclusion that it may not be possible to distinguish between the three lattice models on the basis of interpretive ability. The models may be better distinguished through ease of utilization and physical significance of model parameters. The VJGM model is easiest to apply requiring adjustment of only two continuous parameters in a concise equation. The Bendler and Yaris model involves a numerical integration while the Jones and Stockmayer model, a summation over discrete values. As mentioned, the physical significance of τ_D and τ_0 in the VJGM model is complicated, and the use of a continuous frequency distribution with an upper and lower cutoff is rather different from common dynamic characterizations. The concept of a "range" from the Bendler and Yaris model or segment length from the Jones and Stockmayer model is appealing but more applications are required to test any real significance.

Comparison of Polymer Dynamics

Since none of the lattice models is now clearly superior, the choice for interpretation of spin relaxation in polymers is arbitrary. Familiarity leads us to select the Jones and Stockmayer model so we will now consider application of this model to several well studied polymer systems in order to compare dynamics from polymer to polymer. Also the equations required to consider anisotropic internal rotation of substituent groups and overall molecular tumbling as independent motions in addition to backbone rearrangements caused by the three-bond jump are available for the Jones and Stockmayer model (13).

Some of the best studied dissolved polymers are polystyrene (6, 10, 23-28), polyisobutylene (5, 13, 29, 30), and poly(phenylene oxide) (7, 8). We will also add to these systems polyethylene which is an interesting reference point for polymers interpreted from a tetrahedral lattice viewpoint. The four systems will be abbreviated as PS, PIB, M_2 PPO, and PE in respective order of introduction here. The first of these systems, PS, has data available as a function of molecular weight, temperature, field strength, concentration and from three different types of nuclei. Similar large data bases are available for PIB and M_2 PPO, but the PE results are quoted from a proton and carbon-13 study as a function of molecular weight and temperature (31). The PE data are complicated by an unusual temperature dependence of the relaxation which precluded any estimation of an apparent activation energy. Within experimental error, both the proton and carbon-13 spin lattice relaxation times of PE dissolved in p-xylene are independent of temperature between 90 and 180°C. The proton and carbon-13 relaxation as a function of molecular weight can be consistently interpreted in terms of the model employed here, but no account of the temperature behaviour is offered.

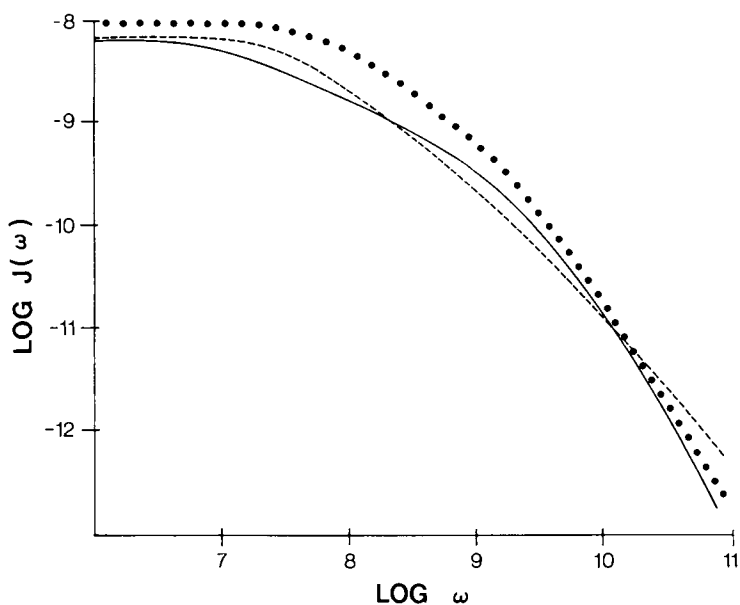


Figure 1. Logarithm of the spectral density vs. logarithm of the frequency, ω . The lines are the fit of the spectral density for the various models to relaxation observed in an *o*-dichlorobenzene solution of isotactic PS: (—), spectral density derived from the Bendler and Yaris model; (---), spectral density derived from the VJGM model; and (· · ·), spectral density derived from the Jones and Stockmayer model.

Table II.
COMPARISON OF BACKBONE DYNAMICS IN DILUTE SOLUTION

Polymer	Solvent	Temperature (°C)	τ_h (ns)	τ_a (ns)	E_a	Segment Length $2m - 1$
PE (21)	d-10 p- xylene	105	0.0052	0.0104	-	5
PIB (27)	CCl ₄	50	0.069	0.14	18 ± 4	5
PS (6)	CDCl ₃	50	0.24	0.72	20 ± 7	9
MePPO (8)	CDCl ₃	50	1.9	3.8	25 ± 5	5

The interpretation of the four systems is summarized in Table II, and the applicability of a lattice model is reasonable since all four backbones can be considered as being at least approximately characterized by a tetrahedral lattice. The correlation time for the three-bond jump, $\tau_h = (2w)^{-1}$, varies considerably among the four systems in general increasing with the size of the three-bond unit. The simple average correlation time, τ_a , also follows the same general trend. Apparent activation energies do not vary so greatly with the size and complexity of the three-bond unit. The segment length for cooperative or coupled motion is relatively short for these simple backbones in dilute solution. Note that in a different solvent, a somewhat longer segment length of 17 bonds produces the best fit for polystyrene (Table I). From another perspective, short segment lengths correspond to rather narrow distributions of correlation times.

Table III.

COMPARISON OF SUBSTITUENT GROUP ROTATION IN DILUTE SOLUTION

<u>Polymer</u>	<u>Group</u>	<u>T (°C)</u>	<u>τ_{ir} (ns)</u>	<u>E_a (kJ)</u>
PIB (<u>21</u>)	methyl	50	0.21	18 ± 5
PS (<u>28</u>)	phenyl	50	1	20 ± 5
M ₂ PPO (<u>8</u>)	phenyl	50	0.23	5 ± 2

In Table III, the characterization of anisotropic rotation of substituent groups is compared for this same collection of polymers. In dilute solution, all three polymers have similar time scales for this motion as indicated by the value of the internal rotation correlation time, τ_{ir} . However, the activation energy for substituent group rotation in PIB and PS is much higher than the activation energy for phenyl group rotation in M₂PPO. This indicates some real differences in the nature of the two local motions. The correlation time for phenyl group rotation in polystyrene is rather uncertain since it is calculated from a very small difference between T_1 values. This same problem precludes a very accurate estimate of the activation energy although some attempts have been made (24, 27, 28).

The relationship between internal rotation of substituents and backbone rearrangements can be considered from the interpretation. The time scales of anisotropic internal rotation and backbone rearrangements are well separated in M₂PPO. In addition, the concentration and temperature dependences of these two quantities are quite different leading us to conclude that the motions are independent. In PIB and PS, internal rotation and backbone

Table IV.

<u>Polymer</u>	<u>Nuclei</u>	<u>Dominant Motional Source of Spin Relaxation</u>	<u>Relationship between Backbone Rearrangement and Internal Rotation</u>
PE (<u>27</u>)	Methylene carbon	backbone rearrangement	
PIB (<u>21</u>)	Methylene carbon	backbone rearrangement	independent
	Methyl carbon	backbone rearrangement	
PS (<u>6</u>)	Methylene carbon	backbone rearrangement	?
	Phenyl carbons	backbone rearrangement	
M ₂ PPO (<u>8</u>)	Phenyl carbon	phenyl group rotation	independent
	Methyl protons	phenyl group rotation	

rearrangements are close in time scale, and the activation energy for methyl group rotation in PIB is equal to the activation energy for backbone rearrangement. However, analysis of the concentration dependence of motions in PIB points to the independence of the two motions (21). The time scale of backbone rearrangements changes by orders of magnitude in traversing the concentration range from dilute solution to the bulk while the time scale of methyl group rotation remains almost constant. This interpretation is at odds with one presented by Heatley (30) but data at two field strengths support the interpretation voiced here. Table IV attempts to summarize these comparisons between backbone rearrangements and substituent group rotation. It also lists the motion which is the major source of spin relaxation through modulation of the dipole-dipole interaction. In only M_2 PPO does internal rotation of a substituent group, the phenyl group, become the dominant source of spin relaxation.

Table V.

COMPARISON OF BACKBONE REARRANGEMENTS IN SOLIDS

<u>Polymer</u>	<u>Temperature</u>	<u>τ_h (ns)</u>	<u>τ_a (ns)</u>	<u>Segment Length $2m - 1$</u>
PE (amorphous) (32)	45°	.030	.060	5
PIB (rubber) (5)	45°	11	22	5
Cis-Polyisoprene (rubber) (10)	35°	0.35	5.25	57
Cis-Polybutadiene (rubber) (10)	35°	0.0065	3.7	23×10^2

Only recently have the lattice models been applied to solid amorphous and rubbery polymers (21-22). Table V contains a summary of the interpretations of several solid polymers. In general, less extensive data are available on these systems. For PE (32) and PIB (5), the interpretation is based on carbon-13 T_1 and NOE values.

For cis-polyisoprene and cis polybutadiene, the interpretation is based on T_1 , NOE and T_2 values (10) although utilization of T_2 is complicated because of the possible presence of systematic errors particularly in spectrometers employing superconducting magnets. The interpretation for PE and to a lesser extent PIB is not unique. Several choices of segment length are possible and the parameters listed in Table V are for a minimum length which

accounts for T_1 and the NOE. The other possible choices of segment length are not much longer since a maximum NOE is observed in PE, and the PIB data are consistent with a very narrow distribution of correlation times (5, 21). Of course coupling between chains is completely neglected in this lattice model so the significance of segment lengths involved in cooperative local motions is unclear. For cis-polyisoprene and cis-polybutadiene, the choice of parameters accounting for T_1 , the NOE, and T_2 is unique. However, the segment length is determined entirely by matching T_2 which is the least reliable piece of input information although great care was taken by Schaefer (10) to limit systematic errors. In PE, the NOE and apparent T_2 depend on the degree of crystallinity (32) and mostly amorphous PE is considered here. As crystallinity increases, the NOE and T_2 decrease which corresponds to an increase in coupling segment length. However, the decrease in T_2 may well reflect macroscopic and microscopic inhomogeneities in magnetic field strength across the sample caused by instrumental problems and sample preparation. These kinds of complications led us and others (2) to doubt the significance of dynamic interpretations which rest strongly on the observed values of T_2 . It is preferable to develop another measure sensitive to low frequency motions which is less susceptible to systematic errors. Possibly $T_{1\rho}$ could play this role in viscous solution and bulk materials as it is now providing insight into glassy materials (33).

One last comment on the interpretation of cis-polyisoprene and cis-polybutadiene is appropriate. Models based on a tetrahedral lattice of equivalent bonds are clearly not strictly applicable to a polymer containing both single and double bonds. Thus, model parameters including the coupling segment length should not be taken too seriously although it is very nearly equal to the ranges obtained by Bendler and Yaris employing their model. One can be somewhat hopeful about the application of the lattice models to non-tetrahedral backbones if the connectivity of the chain is the major factor determining the correlation function and not the tetrahedral geometry (34).

Summary

The lattice models provide useful interpretations of spin relaxation in dissolved polymers and rubbery or amorphous bulk polymers. Very large data bases are required to distinguish the interpretive ability of lattice models from other models, but as yet no important distinction between the lattice models is apparent. In solution, the spectral density at several frequencies can be determined by observing both carbon-13 and proton relaxation processes. However, all the frequencies are rather high unless T_2 data are also included which then involves the prospect of systematic errors. It should be mentioned that only effective rotational motions of either very local or very long range nature are required to account for solution observations. The local

motions which result in rotational averaging of dipolar interactions are the backbone rearrangements caused by the three-bond jump or anisotropic rotation of substituent groups. The only long range motion included is overall rotatory diffusion. Together these account for the molecular weight dependence of spin relaxation although mid-range motions corresponding to Rouse-Zimm modes of an order higher than one are not included. The effects of local motions are seen in the data at high molecular weight and the effects of overall rotatory diffusion, at low molecular weights. However, it has not been necessary to explicitly include other types of cooperative, long range motions. Possibly $T_{1\rho}$ would be sensitive to motions corresponding to the higher Rouse-Zimm modes, and this would aid in the elucidation of the role of these motions in spin relaxation.

The other class of motion only now being introduced into interpretive models is oscillatory motion. Anisotropic oscillatory motions of substituent groups have been considered by Chachaty (12) but not in conjunction with a lattice description of backbone motion. No attempt to develop a model based on oscillatory backbone rearrangements is known to these authors, and this avenue may be very important for the interpretation of concentrated solutions, rubbery or amorphous solids, and especially glassy polymers (32).

Acknowledgments

Many helpful discussions with W. H. Stockmayer are greatly appreciated. The research was carried out with financial support of the National Science Foundation, Grant DMR7716088, Polymers Program. This research was supported in part by a National Science Foundation Equipment Grant No. CHE77-09059.

Abstract

A brief survey of models for the interpretation of spin relaxation in polymers suggests models based on the occurrence of the three-bond jump on a tetrahedral lattice are capable both of accounting for observations and providing some physical insight. The lattice model of Valeur, Jarry, Geny and Monnerie is compared with the more recent revisions of Jones and Stockmayer, and Bendler and Yaris. Since it is found that all three lattice models have comparable interpretive ability and produce very similar descriptions of the spectral density when applied to the same data, one model, the Jones and Stockmayer version, was used to interpret several well studied polymers. The resulting characterization of motions in dissolved polyethylene, polyisobutylene, polystyrene and poly(phenylene oxide) are reviewed for trends in time scale, apparent activation energy and the extent of cooperative motion. Time scales varied from picoseconds to nanoseconds, the activation energies for backbone rearrangements are all about

20kJ, and the length of chain involved in cooperative motion is only 5 to 15 bonds. Spin relaxation in four solid polymers was also interpreted with the model. Amorphous polyethylene and polyisobutylene rubber undergo motion nearly as rapid as dissolved polymers and the segment length for cooperative motion is not appreciably longer either. Cis-polyisoprene and cis-polybutadiene are also very mobile as solid rubbers but the apparent segment length for cooperative motion is much longer than for simple dissolved polymers. Of course, a tetrahedral lattice model is not strictly applicable to these last two polymers, and interchain cooperativity was not properly considered for any of the solid polymers.

Literature Cited

1. For a recent overview, Schaefer, J. in "Topics in Carbon-13 NMR Spectroscopy," Vol. 1, G. C. Levy, Ed. (Wiley-Interscience, New York, 1974).
2. Heatley, F. and Cox, M. K., Polymer, (1977), 18, 225.
3. Doddrell, D., Glushko, V. and Allerhand, A., J. Chem. Phys., (1972), 56, 3683.
4. Bloembergen, N., Purcell, E. M. and Pound, R. V., Phys. Rev., (1948), 73, 679.
5. Komoroski, R. A. and Mandelkern, L., J. Polym. Sci., (1976), Sym. No. 54, 201.
6. Matsuo, K., Kuhlmann, K. F., Yang, H. W.-H., Geny, F., Stockmayer, W. H. and Jones, A. A., J. Polym. Sci., Polym. Phys. Ed., (1977), 15, 1347.
7. Laupretre, F. and Monnerie, L., Eur. Polym. J., (1974), 10, 21.
8. Jones, A. A. and Lubianez, R. P., Macromolecules, (1978), 11, 126.
9. Woessner, D. E., J. Chem. Phys., (1962), 36, 1.
10. Schaefer, J., Macromolecules, (1973), 6, 882.
11. Heatley, F. and Begum, A., Polymer, (1976), 17, 399.
12. Ghesquiere, D., Ban, B. and Chachaty, C., Macromolecules, (1977), 10, 743.
13. Jones, A. A. and Stockmayer, W. H., J. Polym. Sci., Polym. Phys. Ed., (1977), 15, 847.
14. Valeur, B., Jarry, J. P., Geny, F. and Monnerie, L., J. Polym. Sci., Polym. Phys. Ed., (1975), 13, 667.
15. Valeur, B., Monnerie, L. and Jarry, J. P., J. Polym. Sci., Polym. Phys. Ed., (1975), 13, 675.
16. Valeur, B., Jarry, J. P., Geny, F. and Monnerie, L., J. Polym. Sci., Polym. Phys. Ed., (1975), 13, 2251.
17. Glarum, S. H., J. Chem. Phys., (1960), 33, 639.
18. Hunt, B. I. and Powles, J. G., Proc. Phys. Soc., (1966), 88, 513.
19. DuBois-Violette, E., Geny, F., Monnerie, L. and Parodi, O., J. Chem. Phys., (1969), 66, 1865.
20. Schilling, F. C., Cais, R. E. and Bovey, F. A., Macromole-

- cules, (1978), 11, 325.
21. Jones, A. A., Lubianez, R. P., Hanson, M. A. and Shostak, S. L., to appear J. Polym. Sci., Polym. Phys. Ed., (1978).
 22. Bendler, J. and Yaris, R., to appear Macromolecules, (1978).
 23. Allerhand, A. and Hailstone, R. K., J. Chem. Phys., (1972), 56, 3718.
 24. Laupretre, F., Noel, C. and Monnerie, L., J. Polym. Sci., Polym. Phys. Ed., (1977), 15, 2127.
 25. Inoue, Y. and Konno, T., Polym. J., (1976), 8, 457.
 26. Heatley, F. and Begum, A., Polymer, (1976), 17, 399.
 27. Jones, A. A., J. Polym. Sci., Polym. Phys. Ed., (1977), 15, 863.
 28. Matsuo, K. and Stockmayer, W. H., private communication.
 29. Inoue, Y., Nishioka, A. and Chujo, R., J. Polym. Sci., Polym. Phys. Ed., (1973), 11, 2237.
 30. Heatley, F., Polymer, (1975), 16, 493.
 31. Gerr, F. E. and Jones, A. A., unpublished results.
 32. Komoroski, R. A., Maxfield, J., Sakaguchi, F. and Mandelkern, L., Macromolecules, (1977), 10, 550.
 33. Schaefer, J., Stejskal, E. O. and Buchdahl, R., Macromolecules, (1977), 10, 384.
 34. Stockmayer, W. H., private communication.

Discussion

D. Axelson, Florida State University, Florida: One comment and then one question. The polyethylene sample you showed in the last slide I've taken below 45° down to -40°C. The sample develops a broad distribution or non-exponential auto correlation function at low temperatures. One of the giveaways is that in going through the T₁ minimum if it is not 110 ms at 67 MHz and if a distribution exists, it will be higher, and in this case the minimum is almost at 300 ms. The system very quickly develops a distribution and the T₁'s virtually level off for the last 20 or 35 degrees. Even this macromolecule sample is tremendously complicated. The other point is related to the methyl group rotation you were talking about. Some of the polymers we have been looking at in solution as well as in bulk have long side chains. The end methyl with a six or eight carbon alkyl side chain has a T₁ value anywhere from twenty seconds on down. The T₁ value is frequency dependent. It shouldn't be frequency dependent over 200 or 300 ms. I was wondering if you have any feel for what physically is taking place or if you have had any opportunity to investigate models that would try to alleviate this peculiarity,

A. Jones, Clark University, Massachusetts: When the methyl group is attached to the backbone or even to a side chain it has a whole distribution of correlation times associated either with the backbone or the side chain in addition to its methyl group rotation. Our assumption in these models is that they are independent so that the methyl group T₁ would be calculated on the

basis of a very large distribution of correlation times even though there probably is a fast correlation time that is dominant. All those other times are in the correlation function and that may be what you are detecting. I'd like to point out that in some polyethylene data the molecular weight dependence indicated that at a molecular weight of 150, half of the T_1 was still associated with backbone rearrangements which involves a distribution of correlation times. Even a short side chain may have a distribution of correlation times associated with it which you do not observe until the data is closer to the T_1 minimum than in the polyethylene data considered here. It is transparent until you get a sufficient data base.

D. Axelson: We published a paper (G. C. Levy, D. E. Axelson, R. Schwartz and J. Hochmann, *J. Am. Chem. Soc.*, 100, 410 (1978)) late last year on methacrylates in which we tried to interpret the side chains in terms of the effect of the backbone distribution. The side chain was governed by a combination of the distribution and the multiple internal rotations. The problem we had is that when you get to the end of the chain you still can't account for the frequency dependence. We have not proceeded to the stage of doing dilute solution studies because as you mentioned, there may be an intermolecular effect with other chains.

A. Jones: I don't feel the multiple internal rotations model is applicable for side chains any longer than 3 or 4 carbons. As soon as it is longer than 3-4 carbons, a crankshaft type of motion dominates instead of multiple internal rotations. The viewpoint probably reflects some bias on my part.

J. Prud'homme, University of Montreal, Que.: What is the physical explanation for the leveling? When we see the curve of T_1 as a function of molecular weight there is a plateau. This appears at a molecular weight over 1000 and sometimes 10,000. What is the physical explanation for this?

A. Jones: With low molecular weights, the overall molecular tumbling is at a rate not far from the Larmour frequency, the basic frequency of the experiment. As molecular weight increases these overall tumbling motions become very slow and very far removed from the Larmour frequency. The motion closest to the Larmour frequency comes from backbone rearrangements, very local motions, which do not depend upon molecular weight. That three bond jump in the model does not depend on chain length, it only involves three bonds in the middle of a long chain. When those motions are dominant, the relaxation closest to the Larmour frequency stems from local motions yielding molecular weight independent relaxation.

J. Prud'homme: What is amazing is that for some substances this happens at a very high molecular weight.

A. Jones: If the backbone motion is very slow then higher molecular weights are required before the overall tumbling motion becomes much slower than the relatively slow backbone motions. Dr. Bovey mentioned polysulfones. I would expect T_1 to have an extended molecular weight dependence because the backbone motions are relatively slow in this case.

W. G. Miller, University of Minnesota, Minnesota: In visco elastic studies the monomeric friction coefficient is used to describe motion. The same parameter is used to look at translational diffusion of solvent and its concentration dependence. Is there any relationship between this parameter and your three bond motion or is the correlation length way too long?

A. Jones: I don't think I can give you a definitive answer. The three-bond jump should be experiencing a friction partially determined by its environment, i.e., solvent or other chains around it, but most probably it is determined by the rotational potential associated with conformational changes within the chain. For many systems we see rather little concentration dependency of the activation energy for these motions. For instance, the phenyl group rotation which has a very low barrier appears to be a good deal more solvent dependent than some of the other backbone rearrangements involving three bond jumps. Most others seem to be mostly influenced by the shape of potential energy surface associated with the chain conformations.

I. C. P. Smith, NRC - Ontario: When you are testing those models, crankshafts, cut off and so on, there was a paucity of data on frequency dependence. Have you tried to fit the various models to say three magnetic fields?

A. Jones: Yes, I didn't show all the data. Most of the data is from the literature. Polystyrene data and polyisobutylene data at two field strengths can be accounted for by these lattice models based upon the three bond jump. Some recent data by Dr. Bovey on the polybutene was more difficult to fit with regard to the frequency dependence. (F. C. Schilling, R. E. Cais and F. A. Bovey, *Macromolecules* **11**, 325 (1978).) Frequency dependent data I think is important information to acquire when trying to understand the dynamics.

I. C. P. Smith: But does it distinguish between the models? It would be nice to find the best of the three.

A. Jones: It does not distinguish between the lattice models. I don't think we are going to easily distinguish between the lattice models. The same basic equations are employed in each case. The three lattice models I mentioned have spectral densities of the same shape.

P. Sipos, Dupont, Ontario: Using the concept of a chain segment you have shown large differences between polyisobutylene

and other polymer systems. Is it that the basic relaxation mode occurs on a long chain because a very short segment oscillates independently or is it a concerted phenomenon? To get agreement must a large number of segments be considered together?

A. Jones: We would like to know whether it is a long range oscillation or whether it is a complete turning around of a series of units. There are measures of whether it is a complete rotation such as the values of NT_1 for different carbons. If NT_1 is a constant, the motion is probably rotational. In solution, I think these things tend to be rotational. As you go to bulk systems and in particular glassy systems, J. Schaefer (J. Schaefer, E. O. Stejskal and R. Buchdahl, *Macromolecules* 10, 384 (1977)) feels very strongly that oscillatory motion predominates. I would agree, although I can't produce an oscillatory model to interpret the data.

RECEIVED March 13, 1979.

Carbon-13 NMR and Polymer Stereochemical Configuration

JAMES C. RANDALL

Phillips Petroleum Company, Bartlesville, OK 74004

With the discovery of crystalline polypropylene in the early 1950's, polymer stereochemical configuration was established as a property fundamental to formulating both polymer physical characteristics and mechanical behavior. Although molecular asymmetry was well understood, polymer asymmetry presented a new type of problem. Both a description and measurement of polymer asymmetry were essential for an understanding of the polymer structure.

Technically, each methine carbon in a poly(1-olefin) is asymmetric; however, this asymmetry cannot be observed because two of the attached groups are essentially equivalent for long chains. Thus a specific polymer unit configuration can be converted into its opposite configuration by simple end-to-end rotation and subsequent translation. It is possible, however, to specify relative configurational differences and Natta introduced the terms isotactic to describe adjacent units with the same configurations and syndiotactic to describe adjacent units with opposite configurations (1). Although originally used to describe dyad configurations, isotactic now describes a polymer sequence of any number of like configurations and syndiotactic describes any number of alternating configurations. Dyad configurations are called meso if they are alike and racemic if they are unlike (2). Thus from a configurational standpoint, a poly(1-olefin) can be viewed as a copolymer of meso and racemic dyads.

The measurement of polymer configuration was difficult and sometimes speculative until the early 1960's when it was shown that proton NMR could be used, in several instances, to define clearly polymer stereochemical configuration. Bovey was able to identify the configurational structure of poly(methylmethacrylate) in terms of the configurational triads, mm, mr and rr, in a classic example (3). In the case of polypropylene, configurational information appeared available but was not unambiguously accessible because severe overlap complicated the identification of resonances from the mm, mr and rr triads (4). Several papers appeared on the subject of polypropylene tacticity but none totally resolved the problem (5).

0-8412-0505-1/79/47-103-291\$07.00/0
© 1979 American Chemical Society

With the advent of C-13 NMR in the early 1970's, the measurement of polymer stereochemical configuration became routine and reasonably unambiguous.

The advantages of C-13 NMR in measurements of polymer stereochemical configuration arise primarily from a useful chemical shift range which is approximately 20 times that of proton NMR. The structural sensitivity is enhanced through an existence of well separated resonances for different types of carbon atoms. Overlap is generally not a limiting problem. The low natural abundance (~1%) of C-13 nuclei is another favorably contributing factor. Spin-spin interactions among C-13 nuclei can be safely neglected and proton interactions can be eliminated entirely through heteronuclear decoupling. Thus each resonance in a C-13 NMR spectrum represents the carbon chemical shift of a particular polymer moiety. In this respect, C-13 NMR resembles mass spectrometry because each signal represents some fragment of the whole polymer molecule. Finally, carbon chemical shifts are well behaved from an analytical viewpoint because each can be dissected, in a strictly additive manner, into contributions from neighboring carbon atoms and constituents. This additive behavior led to the Grant and Paul rules (6), which have been usefully applied in polymer analyses, for predicting alkane carbon chemical shifts.

The advantages so clearly evident when applying C-13 NMR to polymer configurational analyses are not devoid of difficulties. The sensitivity of C-13 NMR to subtle changes in molecular structure creates a wealth of chemical shift-structural information which must be "sorted out". Extensive assignments are required because the chemical shifts relate to sequences from three to seven units in length. Model compounds, which are often used in C-13 analyses, must be very close structurally to the polymer moiety reproduced. For this reason, appropriate model compounds are difficult to obtain. A model compound found useful in polypropylene configurational assignments was a heptamethylheptadecane where the relative configurations were known (7). To be completely accurate, the model compounds should reproduce the conformational as well as the configurational polymer structure. Thus reference polymers such as predominantly isotactic and syndiotactic polymers form the best model systems. Even when available, only two assignments are obtained from these particular polymers. Pure reference polymers can be used to generate other assignments as will be discussed later (5).

To obtain good quantitative C-13 NMR data, one must understand the dynamic characteristics of the polymer under study. Fourier transform techniques combined with signal averaging are normally used to obtain C-13 NMR spectra. Equilibrium conditions must be established during signal averaging to ensure that the experimental conditions have not led to distorted spectral information. The nuclear Overhauser effect (NOE), which arises from H-1, C-13 heteronuclear decoupling during data acquisition, must also be considered.

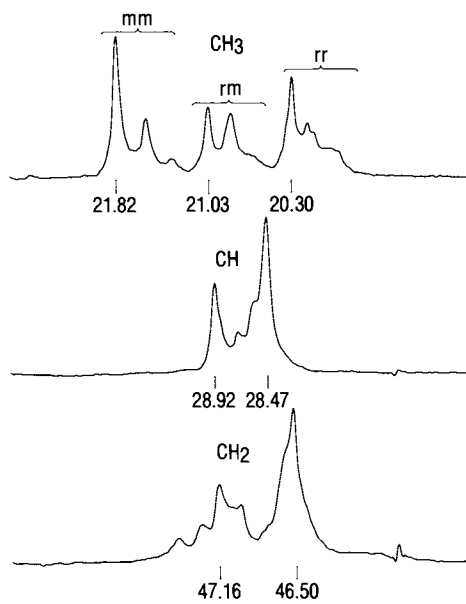
Energy transfer, occurring between the H-1 and C-13 nuclear energy levels during spin decoupling, can lead to enhancements of the C-13 resonances by factors between 1 and 3. Thus the spectral relative intensities will only reflect the polymer's moiety concentrations if the NOE's are equal or else taken into consideration. Experience has shown that polymer NOE's are generally maximal, and consequently equal, because of a polymer's restricted mobility (8) (9). To be sure, one should examine the polymer NOE's through either gated decoupling or paramagnetic quenching and thereby avoid any misinterpretation of the spectral intensity data.

Let us now discuss C-13 NMR spectra from a series of vinyl polymers to survey the information available concerning polymer stereochemical configuration. We will later return to the topics of how C-13 structural sensitivity is established and how assignments are made. As mentioned earlier, the C-13 configurational sensitivity falls within a range from triad to pentad for most vinyl polymers. In noncrystalline polypropylenes, three distinct regions corresponding to methylene (~46 ppm) methine (~28 ppm) and methyl (~20 ppm) carbons are observed in the C-13 NMR spectrum. (Throughout this discussion and in the ensuing discussions, the chemical shifts are reported with respect to an internal tetramethylsilane (TMS) standard.) The C-13 spectrum of a typical amorphous polypropylene is shown in Figure 1.^a Although a configurational sensitivity is shown by all three spectral regions, the methyl region exhibits by far the greatest sensitivity and is consequently of the most value. At least ten resonances, assigned to the unique pentad sequences, are observed in order, mmmmm, mmmmr, rmmr, mmrr, mrrm, rrrr, rrrm and mrrm, from low to high field (7) (10) (11). These assignments will be discussed in more detail later.

The C-13 spectrum of polystyrene, shown in Figures 2 and 3, contains two regions where stereochemical information can be extracted. There are nine methylene resonances and at least 20-22 aromatic quaternary carbon resonances. No other carbons in polystyrene exhibit a configurational sensitivity. Tentative assignments have been made for the methylene carbons based on an assumed Bernoullian behavior (12).

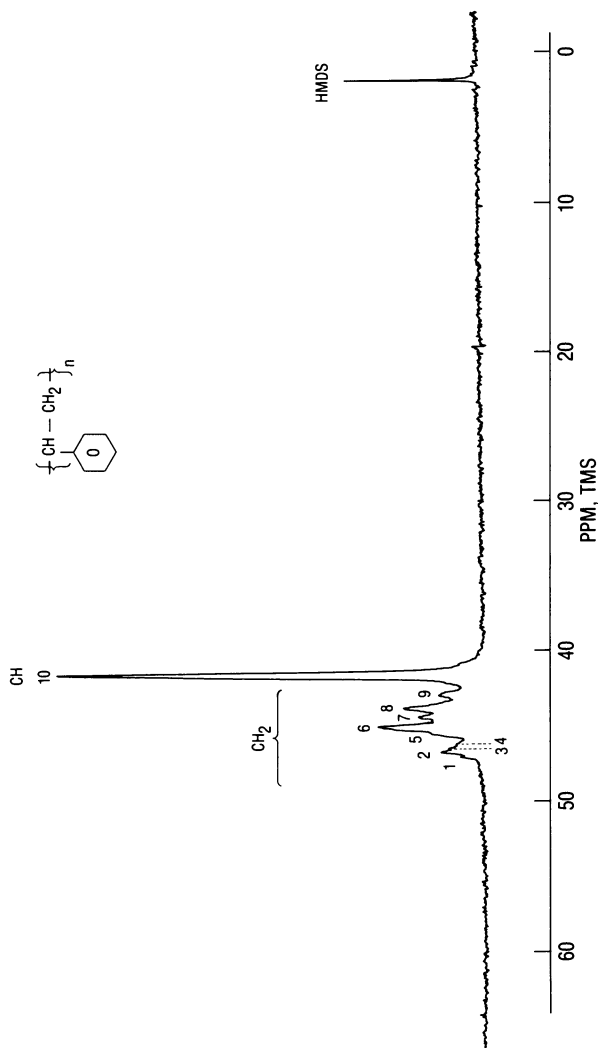
As shown in Figure 4, the methylene and methine carbons of polyvinylchloride show a sensitivity toward configuration and complete, internally consistent assignments have been given by Carman (13). Likewise a similar situation exists in the C-13 NMR spectrum of polyvinylalcohol, shown in Figure 5, where assignments have been given by Wu and Ovenall (14). The general trend among vinyl polymers is for the methylene carbons to exhibit a greater configurational sensitivity than the methine carbons.

^a This spectrum and others shown in this paper were taken from polymers dissolved in 1,2,4-trichlorobenzene at 125°C.



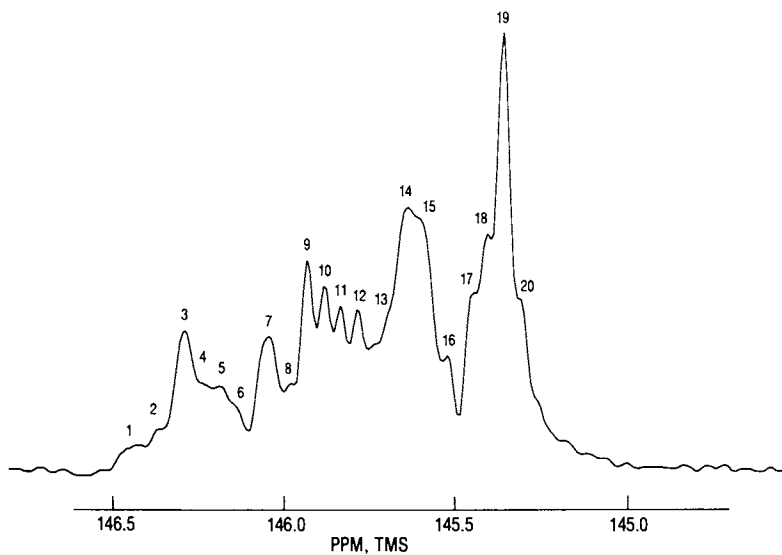
Academic Press

Figure 1. Methyl, methine, and methylene regions of the C-13 NMR spectrum of a noncrystalline PP (30)



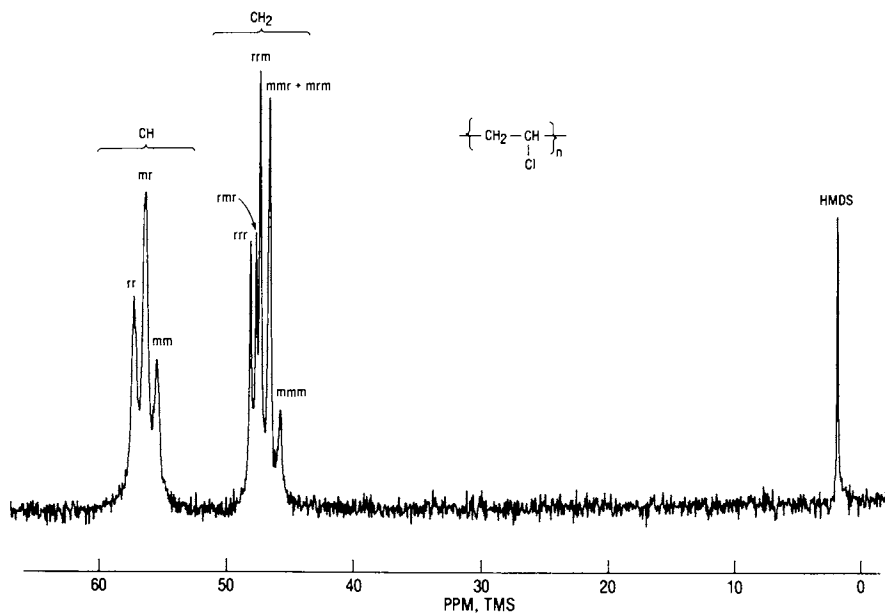
Academic Press

Figure 2. The methylene and methine regions of the C-13 NMR spectrum of PS prepared with a free radical initiator. The internal standard is HMDS, which occurs at 2.03 ppm with respect to TMS (30).



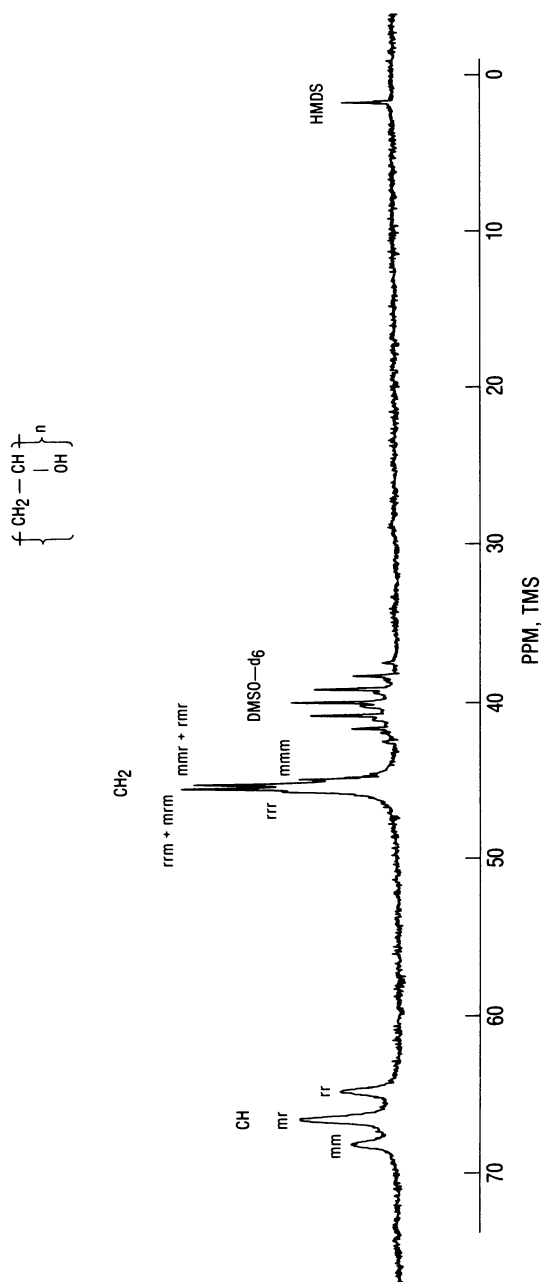
Academic Press

Figure 3. The aromatic quaternary resonances from a free radical PS shown in Figure 2 (30)



Academic Press

Figure 4. $C-13$ NMR spectrum at 25.2 MHz of PVC (30). See Figure 2 for an explanation of the scale.



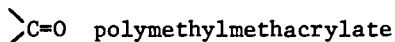
Academic Press

Figure 5. C-13 NMR spectrum at 25.2 MHz of PVC (30). See Figure 2 for an explanation of the scale.

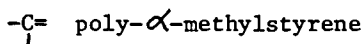
Exceptions do exist as shown in the spectrum of polyacrylonitrile in Figure 6. The methine resonances show a distinct triplet with very little splitting exhibited by the methylene resonances. The greatest sensitivity toward configuration occurs for the nitrile resonances where an almost ideal Bernoullian distribution is observed.

In general, the trends observed among configurational assignments cannot be used to make assignments in a system where the assignments are unknown. Certainly, similarities do exist as shown by the data in Tables I, II and III; however, there are no obvious explanations for some of the glaring exceptions. The methine resonances in polyvinylchloride occur as rr, mr and mm from low to high field. A similar trend appears reasonable for polyacrylonitrile; however the methine resonances in polyvinylalcohol have been shown to occur in the inverse order, mm, mr and rr. In an analogous manner, the methine carbon resonances from polypropylene show a pentad sensitivity with the mmmmm pentad occurring at low field. In this instance, the observed pentads may occur in an order similar to the methyl pentad resonances (15).

An example where independent, but similar, assignments were obtained occurs for the side-chain carbonyl carbons in polymethylmethacrylate (16) and the quaternary aromatic carbons of poly- α -methylstyrene (17). Both are sp^2 carbons; otherwise they are in completely different environments. The assignments, as they occur from low to high field, are:



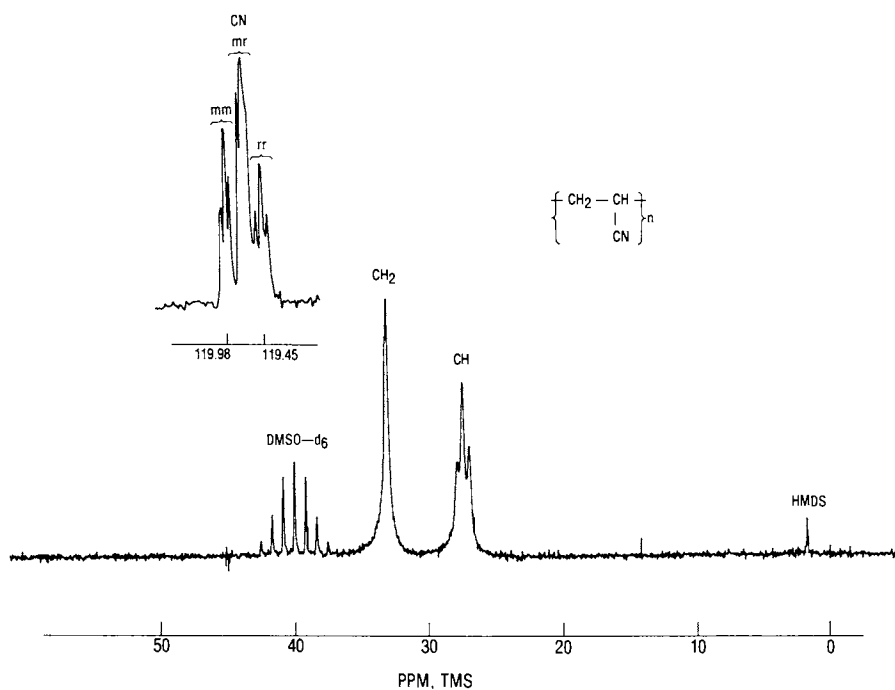
mrrm, rrrm, rrrr, rrrm+mmrm, rrrr+mmrr, mmmmm, mmmmr, rmmr



mrrm, rrrm, rrrr, rrrr, mrrr, mrrm, rrrm+mmrr, mmmmm +mmmr +rmmr

Listed in Tables I, II and III are representative examples of the chemical shift behavior for both the backbone methine, methylene carbons and for the various pendant side-chain carbons.

In almost all of the vinyl polymers examined so far, an unambiguous source for configurational information has been found (18). For example, the backbone methylene resonances for the various polyvinylethers show a basic dyad sensitivity with the r resonance occurring downfield from the m resonance (19) (20). In poly-(methylacrylonitrile) one can utilize the methyl resonances (21). The backbone carbons yield configurational information in the C-13 spectra of the various polyalkylacrylates (22) (23). The extraction of configurational information, of course, depends upon the availability of correct assignments in spectra which are often detailed and complex. Let us now turn to the determination of the various spectral sensitivities and the establishment of configurational assignments.



Academic Press

Figure 6. ^{13}C NMR spectrum at 25.2 MHz of PAN (30). See Figure 2 for an explanation of the scale.

Table I

The Triad Chemical Shift Sequence with Respect to an Increasing Field Strength for Some Representative Vinyl Polymer Backbone Methine Carbon Resonances

	δ_{-CH-}		
	\xrightarrow{H}		
Poly(vinylchloride)	rr	mr	mm
Poly(isopropyl acrylate)	rr	mr	mm
Poly(vinyl alcohol)	mm	mr	rr
Polypropylene	mm	mr	rr
Poly(vinyl acetate)	mm	(mr)	(rr)

Polymers with only Singlet Backbone Methine Resonances

Polystyrene
 Poly(ethyl vinyl ether)
 Poly(isobutyl vinyl ether)
 Poly(methyl acrylate)
 Poly(methyl vinyl ether)

Table II

The Triad Chemical Shift Sequence with Respect to an Increasing Field Strength for Some Representative Vinyl Polymer Side-Chain Carbon Resonances

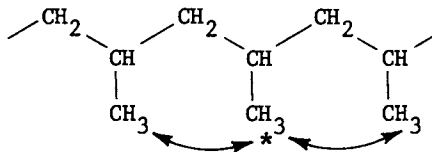
	δ_C		
	\xrightarrow{H}		
Polyacrylonitrile (CN)	mm	mr	rr
Poly(tertiary vinyl ether) (-C-)	mm	mr	rr
Polypropylene (CH ₃)	mm	mr	rr
Polystyrene (-C=)	mm	--	--
Poly(methyl vinyl ether) (OCH ₃)	rr	mr	mm

Table III

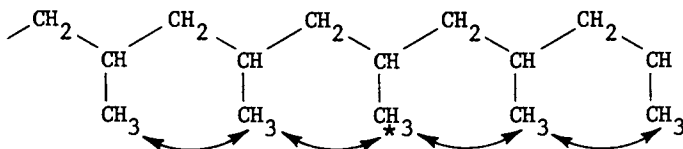
The Tetrad Chemical Shift Sequence with Respect to an Increasing Field Strength for Some Representative Vinyl Polymer Backbone Methylene Carbon Resonances

	δ -CH ₂ - Poly(isopropyl acrylate)					
→ H	rrr +	rmr,	mmr +	mrn +	mrr,	mmn
	δ -CH ₂ - Poly(methyl acrylate)					
→ H	rrr	rmr,	mmr +	mrr,	mrn +	mmn
	δ -CH ₂ - Polypropylene					
→ H	mrn	rrr	mrr		m	
	δ -CH ₂ - Polystyrene					
→ H	mrr	(rmr)	mmr	(mrn)	mmn	(rrr)
	δ -CH ₂ - Poly(vinyl acetate)					
→ H	rrr	mrr	mrn +	rmr,	mmr	mmn
	δ -CH ₂ - Poly(vinyl alcohol)					
→ H	rrr	mrr +	mrn,	rmr +	mmr,	mmn
	δ -CH ₂ - Poly(vinyl chloride)					
→ H	rrr	rmr	mrr	mrn +	mmr,	mmn
	δ -CH ₂ - Poly(ethyl vinyl ether)					
→ H		r			m	
	δ -CH ₂ - Poly(isobutyl vinyl ether)					
→ H		r			m	
	δ -CH ₂ - Poly(methyl vinyl ether)					
→ H		r			m	

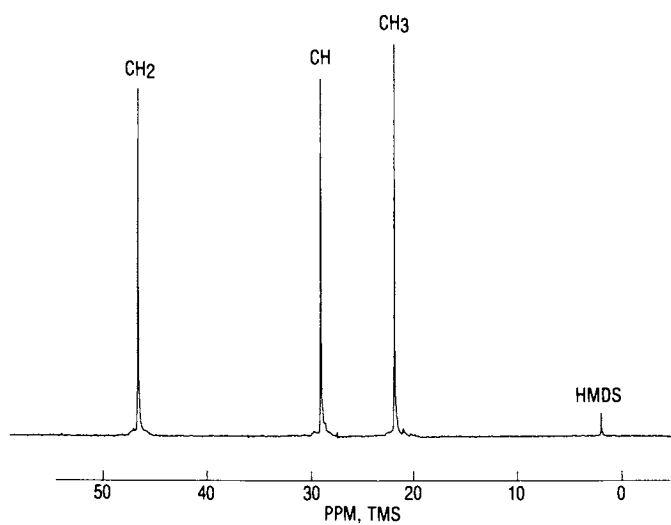
The vinyl polymer studied most thoroughly with respect to configuration has been polypropylene (5) (7) (10) (11) (24) (25) (26). The C-13 NMR spectrum of a crystalline polypropylene shown in Figure 7 contains only three lines which can be identified as methylene, methine and methyl from low to high field by off-resonance decoupling. An amorphous polypropylene exhibits a C-13 spectrum which contains not only these three lines but additional resonances in each of the methyl, methine and methylene regions as shown previously in Figure 1. The crystalline polypropylene must, therefore, be characterized by a single type of configurational structure. In this case, the crystalline polypropylene structure is predominantly isotactic, thus the three lines in Figure 7 must result from some particular length of meso sequences. This sequence length information is not available from the spectrum of the crystalline polymer but can be determined from a corresponding spectrum of the amorphous polymer. To do so, one must examine the structural symmetry of each carbon atom in the various possible monomer sequences. Let us begin by an inspection of the polypropylene methyl group in triad and pentad configurational environments. (These arguments can be applied, of course, in a related discussion of any vinyl polymer.) If the methyl group chemical shift is sensitive to just nearest neighbor configurations, then the simplest configurational sensitivity must be triad, that is,



There are only three unique triad combinations, mm, mr and rr; thus a methyl configurational sensitivity to just nearest neighbor configurations would produce only three resonance in the methyl region of the C-13 spectrum. From an earlier spectrum of the amorphous polymer, we noted at least ten methyl resonances. We must therefore consider the situation where the next-nearest as well as nearest neighbor configurations are affecting the chemical shift, that is,



An inspection of the number of unique configurations taken five at a time (pentads) shows that there are ten unique pentad configurations: mmmmm, mmmmr, rmmmr, mmmrm, mmrrr, rmmrr, mrrrm, mrrrr and



Academic Press

Figure 7. *C-13* NMR spectrum at 25.2 MHz of crystalline PP (30). See Figure 2 for an explanation of the scale.

rrrr. Note that there are three pentads with common mm centers, three with common rr centers and four with common mr triad centers. By now, we can see a pattern developing. The side chain groups of a vinyl polymer will be configurationally sensitive to an odd sequence of structural units; that is, three, five, seven, etc. The number of unique combinations (N) for a particular sequence length (n) can be predicted with the following equation (27):

$$N_{(n)} = 2^{n-2} + 2^{(n-3)/2}$$

Therefore, when

$$\begin{array}{ll} n = 3, & N = 3 \\ n = 5, & N = 10 \\ n = 7, & N = 36 \quad \text{etc.} \end{array}$$

These same arguments apply to the polymer backbone methine carbons where the structural sensitivity, required by symmetry, will be to an odd number of continuous units.

In a similar manner, it can be shown that the backbone methylene carbons are sensitive to an even number of structural units with the simplest configurational sensitivity being dyad. The number of unique combinations for a particular sequence length is given by (27):

$$N_{(n)} = 2^{n-2} + 2^{(n-2)/2}$$

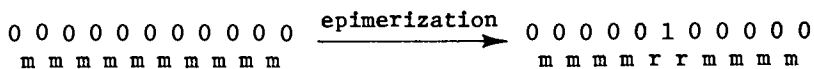
where

$$\begin{array}{ll} n = 2, & N = 2 \\ n = 4, & N = 6 \\ n = 6, & N = 20 \quad \text{etc.} \end{array}$$

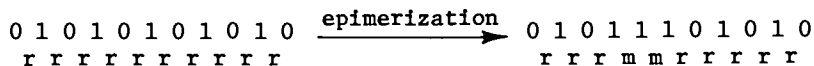
If the structural sensitivity exhibited by C-13 NMR extends beyond next-nearest neighbor configurations, an unwieldy number of configurations must be considered. Such has been the case for the aromatic quaternary carbon resonances in polystyrene where 20-22 resonances are observed as shown in Figure 3. When making these assignments, one must consider thirty-six possible heptads. Even "clearcut" analyses may be deceptively simple. The methyl spectrum of polypropylene apparently consists of three triad regions which are further subdivided into pentads. However, some of the pentads may be composites of overlapping heptads. This is particularly true for the resonances from the rr-centered sequences.

Thus only the assignment problem faced in C-13 NMR polymer configurational studies can be defined by simply counting the number of observed resonances. Fortunately, for most vinyl polymers, the configurational sensitivity is predominantly triad-pentad for the backbone methine and pendant side-chain carbons and dyad-tetrad for the backbone methylene carbons. Caution should be exercised when making assignments because even with the existence of only a few configurational resonances, the assignments are often neither straightforward nor unambiguous.

Let us now examine the techniques used in making configurational assignments in polypropylene. In principle, they could be applied in any configurational study. The polypropylene assignments have been well established and are probably the least equivocal of any reported for vinyl polymers. Two configurational assignments can be made without difficulty by comparing the amorphous polymer spectrum with those obtained from reference polymers consisting predominantly of isotactic and syndiotactic sequences. Roberts, et al., identified both the *mmmm* and *rrrr* pentads in the methyl region using this approach (24). Randall later correctly identified the configurational sequence of assignments through an extension of the Grant and Paul parameters to account for configurational differences (10). Final proof of the assignments, however, came from a model compound study by Zambelli, et al., (7) and an epimerization study by Stehling and Knox (5). Let's first consider the epimerization study because it can be more easily adapted to other polymer systems. An addition of 1% dicumylperoxide plus 4% tris(2,3-dibromopropyl)phosphate to a configurationally pure polypropylene results in well-spaced inversions if the conversions are deliberately kept low.^a



The new configurational sequences, *mmmr*, *mmrr* and *mrrm* will be produced from the isotactic polymer by isolated inversions of configuration. The relative intensities for the *mmmr*, *mmrr* and *mrrm* pentads will be 2:2:1 which allows a positive identification of the *mrrm* pentad. Correspondingly, the predominantly syndiotactic polymer will have *mm* triads inserted into *rr.....r* sequences by the inversion process.



Consequently, the new pentads, *mmrr*, *rrrm* and *rmmr* will be produced in a 2:2:1 ratio. Once again, the *rmmr* pentad can be identified by its unique relative intensity. The *mmrr* pentad is

^a A "0" represents one monomer unit configuration, "1" is its opposite configuration.

commonly produced between the two inversion experiments and therefore can be positively identified. The rrrm and mmmr pentads are assigned by default. The epimerization experiment, therefore, leads to an identification of all but the mrrr, mrrm and rrrr pentads. Most importantly, the resonances from common triad centers, that is, mm, mr and rr have been positively identified. This triad information is sufficient for a complete determination of the configurational structure.

The probable "order" of the configurational assignments in polypropylene have been established by Zambelli, et al., (7) and A. Provasoli and D. R. Ferro (11) in a study of C-13 labelled compounds, 3(S), 5(R), 7(RS), 9(RS), 11(RS), 13(R), 15(S)-heptamethylheptadecane (compound A) and a mixture of A with 3(S), 5(S), 7(RS), 9(RS), 11(RS), 13(R), 15(S)-heptamethylheptadecane. The pentad assignments from low to high field are mmmmm, mmmmr, rmmmr, mrrrr, mrrrm, rrrrr, mrrrr, mrrrm and mrrm in agreement with the eight assignments by Stehling and Knox and the order predicted earlier using the NMR additivity relationships. The polypropylene spectrum appears consistent with the assignment order observed in the model compounds; however, heptad splitting in the mr and rr-centered pentads could obscure the true situation and lead to misplaced pentad assignments. In spite of this difficulty, the triad distribution can be obtained with confidence. Quantitative results can be acquired by either integrating or curve fitting the methyl region for the mm, mr and rr relative intensities (28).

The most commonly used technique for making C-13 NMR spectral assignments for vinyl homopolymers has been through the use of Bernoullian statistics (29) (30). If one knows the relative concentrations of either m or r, any particular "n-ad" distribution can be calculated because

$$P_m = X_m \quad (3)$$

and

$$1 - P_m = X_r \quad (4)$$

Accordingly, the relative triad and tetrad concentrations are given by,

<u>Triad</u>		<u>Tetrad</u>	
mm	P_m^2	mmm	P_m^3
mr, rm	$2(1-P_m)P_m$	mrr, rmm	$2P_m^2(1-P_m)$
rr	$(1-P_m)^2$	rrr	$P_m(1-P_m)^2$
		mrrm	$P_m^2(1-P_m)$
		mrrr, rrrm	$2P_m(1-P_m)^2$
		rrrr	$(1-P_m)^3$

If the m and r mole fractions are unknown, one usually iterates over values for P_m until satisfactory agreement is obtained between the calculated and observed n -ad distributions. The success of the method depends upon the existence of discriminating differences among the relative intensities and a well-resolved triad, tetrad or " n -ad" distribution. It is generally best applied when more than one spectral region (for example, a triad methine distribution and a tetrad methylene distribution) are available for the calculated versus observed fit. Even under the best of circumstances, unique fits may not be obtained. If independent partial assignments are available (for example, the isotactic " n -ad" resonance) the method can be applied with more confidence. In cases where Bernoullian fits cannot be obtained, one then resorts to higher order statistical analyses, that is, first order Markov or Coleman-Fox (29). Care must be exercised, however, because more parameters are used to fit the data, thereby reducing the number of degrees of freedom in the analysis. In spite of these difficulties, more assignments have been proposed utilizing conformity to statistical behavior than any other technique. An example where good Bernoullian fits were obtained between the methine and methylene regions has been reported by Carman for polyvinylchloride (13).

Methine Resonances

<u>Triad</u>	<u>Observed</u>	<u>Calculated $P_m = 0.45$</u>
rr	0.291	0.303
mr	0.520	0.496
mm	0.188	0.202

Methylene Resonances

<u>Tetrad</u>	<u>Observed</u>	<u>Calculated $P_m = 0.45$</u>
rrr	0.161	0.166
rmr	0.146	0.130
rrm	0.282	0.272
mmr+mrn	0.320	0.334
mmm	0.092	0.091

Once assignments are made, C-13 NMR " n -ad" distributions are available. In general, one would like to obtain a distribution over the longest possible sequence length. Relationships, often referred to as the "necessary relationships", exist between n -ad sequences of different lengths. It is possible to reduce any n -ad distribution to m versus r , which correspond to the simplest comonomer distribution but is devoid of any information concerning sequence length.

More generally,

$$\bar{n}_{\text{like}} = \frac{\sum_{i=0}^{i=n} iN_{1(0)}_{i1} + \sum_{i=0}^{i=n} iN_{0(1)}_{i0}}{\sum_{i=1}^{i=n} N_{1(0)}_{i1} + \sum_{i=1}^{i=n} N_{0(1)}_{i0}} = \frac{1}{(r)} \quad (7)$$

As was true in the case of mean sequence lengths for meso and racemic dyads, the necessary relationships can be used to develop corresponding equations for any particular n-ad distribution. A favorable point concerning the concept of "like" configurations is that it attaches a physical significance to the racemic distribution.

The mean sequence lengths may offer a better way to present the simple comonomer distribution than meso, racemic distributions because they do reflect the polymer sequential structure. In addition, mean sequence lengths may be useful in evaluating statistical fits. For example,

$$\bar{n}_m = \bar{n}_r = \bar{n}_{\text{like}} = 2.0 \quad (8)$$

for ideally random Bernoullian distributions when $P_m = 0.5$. Generally, for Bernoullian distributions,

$$\bar{n}_m = \bar{n}_{\text{like}} = 1/(1-P_m) \quad (9)$$

and

$$\bar{n}_r = 1/P_m \quad (10)$$

For first order Markov,

$$\bar{n}_m = 1/(P_{m/r}) \quad (11)$$

$$\bar{n}_r = 1/(P_{r/m}) \quad (12)$$

$$\bar{n}_{\text{like}} = 1 + (P_{r/m})/(P_{m/r}) \quad (13)$$

Mean sequence lengths can also be used to write average configurational structures, that is,

$${}^{(m)}\bar{n}_m \quad {}^{(r)}\bar{n}_r$$

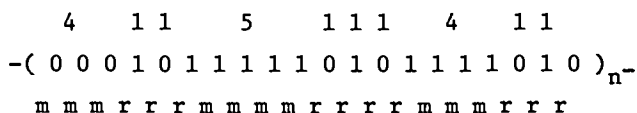
In the amorphous polypropylene, shown in Figure 1, the mean sequence lengths are:

$$\bar{n}_{\text{like}} = 2.0$$

$$\bar{n}_m = 3.3$$

$$\bar{n}_r = 3.4$$

The sequence distributions are clearly non-Bernoullian in this case although they could satisfy conformity to Markovian behavior as indicated by equations 11-13. The following average structure is suggested by these observed mean sequence lengths.



This amorphous polypropylene, therefore, has a tendency toward short blocks of meso and racemic sequences. This structural conclusion is not readily apparent from a simple inspection of the pentad distribution although the dominant pentads are mmmmm, mmmmr, rrrr, rrrm and mmrr as indicated by the above formula for the average configurational structure.

In conclusion, we can see that C-13 NMR offers possibly the best experimental approach now available to determine polymer tacticity or stereochemical sequence distributions. The chemical shift sensitivity is generally in the ideal range of dyad to pentad. Higher sensitivities could lead to n-ad distributions which are unwieldy or difficult to assign. From the work reported in the literature, we find that free radical catalysts generally produce configurational distributions which conform to Bernoullian statistics (30). Cationic or anionic catalysts or initiators produce distributions conforming to either Coleman-Fox or higher order Markovian models. Bernoullian statistics have proven to offer a reasonable approach to assignments for polymers produced by free radical initiators. The model compounds for polypropylene have apparently given the correct pentad methyl assignments and may offer a useful approach to assignments in other polymers. Epimerization is one of the more imaginative techniques but so far has seen only a limited application.

When describing polymer tacticity, one should attempt to obtain the highest complete "n-ad" distribution available as well as a simple "comonomer" distribution. In connection with such a measurement, the mean sequence lengths may offer a viable alternative to the simple m versus r distribution. Useful relationships, which are helpful in establishing particular statistical behaviors, are available.

I believe we are just on the threshold of the C-13 NMR applications in studies of polymer chemistry. Detailed structures are available for correlations with physical properties. Differences in configurational structure produced by various catalysts can be accurately determined. For example, the "irregularities" in crystalline polypropylene structure have been shown to be

```

0 0 0 0 0 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0
m m m m m m m m m m m r r m m m m m m m m m m

```

as opposed to

```

0 0 0 0 0 0 0 0 0 0 0 0 1 1 1 1 1 1 1 1 1 1 1 1
m m m m m m m m m m m r m m m m m m m m m m m m

```

Although not presently reported, the latter structure could be easily detected if it were, in fact, produced by an appropriate catalyst system.

Finally, the configurational structure in copolymers is currently a topic investigated by many industrial scientists. Is configuration retained when inserting ethylene into a sequence of isotactic propylene units? This question has been recently answered for a particular catalyst system where it was shown that configuration was retained and catalyst stereospecificity was considered to be the controlling factor (33). Now that the groundwork has been laid, there should be more extensive and varied applications of C-13 NMR in determining polymer structure. Certainly, an indispensable tool has become available for determining polymer stereochemical configuration.

REFERENCES

1. G. Natta and F. Danusso, J. Polym. Sci., XXXIV, 3 (1959).
2. F. A. Bovey, "Polymer Conformation and Configuration", Academic Press, New York, N. Y., 1969, p. 8.
3. F. A. Bovey and G. V. D. Tiers, J. Polymer Sci., 44, 173 (1960).
4. F. A. Bovey, "Polymer Conformation and Configuration", Academic Press, New York, N. Y., 1969, p. 32.
5. F. C. Stehling and J. R. Knox, Macromolecules, 8, 595 (1975).
6. D. M. Grant and E. G. Paul, J. Amer. Chem. Soc., 86, 2984 (1964).

7. A. Zambelli, P. Locatelli, G. Bajo, and F. A. Bovey, Macromolecules, 8, 687 (1975).
8. J. Schaefer and D. F. S. Natusch, Macromolecules, 5, 416 (1972).
9. D. E. Axelson, L. Mandelkern and G. C. Levy, Macromolecules, 10, 557 (1977).
10. J. C. Randall, J. Polym. Sci., Polym. Phys. Ed., 12, 703 (1974).
11. A. Provasoli and D. R. Ferro, Macromolecules, 10, 874 (1977).
12. J. C. Randall, J. Polym. Sci., Polym. Phys. Ed., 13, 889 (1975).
13. C. J. Carman, Macromolecules, 6, 725 (1973).
14. T. K. Wu and D. W. Ovenall, Macromolecules, 6, 582 (1973).
15. J. C. Randall, J. Polym. Sci., Polym. Phys. Ed., 14, 1693 (1976).
16. I. R. Peat and W. F. Reynolds, Tetrahedron Letters, No. 14, 1359 (1972).
17. K. F. Elgert, R. Wicke, B. Stützel and W. Ritter, Polymer, 16, 465 (1975).
18. K. Matsuzaki, H. Ito, T. Kawamura and T. Uryu, J. Polym. Sci., Polym. Chem. Ed., 11, 971 (1973).
19. J. C. Randall, "Polymer Sequence Determination: Carbon-13 NMR Method", Academic Press, New York, N. Y., 1977, Chapter 6.
20. L. F. Johnson, F. Heatley and F. A. Bovey, Macromolecules, 3, 175 (1970).
21. Y. Inoue, K. Koyama, R. Chûjô and A. Nishioka, Makromol. Chem. 175, 277 (1975).
22. H. Girad and P. Monjol, C. R. Hebd. Seances Acad. Sci., Ser. C., 279(13), 553 (1974).
23. H. Matsuzaki, T. Kanai, T. Kawamura, S. Matsumoto and T. Uryu, J. Polym. Sci., Polym. Chem. Ed., 11, 961 (1973).

24. W. D. Crain, Jr., A. Zambelli, and J. D. Roberts, Macromolecules, 3, 330 (1970).
25. Y. Inoue, A. Nishioka, and R. Chûjô, Makromol. Chem., 152, 15 (1972).
26. A. Zambelli, D. E. Dormann, A. I. Richard Brewster and F. A. Bovey, Macromolecules, 6, 925 (1973).
27. F. A. Bovey, "Polymer Conformation and Configuration", Academic Press, New York, N. Y., 1969, p. 16.
28. J. C. Randall, J. Polym. Sci., Polym. Phys. Ed., 14, 2083 (1976).
29. F. A. Bovey, "Polymer Conformation and Configuration", Academic Press, New York, N. Y., 1969, Chapter 2.
30. J. C. Randall, "Polymer Sequence Determination: Carbon-13 NMR Method", Academic Press, New York, N. Y., 1977, Chapter 4.
31. H. L. Frisch, C. L. Mallows, and F. A. Bovey, J. Chem. Phys. 45, 1565 (1966).
32. J. C. Randall, "Polymer Sequence Determination: Carbon-13 NMR Method", Academic Press, New York, N. Y., 1977, p. 35.
33. J. M. Sanders and R. A. Komoroski, Macromolecules, 10, 1214 (1977).

DISCUSSION

T. K. Wu, Du Pont de Nemours, Delaware:

I have a minor comment concerning the polyvinyl alcohol spectrum. The one shown (Figure 5) is similar to one obtained in our laboratory in 1973. Last year, Dr. Lana Sheer obtained a polyvinyl alcohol spectrum where the methine resonance was resolved into a triplet of triplets. By studying the effect of temperature upon the spectrum and by tuning the spectrometer carefully, one sometimes obtains better hyperfine structure.

J. C. Randall:

Have you obtained satisfactory assignments?

T. K. Wu:

Yes, the assignments have been published in Macromolecules (10, 529 (1977)).

J. C. Randall:

The spectra shown here were obtained from our equipment so, in principle, spectra could be presented which were recorded under similar operating conditions.

W. M. Pasika, Laurentian University, Ontario:

What dictates how far down the chain configurational effects are sensed by the resonance from your reference carbon?

J. C. Randall:

I believe the evidence points to conformational factors. Some people have recently observed chemical shift effects between carbons as far as six bonds away. Originally, Grant and Paul (reference 6) defined chemical shift "contributions" among five neighboring carbons, which were described as α through ϵ . I believe it is conformational differences which are responsible for the chemical shift behavior observed for various configurations. Depending upon the particular set of circumstances, these effects can involve five to seven consecutive monomer units. Sometimes, these chemical shift differences can be treated by an additive scheme which considers the contributions from each possible configurational array.

W. M. Pasika:

Do the observed differences in configurational chemical shifts disappear at higher temperatures?

J. C. Randall:

There is a temperature effect in many systems. The peaks either collapse or separate. Again, this goes back to the concept that it is the conformation of the configuration that you are dealing with. This is why we see so much unusual chemical shift behavior when we examine trends among vinyl polymers.

G. Babbitt, Allied Chemical, New Jersey:

With regard to branching in polyethylene, I was wondering if you have arranged your experiments in such a way that NOE's and T_1 's are taken into account so you can assume

that your areas represent accurate quantitative data. In studies of polyethylene branching, there is a strong major methylene resonance with much weaker methylene and methyl resonances. What is the best way to handle such data quantitatively as far as the area measurements are concerned?

J. C. Randall:

In the systems that I have examined, I can satisfy the dynamic requirements with a ten second pulse delay. The longest methyl T_1 may be 3 seconds. In general, the longer the side chain, the longer will be the methyl T_1 . We will hear more about this subject later on. We need not be too concerned about NOE factors because they are usually full under the experimental conditions ($T = 120-130^\circ\text{C}$) used for polymer quantitative measurements. The T_1 problem can be handled, even under non-equilibrium conditions, by utilizing resonances from the same types of carbon atoms in a quantitative treatment. Such an approach can sometimes lead to more efficient quantitative NMR measurements. Adequate pulse spacings will have to be used whenever one wishes to utilize all of the observed resonances. Quantitative measurements in branched polyethylenes are very desirable because this is one of the best applications of analytical polymer C-13 NMR.

G. Babbitt:

I was wondering more about the physical problem of obtaining areas, for example, cutting and weighing, when some of the methylene resonances may be truncated.

J. C. Randall:

The problems associated with dynamic range are ones which we would all like to see solved. I am using peak heights with some success. It may be the best method when the relative heights are 500:1 or higher. In any event, one would like to have an independent measurement or reference standard. I have checked with infrared analyses because it can measure the methyl content. Unfortunately, IR does not discriminate the polymer methyl end group from methyls on side-chain short branches. Nevertheless, with this consideration, I have obtained good correlations between IR and NMR measurements using NMR peak heights. Such a comparison can often serve as a guideline when measuring an internal branching distribution. More confidence is placed in the NMR result, if, overall, similar results are achieved. Truncation, which occurs among resonances with relatively low intensities, is a constant worry. Another way of checking for possible truncation in a specific ratio range is through number average molecular weight measurements on

NBS standard polyethylenes where the M_n is known. Commercial polyethylenes may also be useful because the M_n is frequently in the 10 to 15 thousand range which corresponds to the sensitivity available with most C-13 NMR spectrometers.

D. E. Axelson, Florida State University, Florida:

We have been investigating branched polyethylenes in solution and would like to respond to a couple of questions which were brought up. In a recent paper in Macromolecules (10, 557 (1977)), we showed that some methyl groups, particularly butyl and longer, have T_1 's as long as 7 seconds; hence 5 T_1 could be of the order of 35 seconds at 120°C in solution. We have also measured the NOE's of most of the small branches because of the high sensitivity of our 270 MHz spectrometer, and, as you indicated, the NOE enhancements have uniformly been full. We have been able to measure peak areas because of our good sensitivity. Electronic integrations have not given satisfactory results. More consistent results have been obtained with a planimeter. Dr. Cudby of ICI in England has been very kind to send us some comparisons between NMR and IR total methyl contents. Close agreement was obtained when the two experiments were done carefully.

J. C. Randall:

Have you had an opportunity to evaluate the results from peak heights versus peak area measurements?

D. E. Axelson:

The peak heights worked out fairly well. I do not believe you get into a peak height versus peak area problem if the resolution is sufficiently high and overlap is not a problem. We found that results from one spectrum to another were within an order of one to two branches per thousand carbons as far as consistency was concerned. Spectra, which were obtained on other systems under less than equilibrium conditions, show that the total methyl content, surprisingly, stays very constant. You find that the low end is enhanced and the long branches are saturated slightly; however, the total remains the same.

J. C. Randall:

Yes, I believe more efficient NMR methods can be developed by working on systems with known amounts of branching.

D. E. Axelson:

A comparison to IR results can be dangerous in the absence of a good standard for the IR experiment. We ran into a problem where the IR results were one-half of that obtained from NMR. This discrepancy can usually be attributed to the IR method.

J. C. Randall:

Yes, since IR measures total methyl groups, problems can develop if the branching content is comparable to the end group concentration.

RECEIVED March 13, 1979.

INDEX

A	
Absolute signal intensities	252 <i>t</i>
Agaro-oligosaccharides, C-13 NMR spectra of	131 <i>f</i>
Agarose	130-132
C-13 NMR spectra of	131 <i>f</i>
degradation of	130
Alkali-metal compounds, allyl	90
Allyl	
alkali-metal compounds	90
carbons in I-M and II-M chemical shifts of	93 <i>t</i>
compounds, deuterated	90
compounds, undeuterated	90
M C-13 chemical shifts at 0°C	91 <i>t</i>
Allylic positions, calculated charges on	95 <i>t</i>
Alternating copolymers, C-13 NMR analysis in characterization of	215-233
Amorphous PP	311
Amylose, C-13 NMR spectra of	41 <i>f</i>
Amylose, synthetically branched	38-40
Anionic polymerization systems, C-13 NMR studies of	89-95
Anisotropic rotation in dilute solution, comparison of substituent group	281 <i>t</i>
Anisotropy, chemical shift	68, 69
Anisotropy, conformational	80
Anomeric	
carbons, chemical shifts of	123
region of the C-13 NMR spectra	39 <i>f</i>
resonances, analysis of	46
Aromatic quaternary resonances from a free-radical polystyrene	296 <i>f</i>
B	
Backbone	
dynamics in dilute solution, comparison of	280 <i>t</i>
dynamics and substituent group rotation, comparison between	282 <i>t</i>
methylene resonances for poly(vinyl ethers)	299
motions in poly (but-1-ene sulfone), correlation times for	23 <i>t</i>
rearrangements	281-284
in solids, comparison of	283 <i>t</i>
Benzene, proton relaxation of	153
Bernoullian	
behavior	293
distributions	310
statistics	307-308
Bound species, correlation times of	174-177
Branched PE, spin-lattice relaxation time vs. temperature for	193 <i>f</i>
Branched polymers, comparison of spectra of linear and	53
Branching and the polymerization mechanism	64-65
2-Bromo-4,6-dichlorophenoxide, C-13 NMR spectra of polymers derived from	56 <i>f</i>
4-bromo-2,6-dichlorophenoxide and	59 <i>f</i>
2-Bromo-4,6-dichlorophenoxide reaction	63
4-Bromo-2,6-dichlorophenoxide	
C-13 NMR spectra of copolymers derived from 2-bromo-4,6-dichlorophenoxide and	59 <i>f</i>
C-13 NMR spectra of copolymers derived from 2,4,6-trichlorophenoxide and	59 <i>f</i>
polymerization of	57-58
<i>cis</i> -2-Butene-ethylene alternating erythrodiisotactic copolymer, C-13 chemical shifts of	227 <i>t</i>
Butyl rubber as a solid, C-13 NMR spectrum of	114 <i>f</i>
C	
Carbohydrate polymers by C-13 NMR spectroscopy, characterization of	123-138
C-C coupling constants	90
C-1 signal of methyl β -cellobioside- <i>d</i> ₅	137 <i>f</i>
C-13-C-13 NMR coupling constants ..	91
C-13 { ¹ H} NMR of <i>cis</i> -1,4-polyisoprene	110 <i>f</i>
C-13 { ¹ H} NMR of propylene butadiene copolymer	108 <i>f</i>

[2-[2-C-13]glycine] methionine	
enkephalin	160, 172f, 173f
C-13 chemical shift of the glycol	
residues in	164f
correlation times of	170f
K _a vs. pH for	175f
linewidths of	166t
to other lipids, binding of	178
pH-dependence of T ₁ of	167f, 168f
C-13 NMR	1-23, 181
of agaro-oligosaccharides	131f
of agarose	131f
of alternating copolymers	215-233
of amylose	41f
of 2-bromo-4,6-dichlorophenoxide	
polymers	56f
4-bromo-2,6-dichlorophenoxide	
polymers	56f
of butyl rubber as a solid	114f
calibration curve	253f
of carbohydrate polymers	123-138
of carbon-black-loaded polyiso-	
prene, FT	110f
chemical shifts	
at 0°C, allyl M	91t
of allyl carbons in I-M and II-M	93t
anisotropy	68, 69
of anomeric carbons	123
of <i>cis</i> -2-butene-ethylene alternat-	
ing erythrodiisotactic co-	
polymer	227t
of C-β of tyrosine in enkephalin	162f
characteristic	123-124
correlations	127-130
dependence on temperature	
(Δδ/ΔT)	28
for ethylene propylene rubbers	100t
gamma carbon	92-95
of the glycol residues in 2-[2-C-	
13]glycine methionine enke-	
phalin	164f
of the glycol-2 residues of methio-	
nine enkephalin	165f
of glycol-3 residues of methio-	
nine enkephalin	165f
for hydrochlorinated 1,4-polydi-	
methylbutadiene, observed	
and predicted	232t
of hydrogenated 1,4-polydi-	
methylbutadiene	227t
for hydrogenated 1,4-polyiso-	
prene, observed and	
predicted	224t
of oxonium ions	260f
carbon	258
resonances in PS sulfones	3-8
in styrene-SO ₂ copolymers, main	
chain carbon	8t
C-13 NMR (<i>continued</i>)	
chemical shifts (<i>continued</i>)	
tensor	67
of tetramethylene ethers	251t
upfield movement of	92
of chloroprene-SO ₂ copolymer	15f
of chondroitins A, B, and C	129f
of copolymers	107
derived from chlorophenoxides	59f
to copolymerizations, application	
of	258-263
coupling constants, C-13-	91
20-crown-4 in CDCl ₃	250f
of crystalline PP	304f
of a cured epoxy, three stages of	
resolution in	71f
for determination of polymer	
tacticity	291
of dextran(s)	29-38
B-742	48f
B-1142	39f
B-1299	31f, 33f
B-1396	31f
B-1402	39f
B-1501	35f
B-1526	39f
of elastomers	97-117
of EPDM rubbers	98f, 104t
Fourier transform	28
- ¹ H coupling in oligosaccharide	
models, stereochemistry of	135-138
of heparin A	126f
of heparin B	126f
of hyaluronic acid	125f
of hydrogenated 1,4-polydimethyl-	
butadiene	225f, 231f
of hydrogenated 1,4-polyiso-	
prene	223f, 228f
interpretation of	3
for levan B-133	31f
magnetization	74
of the main chain of styrene-SO ₂	
copolymers	6f
mannan Y-1842	39f
mannan YB-1344	39f
of methyl β-cellobioside	125f
of methyl α-D-glucopyranoside	41f
molecular weight effects on	53
n-ad distributions	308
of neoagaro-oligosaccharides	131f
nuclei, spin-lattice relaxation of	80
in organic solids	67
of the oxonium ion region	243f
of PAN	300f
of PEMA solutions	145f
of polyalkane	109f
of poly(dichlorophenyleneoxides)	53
of polydienes	222-233

- C-13 NMR (*continued*)
- and polymer stereochemical configuration 291-312
 - of polymer from 2,4,6-trichlorophenoxide 61*f*
 - of PP 294*f*, 303
 - polysaccharide branching and 27-50
 - of PS 293, 295*f*
 - of a polytetrahydrofuran 239
 - of PTMEG in THF 240*f*
 - of PVC 297*f*, 298*f*
 - quantitative applications of 249-258
 - of rabbit liver glycogen 41*f*
 - relaxation 143
 - behavior of chloroform as a function of polymer concentration 144
 - behavior of chloroform as a function of temperature 144
 - concentration dependence of methyl 150*f*
 - concentration dependence of methylene 149*f*
 - concentration dependence of quaternary 148*f*
 - as a function of temperature, chloroform 146*f*
 - measurements 21
 - and polymer concentration, relationship between solvent temperature of methyl 150*f*
 - temperature of methylene 149*f*
 - temperature of quaternary 148*f*
 - of SBR/polybutadiene rubber blend 113*f*, 114*f*
 - sensitivity of 292
 - spectroscopy, quantitative 252*t*
 - spin relaxation
 - parameters of linear PE 195*t*
 - parameters of semicrystalline polymers 181-209
 - polymers 271-286
 - studies
 - of anionic polymerization systems 89-95
 - on cationic ring-opening polymerization of cyclic ethers 237-267
 - of hormone binding to receptor membranes 159-179
 - of styrene-SO₂ copolymers 7*f*
 - of THF-CH₃OSO₂F 242*f*
 - polymerization mixture in CH₃NO₂ 247*f*
 - of THF-methyltriflate polymerization mixtures 241
 - of a THF-OMP-CF₃SO₃CH₃ polymerization mixture of CH₃NO₂ 262*f*
- C-13 NMR (*continued*)
- of 2,4,6-trichlorophenoxide polymers 56*f*
 - two-line 90
 - of vinyl polymers 293
 - α Carbon chemical shifts of oxonium ions 258
 - α Carbon resonance region of a THF/OMP copolymerization in CH₃NO₂ 258
 - Carbon(s)
 - black-filled polyisoprene, cross polarization spectra of 112*f*
 - black-loaded polyisoprene 110*f*
 - olefinic 16
 - protonated and unprotonated 82*f*
 - species as a function of a sequence, number of 102*t*
 - Carr-Purcell pulse sequence 68
 - Catalyst system, Ziegler 105
 - Catalysts, Ziegler-Natta 222
 - Cationic polymerization of cyclic ethers, chain transfer during 245
 - Cationic ring-opening polymerization of cyclic ethers, C-13 NMR studies on 237-267
 - CH₃NO₂, C-13 NMR spectrum of a THF-CH₃OSO₂F polymerization mixture in 247*f*
 - Chain
 - dynamics of polysulfones 1-23
 - structures within a semicrystalline polymer, regions of 181
 - transfer during cationic polymerization of cyclic ethers 245
 - Characteristic chemical shifts 123-124
 - Chemical change, monitoring 132
 - Chloroform
 - C-13 relaxation as a function of temperature 146*f*
 - as a function of polymer concentration, C-13 relaxation behavior of 144
 - as a function of temperature, C-13 relaxation behavior of 144
 - ¹H relaxation, concentration dependence of 152*f*
 - ¹H relaxation, temperature of 152*f*
 - nuclear Overhauser enhancement factor as a function of temperature 147*f*
 - rotation diffusion coefficients 155
 - Chloroprene
 - centered sequences 12
 - SO₂ copolymers 11
 - C-13 spectrum of 15*f*
 - preparation of 13*t*
 - Chondroitin(s)
 - A, B, and C, C-13 spectra of 129*f*

CLC-MS, permethylation fragmen- tations	27	Copolymer(s) (<i>continued</i>)	
Cole-Cole distribution	155	methylene carbons in	17f
Compositional tetrads in chains of vinyl (<i>M</i>) copolymers	5f	olefin carbons in	18f
Compositional triads in chains of vinyl (<i>M</i>) copolymers	4f	preparation of chloroprene-SO ₂	13t
Computer fit of a C-13 NMR spectrum using conditional probabilities	103t	styrene-SO ₂	2
Configuration(s)		Copolymerization(s)	261
advantages of C-13 NMR in meas- urements of polymer stereo- chemical	292	application of C-13 NMR to	258-263
C-13 NMR polymer stereo- chemical	291-312	in CH ₃ NO ₂ , α carbon resonance region of a THF/OXP	258
dyad	291	of THF/OXP at 20°C	263t
mean sequence lengths for meso ...	309	Correlation	
mean sequence lengths for racemic	309	function	277
measurement of polymer	291	times	
meso dyad	291	of bound species	174-177
pentad	303, 305	of [2-[2-C-13]glycine]methio- nine enkephalin	170f
racemic dyad	291	exponential	272-273
syndiotactic	291	weighted harmonic average (τ_h^*) ..	277
tactic	291	Counter ion on the % 1,2 structure in polybutadiene, effect of	95t
Configurational		Coupling constants	
assignments in PP, techniques for making	306	at 0°C	91t
dyads in chains of vinyl (<i>M</i>) copolymers	4f	C-C	90
sensitivity		C-13-C-13 NMR	91
methyl	303	of the enkephalins in solution, ¹ H spin-spin	159
triad	303	Coupling, proton-carbon dipolar	69, 81f
vinyl polymers and	293	Cross-polarization	70
triads in chains of vinyl (<i>M</i>) copolymers	5f	magnetization for the PIP-cured epoxy under the SL condition ..	76f
Conformational anisotropy	80	process	74
Copolymer(s)		influence of magic angle spinning on	75
C-13 NMR analysis in characteriza- tion of alternating	215-233	rate	83
C-13 spectrum of		spectra of carbon-black-filled polyisoprene	112f
chloroprene-SO ₂	15f	spin lock	70
the main chain of styrene-SO ₂ ...	6f	time constant, influence of magic angle spinning on the SL	77f
styrene-SO ₂	7f	time constant, rf pulse sequences for determining SL	79f
chloroprene-SO ₂	11	without magic angle spinning	111
derived from 2-bromo-4,6-dichloro- phenoxide and 4-bromo-2,6-di- chlorophenoxide, C-13 NMR spectra of	59f	Cross-relaxation, dipolar	70
derived from 2,4,6-trichlorophen- oxide and 4-bromo-2,6-dichlo- rophenoxide, C-13 NMR spectra of	59f	Crown ether nomenclature	246
ethylene-butene-1	201	Crystalline PP, C-13 NMR spectrum of	304f
main chain carbon chemical shifts in styrene-SO ₂	8t	Crystalline regions on segmental motions, influence of	200
		Crystallinity, degree of	185
		Crystallinity, homopolymers of low levels of	201
		Cured epoxy(ies)	
		molecular motion in	70-80
		spectral fidelity in	70-80
		spectral resolution in	70-80
		three stages of resolution in a C-13 spectrum of	71f

- Cyclic ethers
 C-13 NMR studies on the cationic ring-opening polymerization of 237-267
 chain transfer during cationic polymerization of 245
 mechanism of polymerization of 238-239
 polymerization of 238-249
 polymerization by Fourier transform NMR, kinetic analysis of 264-267
 polymerization, kinetics of a reversible 254-258
 spectra of polymerization of 238-239
 Cyclic oligomers, formation of 246-249
- D**
- Decoupling
 experiments, off-resonance 233
 gated 249
 "magic angle" for dipolar 69
 two-level 182-183
 Defect-diffusion model 155
 Delta effect, oxygen 19
 Density, spectral 271-276
 Density vs. logarithm of frequency, logarithm of spectral 279f
 Deuterated allyl compound 90
 Dextran(s) 28, 29-38
 B-742 34, 47
 C-13 NMR spectrum of 48f
 B-1142, anomeric region of the C-13 NMR spectra for 39f
 B-1299 32, 46-47, 49
 C-13 NMR spectra for 31f, 33f
 B-1396, C-13 NMR spectra for 31f
 B-1402, anomeric region of the C-13 NMR spectra for 39f
 B-1501, C-13 NMR spectra for 35f
 B-1526, anomeric region of the C-13 NMR spectra for 39f
 branching through 2,6-di-O-substituted α -D-glucopyranosyl residues 32
 C-13 NMR spectral analysis of 29-38
 series structures 32-38
 DGEBA (*see* Diglycidyl ether of bisphenol A)
 Diastereoisomerism, erythro 219-233
 Diastereoisomerism, threo 219-233
 Dielectric relaxation, analysis of 273
 Diglycidyl ether of bisphenol A (DGEBA) 70
 -based epoxies, composition for 72t
 -based epoxies, cure cycle for 72t
- Dilute polymer solutions, NMR spectroscopy of 143
 Dioxane, selective irradiation of 203f
 Dipolar
 cross-relaxation 70
 coupling between spin species, heteronuclear 78
 coupling, proton-carbon 69, 81f
 decoupling 69
 "magic angle" for 69
 Dissociation constants 169
 2,6-Di-O-substituted α -D-glucopyranosyl residues, dextrans branching through 32
 Dormant ion 245-246
 Dyad configurations 291
 meso 291
 racemic 291
 Dyad-triad relationships 309
- E**
- Elastomers
 C-13 NMR high-resolution characterization of 97-117
 high-resolution solution spectra of 97-107
 identification of 97
 Endo-cyclic methylene carbons 241
 Enkephalin
 C-13 chemical shift of C- β of tyrosine in 162f
 morphine antagonism of 178
 opioid peptide 159-179
 -PS interaction 169
 in solution, ^1H spin-spin coupling constants of 159
 Enthalpy of polymerization 254
 of THF 257t
 Entropy of polymerization 254
 of THF 257t
 Enzymes, polymer-degrading 130
 Enzymic applications 130-132
 EPDM A (*see* Ethylene propylene rubber)
 Epimerization 306
 Epoxy(ies) 73-78
 composition for DGEBA-based 72t
 cure cycle for DGEBA-based 72t
 solid-state spectra of 73f
 spectra of the PIP-cured 81f
 three stages of resolution in a C-13 spectrum of a cured 71f
 under the SL condition, cross-polarization magnetization for the PIP-cured 76f

- High-resolution
 characterization of elastomers,
 C-13 NMR 97-117
 solution spectra of elastomers 97-107
 spectra of solids 107-115
 of styrene butadiene rubbers 111
 Hole burning experiment 202
 Homogeneous
 line broadening 78
 linewidth, natural 202, 205
 resonance lines 205
 Homopolymer, vinyl 309
 Homopolymers of low levels of
 crystallinity 201
 Hormones to receptor membranes,
 C-13 NMR to study binding
 of 159-179
 Hyaluronic acid, C-13 NMR
 spectrum of 125*f*
 Hyaluronic acid, ¹H-NMR spectrum
 of 125*f*
 Hydrobrominated 1,4-polyisoprene 226-230
 Hydrochlorinated 1,4-polydimethyl-
 butadiene 230-233
 microstructure of 233
 ¹H NMR spectrum of 221*f*
 observed C-13 chemical shifts for .. 232*t*
 predicted C-13 chemical shifts for .. 232*t*
 proton noise-decoupled C-13 NMR
 spectrum of 231*f*
 Hydrochlorinated 1,4-polyisoprene 226-230
 Hydrogen halide addition to 1,4-poly-
 isoprene, Markownikoff's rule for 226
 Hydrogenated 1,4-polydimethyl-
 butadiene 222-226
 C-13 chemical shifts of 227*t*
 proton noise-decoupled C-13 NMR
 spectrum of 225*f*
 Hydrogenated 1,4-polyisoprene 222
 observed C-13 chemical shifts for .. 224*t*
 predicted C-13 chemical shifts for .. 224*t*
 proton noise-decoupled C-13 NMR
 spectrum of 223*f*
 Hydrogenation reactions of poly-
 dienes 216-219
 Hydrohalogenated 1,4-polyisoprenes,
 ¹H NMR spectra of 220*f*
 Hydrohalogenated 1,4-polyisoprene,
 proton noise-decoupled C-13
 NMR spectrum of 228*f*
 Hydrohalogenation reactions of
 polydienes 216-221
- I**
- Incomplete motional narrowing 205-206
 Inhomogeneity in the static magnetic
 field, line broadening owing to 200-201
- Inhomogeneous broadening 202
 Inhomogeneous line broadening 78
 Insertion, primary 99
 Insertion, secondary 99
 Internal rotation of substituents 281-284
 Interpretational models of C-13 NMR
 spin relaxation in polymers
 survey of 271-273
 Inverse first moment, weighted 277
 Ionic strength on the interaction of
 opiates with PS, effect of 177
 Isomerism, head-tail head-to-head 19
 Isotropic tumbling 272
- J**
- Jones and Stockmayer model 275-284
- K**
- Kinetic analysis of an equilibrium
 polymerization 266*f*
 Kinetic constants, determination of 259*f*
- L**
- Lattice models 273-278
 and rubbery polymers 283
 and solid amorphous polymers 283
 Levan B-133, C-13 NMR spectra for 31*f*
 Levans and β -D-fructofuranosyl
 compounds 44-49
 Lewis-Mayo copolymerization
 equation 2
- Line broadening
 component, residual zero frequency 205
 due to inhomogeneity in the static
 magnetic field 200-201
 homogeneous 78
 inhomogeneous 78
- Linear
 and branched polymers, comparison
 of spectra of 53
 poly(2,6-dichloro-1,4-phenylene-
 oxide) 57
 polyethylene(s)
 C-13 spin relaxation parameters
 of 195*t*
 linewidth vs. temperature for 186*f*-188*f*
 selective irradiation of 203*f*, 204*f*
 spin-lattice relaxation time vs.
 temperature for 193*f*
- Linewidth(s) 198-208
 chain entanglements influencing 206
 factors contributing to 200
 natural homogeneous 202, 205
 PE 207
 for PEO, resonant 201
 for PE, resonant 201

Linewidth(s) (<i>continued</i>)	
of <i>cis</i> -polyisoprene	206
of semicrystalline polymers	184-190
-temperature relation of the PEO ..	185
-temperature relations, influence of morphology on	185
vs. temperature for ethylene-butene-1 random co- polymers	191f
linear PE	186f-188f
PEO	189f
Living polymerization	238
Long pulse delay	249
Low-molecular-weight PEO with high-molecular-weight PEO, comparison of	196-198
M	
Macrocyclic ethers	246
Macroion, propagation \rightleftharpoons depropaga- tion equilibria of	239
Macromolecules, C-13 NMR studies on	123
Magic angle for dipolar decoupling	69
Magic angle spinning	67-69
of carbon-black-loaded polyiso- prene	110f
on the cross-polarization process, influence of	75
cross polarization without	111
on the SL cross-polarization time constant, influence of	77f
Magnetization, C-13	74
Magnetization for the PIP-cured epoxy under the SL condition, cross-polarization	76f
Main chain carbon chemical shifts in styrene-SO ₂ copolymers	8t
Mannan(s)	
extracellular	40-44
YB-1344	42
anomeric region of C-13 NMR spectra for	39f
Y-1842	42-44
anomeric region of C-13 NMR spectra for	39f
Mark-Houwink plot for polymers obtained from trichlorophenoxide ..	55f
Markov distributions, first-order	310
Markownikoff's rule for hydrogen halide addition to 1,4-polyiso- prene	226
Mean sequence lengths	311
for meso configurations	309
for racemic configurations	309
Meso dyad configuration	291
Meso configurations, mean sequence lengths for	309
Methine	
carbon relaxation in dissolved PS, comparisons of interpretations of	276t
carbon resonances, triad chemical for vinyl polymer backbone	301t
regions of the C-13 NMR spectrum of PS	295f
resonances	299
polymers with singlet backbone ..	301t
Methionine enkephalin, C-13 chemical shifts of the glycyL-2 and glycyL-3 residues of	165f
Methionine enkephalin, rotational correlation times of the glycyL residues in	169
Methyl	
C-13 relaxation, concentration dependence of	150f
C-13 relaxation, temperature of	150f
β -cellobioside, C-13 spectra of	125f
β -cellobioside- <i>d</i> ₈ , C-1 signal of	137f
configurational sensitivity	303
α -D-glucopyranoside, C-13 NMR spectra of	41f
β -Maltoside	135
Methylene	
C-13 relaxation, concentration dependence of	149f
C-13 relaxation, temperature of	149f
carbon(s)	
in copolymers	17f
endo-cyclic	241
resonances, tetrad chemical shift sequence for vinyl polymer backbone	302t
region, sequence assignments in	19
regions of C-13 NMR spectra of PP ..	294f
resonances	308
for poly(vinyl ethers), backbone ..	299
sequence(s)	99
of different lengths	101t
distribution of EPDM rubbers	101, 104t
length in ethylene propylene rubber, measurement of	105
Modified spectral density	274
Molecular	
motion in cured epoxies	70-80
weight effects on C-13 spectra	53
weight effects on ¹ H spectra	53
Monomer sequences, assignments of resonances to	3
Morphine antagonism of enkephalin ..	178
Motional narrowing, incomplete	205-206
Motional narrowing, line broadening caused by partial	206
Mucopolysaccharides	124-127

- N**
- n-ad distribution 307-308
 C-13 NMR 308
 Natural homogeneous linewidth 202, 205
 Neogaro-oligosaccharides, C-13
 NMR spectra of 131f
 Nitromethane at 0°C, polymerization
 of THF in 257t
 NMR spectroscopy of dilute polymer
 solutions 143
 NOE (*see* Nuclear Overhauser
 enhancement)
 NOEF (*see* Nuclear Overhauser
 enhancement factor)
 Noise-decoupled C-13 NMR
 spectrum of
 hydrochlorinated 1,4-polydimethyl-
 butadiene, proton 231f
 hydrogenated 1,4-polydimethyl-
 butadiene, proton 225f
 hydrogenated 1,4-polyisoprene,
 proton 223f
 hydrohalogenated 1,4-polyisoprene,
 proton 228f
 polytetrahydrofuran, proton 239
 Noncrystalline regions 196
 Nuclear Overhauser enhancement
 (NOE) 21, 249, 292
 factor (NOEF) 144, 183, 194-196, 249
 as a function of temperature,
 chloroform 147f
 Number fraction of M sequences 8
 Number fraction of SMS sequences .. 9
 plot of 10f
- O**
- Off-resonance decoupling experiments 233
 Olefin carbon in copolymers 18f
 Olefin-SO₂ polymerizations 1
 Olefinic carbons 16
 Oligomers, formation of cyclic 246-249
 Oligosaccharide models, stereochem-
 istry of C-13-¹H coupling in 135-138
 Opiates with PS, effect of ionic
 strength on the interaction of 177
 Opioid peptide enkephalin 159-179
 Organic solids, C-13 NMR in 67
 Overhauser enhancement, nuclear
 (NOE) 21
 factor (NOEF) 183, 194-196
 Oxepane (OXP) 237
 Oxonium ion(s) 245-246
 C-13 NMR chemical shift assign-
 ments of 260f
 α carbon chemical shifts of 258
- Oxonium ion(s) (*continued*)
 propagation ⇌ depropagation
 equilibria of 239
 region, C-13 NMR spectra of 243f
 OXP (oxepane) 237
 Oxygen delta effect 19
- P**
- Paramagnetic additives on the ¹H
 spectrum of poly(dichlorophenyl-
 eneoxide), effects of 58
 Partial motional narrowing, line
 broadening caused by 206
 PE (*see* Polyethylene)
 PEMA-CHCl₃ (*see* Poly(ethyl meth-
 acrylate)-chloroform)
 PEO (*see* Polyethylene oxide)
 Pentad assignments, heptad splitting
 in 307
 Pentad configurations 303, 305
 Peptide-lipid complex (PL) 169
 Permethylation fragmentation
 CLC-MS 27
 Phenoxy end-capping 239
 Phosphatidyl serine 159
 PIP-cured epoxy under the SL condi-
 tion, cross-polarization magneti-
 zation for 76f
 PIP-cured epoxy, spectra of 81f
 PL (peptide-lipid complex) 169
 Polyacrylonitrile (PAN), C-13 NMR
 spectrum of 300f
 Polyalkane, pulsed (FT) C-13 NMR
 spectrum of 109f
 Polybutadiene, effect of counter ion
 on the % 1,2 structure in 95t
 cis-Polybutadiene, interpretation of 283-284
 Poly(but-1-ene sulfone), correlation
 times for backbone and side-
 chain motions in 23t
 Poly(dichlorophenyleneoxide), effects
 of temperature and paramagnetic
 additives on the ¹H spectrum of .. 58
 Poly(dichlorophenyleneoxide),
 structure of 58-62
 using C-13 NMR spectroscopy 53
 Poly(2,6-dichloro-1,4-phenylene-
 oxide), linear 57
 Polydienes
 C-13 NMR characterization of 222-233
¹H NMR characterization of 216-221
 hydrogenation reactions of 216-219
 hydrohalogenation reactions of 216-221
 preparation of 216-221
 4-Polydienes, chemical modifications
 of 215-233
 Polydimethylbutadiene 216

1,4-Polydimethylbutadiene, hydrochlorinated	230-233	<i>cis</i> -Polyisoprene, linewidths of	206
¹ H NMR spectrum of	221f	1,4-Polyisoprene	
microstructure of	233	¹ H NMR spectra of the hydro-	
observed C-13 chemical shifts for ..	232t	halogenated	220f
proton noise-decoupled C-13 NMR		hydrobrominated	226-230
spectrum of	231f	hydrochlorinated	226-230
predicted C-13 chemical shifts for ..	232t	hydrogenated	222
1,4-Polydimethylbutadiene,		observed and predicted C-13	
hydrogenated	222-226	chemical shifts for	224t
C-13 chemical shifts of	227t	proton noise-decoupled C-13	
proton noise-decoupled C-13 NMR		NMR spectrum of	223f
spectrum of	225f	Markownikoff's rule for hydrogen	
1,4-Polydimethylbutadiene, upfield		halide addition to	226
region of ¹ H NMR spectra of	218f	proton noise-decoupled C-13 NMR	
Polydimethyl siloxane	207	spectra of hydrohalogenated ..	228f
Polyethylene(s) (PE)	185	upfield region of ¹ H NMR spectra	
C-13 spin relaxation parameters		of	217f
of linear	195t	<i>cis</i> -1,4-Polyisoprene, C-13 (¹ H) NMR	
linewidths	207	spectra of	110f
vs. temperature for linear	186f, 188f	Polymer(s)	
melts, proton spin-spin relaxation		C-13 NMR spin relaxation in	271-286
decay of unfractionated	208	C-13 spin relaxation parameters of	
oxide (PEO)	185	semicrystalline	181-209
comparison of low-molecular-		characterization	67
weight PEO with high-		comparison of spectra of linear and	
molecular-weight	196-198	branched	53
linewidth-temperature relation		concentration, relationship between	
of	185	solvent C-13 relaxation and	153
linewidth vs. temperature for	189f	configuration, measurement of	291
resonant linewidths for	201	-degrading enzymes	130
spin-lattice relaxation time vs.		derived from chlorophenoxides	56f
temperature for	192f	derived from trichlorophenoxide,	
resonant linewidths for	201	¹ H NMR spectra of	54f
selective irradiation of linear ..	203f, 204f	differentiation of	124-127
spin-lattice relaxation time vs		dynamics, comparison of	278-284
temperature for branched	193f	lattice models and rubbery	283
spin-linear relaxation time vs.		lattice models and solid amorphous	283
temperature for linear	193f	linewidths of semicrystalline	184-190
Poly(ethyl methacrylate)-chloroform		obtained from trichlorophenoxide,	
(PEMA-CHCl ₃)	143	Mark-Houwink plot for	55f
system by carbon-13 NMR, relaxa-		segmental motion	153
tion studies in	143-156	with singlet backbone methine	
system by proton NMR, relaxation		resonances	301t
studies in	143-156	solutions, NMR spectroscopy of	
Poly(ethyl methacrylate) (PEMA)		dilute	143
solutions, C-13 spectrum of	145f	stereochemical configuration, C-13	
Poly(halophenyleneoxides)	53	NMR and	291-312
Polyisoprene	216	survey of interpretational models	
cross polarization spectra of carbon-		of C-13 NMR relaxation in ..	271-273
black-filled	112f	synthesis of	65
FT C-13 NMR spectrum of carbon-		tacticity, C-13 NMR for determina-	
black-loaded	110f	tion of	291
high-power proton decoupling of		Polymerization	
carbon-black-loaded	110f	of 4-bromo-2,6-dichlorophenoxide	57-58
magic angle spinning of carbon-		of cyclic ethers	238-249
black-loaded	110f	C-13 NMR studies on the cationic	
<i>cis</i> -Polyisoprene, interpretation of ..	283-284	ring-opening	237-267
		chain transfer during cationic	245

- Polymerization (*continued*)
of cycle ethers (*continued*)
mechanism of 238-239
spectra of 238-239
equilibria 239
by Fourier transform NMR, kinetic
analysis of cyclic ether 264-267
kinetic of a reversible ether 254-258
mechanism, branching and 64-65
mixture of THF-CH₃OSO₂F, C-13
NMR spectra of 242f
models, spectral region for 105t
olefin-SO₂ 1
reactions, regioselectivity in 62-64
systems, C-13 NMR studies of
anionic 89-95
thermodynamic data for an
equilibrium 252-254
of THF
enthalpy of 257t
entropy of 257t
with esters of fluorosulfonic acids
in nitromethane at 0°C 257t
with OXP initiated with methyl-
triflate 258
Polypropylene (PP) 303
amorphous 311
C-13 NMR spectrum of 303
crystalline 304f
head-to-head 222
methyl, methine, and methylene
regions of the C-13 NMR
spectrum of 294f
techniques for making configura-
tional assignments in 306
Polysaccharide branching and C-13
NMR 27-50
Polysaccharide C-13 NMR spectral
analysis 29
Polystyrene (PS)
aromatic quaternary resonances
from a free-radical 296f
C-13 NMR spectrum of 293
comparisons of interpretations of
methine carbon relaxation in
dissolved 276t
effect of ionic strength on the inter-
action of opiates with 177
-enkephalin complex 174
methylene regions of the C-13
NMR spectrum of 295f
sulfones, chemical shifts of
resonances in 3-8
Polysulfone chain dynamics 1-23
Polysulfones, structure of 1-23
Polytetrahydrofuran, proton noise-
decoupled C-13 NMR spectrum
of 239
Polytetramethylene ether glycol
(PTMEG) 239
in THF, C-13 NMR spectrum of 240f
Polytrimethylene oxide, spin-lattice
relaxation time vs. temperature
for 192f
Poly(vinyl ethers), backbone methyl-
ene resonances for 299
Positions, calculated charges on allylic 95t
PP (*see* Polypropylene)
Propagation-depropagation equi-
librium 11, 239
Propylene butadiene copolymer, C-13
{¹H} NMR spectrum of 108f
Proton
-carbon dipolar coupling 69, 81f
-chlorine interaction (H-Cl) 151
decoupling of carbon-black-loaded
polyisoprene, high-power 110f
NMR, relaxation studies in the
PEMA-CHCl₃ system by 143-156
noise-decoupled C-13 NMR
spectrum of
hydrochlorinated 1,4-polydi-
methylbutadiene 231f
hydrogenated 1,4-polydimethyl-
butadiene 225f
hydrogenated 1,4-polyisoprene 223f
hydrohalogenated 1,4-polyiso-
prene 228f
polytetrahydrofuran 239
with the polymer (H-P) 151
-proton interaction (H-Hs) 151
relaxation 143
of benzene 153
spin-spin relaxation decay of
unfractionated PE melts 208
Protonated carbons 82f
Primary insertion 99
PTMEG (*see* Polytetramethylene
ether glycol)
Pulse sequence, Carr-Purcell 68
Pulsed FT C-13 NMR spectrum of
EPDM rubbers 98f
Pulsed FT C-3 NMR spectrum of
polyalkane 109f
PVC, C-13 NMR spectrum of 208f, 297f
- Q
- Quadrature detection 182-183
Quaternary
C-13 relaxation, concentration
dependence of 148f
C-13 relaxation, temperature of 148f
resonances from a free-radical PS,
aromatic 296f

R	S
Rabbit liver glycogen, C-13 NMR spectra of	SBR (<i>see</i> Styrene butadiene rubbers)
Racemic configurations	Secondary insertion
Racemic dyad configuration	Segmental motions, influence of
Receptor membranes, C-13 NMR to study binding of hormones to	crystalline regions on
Regioselectivity in the polymerization reactions	Semicrystalline polymer(s)
Relative signal intensities	C-13 spin relaxation parameters
Relaxation	of
analysis of dielectric	linewidths of
C-13	regions of chain structures within ..
decay of unfractionated polyethylene melts, proton spin-spin	Sequence assignments in the methylene region
parameters of linear PE, C-13 spin	Sequence lengths, mean
proton	for meso configurations
rates	for racemic configurations
spin-lattice (<i>see</i> Spin-lattice relaxation)	Sequence lengths, number of unique combinations for
studies in the PEMA-CHCl ₃ system	Side-chain carbon resonances, triad chemical shift sequence for vinyl polymer
time, spin-spin (T ₂)	Side-chain motions in poly(but-1-ene sulfone), correlation times for
Residual zero frequency line broadening component	Signal
Resolution in a C-13 spectrum of a cured epoxy, three stages of	averaging
Resonance(s)	intensities, absolute
in C-13 NMR spectrum of low molecular weight polymer from 2,4,6-trichlorophenoxide, assignment of	intensities, relative
from a free-radical polystyrene, aromatic quaternary	Singlet backbone methine resonances, polymers with
lines, homogeneous	SL (<i>see</i> Spin lock)
methine	SMS sequences, plot of the number fraction of
methylene	Solid(s)
to monomer sequences, assignments of	amorphous polymers, lattice models and
in PS sulfones, chemical shifts of	comparison of backbone rearrangements in
for poly(vinyl ethers), backbone methylene	high-resolution spectra of
relaxation measurements	-state spectra of epoxies
Resonant linewidths for PE and PEO	Solution spectra of elastomers, high-resolution
Reversible cyclic ether polymerization, kinetics of	Solvent
rf pulse sequences for determining SL cross-polarization time constants	C-13 relaxation and polymer concentration, relationship between
Ring-opening polymerization of cyclic ethers, C-13 NMR studies on cationic	proton-polymer proton relaxation rate, temperature and concentration dependence of
Rotation diffusion coefficients, chloroform	proton-solvent proton relaxation rate, temperature and concentration dependence of
Rotational correlation times of the glyceryl residues in methionine enkephalin	Spectral
Rubbery polymers, lattice models and	density
	vs. logarithm of frequency, logarithm of
	fidelity in cured epoxies
	narrowing
	resolution
	in cured epoxies

- Spin-lattice relaxation
of C-13 nuclei 80
parameters 190-194, 196-198
in the rotating frame 80-83
times 183
vs. temperature for
 branched polyethylene 193*f*
 linear PE 193*f*
 PEO 192*f*
 for polytrimethylene oxide 192*f*
- Spin
lock (SL) 70
 condition, cross-polarization mag-
 netization for the PIP-cured
 epoxy under 76*f*
 cross-polarization 70
 time constant, influence of
 magic angle spinning of
 time constants, rf se-
 quences for determining .. 79*f*
relaxation parameters C-13 NMR
of linear polyethylene 195*t*
relaxation in polymers, C-13
 NMR 271-286
 survey of interpretational models
 of 271-273
species, heteronuclear dipolar
 coupling between 78
-spin relaxation decay of unfrac-
 tionated polyethylene melts,
 proton 208
-spin relaxation time (T_2) .. 198-208, 272
 stirring 69
- Static magnetic field, line broadening
due to inhomogeneity in 200-201
- Stereochemical configuration, C-13
 NMR and polymer 291-312
- Steric effect, gamma 16
- Stokes-Einstein equation 176
- Styrene butadiene rubbers (SBR) 111
 high-resolution spectra of 111
 -polybutadiene rubber blend, C-13
 NMR spectra of 113-114*f*
- Styrene-SO₂ copolymer(s) 2
 C-13 spectra of 7*f*
 main chain of 6*f*
 main chain carbon chemical
 shifts in 8*t*
- Substituent(s)
 group anisotropic rotation in dilute
 solution, comparison of 281*t*
 group rotation, comparison between
 backbone dynamics and 282*t*
 internal rotation of 281-284
- Sulfur dioxide 1
- Syndiotactic configurations 291
- Synthetically branched amylose 38-40
- T**
- T_1 (spin-lattice relaxation param-
eters 190-194
 T_2 (spin-spin relaxation time) 198-208
- Tactic configurations 291
- Tacticity, C-13 NMR for determina-
tion of polymer 291
- Temperature on the ¹H spectrum of
poly(dichlorophenyleneoxide),
effects of 58
- Tetrad chemical shift sequence for
vinyl polymer backbone methyl-
ene carbon resonances 302*t*
- Tetrad concentrations 307
- Tetrahydrofuran (THF) 237
 -CD₃NO₂-Me₃OBf₄ after equili-
 bration 250*f*
 -CH₃OSO₂F, C-13 NMR spectra of
 the polymerization mixture of 242*f*
 -CH₃OSO₂F polymerization mix-
 ture in CH₃NO₂, C-13 NMR
 spectrum of 247*f*
 enthalpy of polymerization of 257*t*
 entropy of polymerization of 257*t*
 with esters of fluorosulfonic acids,
 polymerization of 244
 -methyltriflate polymerization mix-
 tures, C-13 NMR spectra of 241
 in nitromethane at 0°C, polymeriza-
 tion of 257*t*
 -OXP
 at 20°C, copolymerization of 263*t*
 -CF₃SO₃CH₃ polymerization mix-
 ture in CH₃NO₂, C-13 NMR
 spectrum of 262*f*
 copolymerization in CH₃NO₂, α
 carbon resonance region of .. 258
 with OXP initiated with methyltri-
 flate, polymerization of 258
 polymerization, determination of
 thermodynamic constants of 255*f*
 tetramer 20-crown-4 in CDCl₃, C-13
 NMR spectrum of the cyclic .. 250*f*
- Tetrahydropyran (THP) 241
- Thermodynamic data for an equi-
librium polymerization 252-254
- THF (*see* Tetrahydrofuran)
- THP (tetrahydropyran) 241
- Three-bond jump 273
- Threo diastereoisomerism 219-233
- Titanium catalyst, C-13 (¹H) NMR
spectrum of polypropylene buta-
diene copolymer made with 108*f*
- α Transition region 207
- Triad
 chemical shift sequence for vinyl
 polymer backbone methine
 carbon resonances 301*t*

Triad (<i>continued</i>)		Vanadium catalyst, C-13 $\{^1\text{H}\}$ NMR spectrum of propylene butadiene copolymer made with	108f
chemical shift sequence for vinyl polymer side-chain carbon resonances	301f	Vinyl copolymers	
concentrations	307	compositional tetrads in chains of	5f
configurational sensitivity	303	compositional triads in chains of	4f
-tetrad relationships	309	configurational dyads in chains of	4f
Trichlorophenoxide	64	configurational triads in chains of	5f
^1H NMR spectra of polymers derived from	54f	homopolymer	309
Mark-Houwink plot for polymers obtained from	55f	polymer(s)	
2,4,6-Trichlorophenoxide		backbone methylene carbon resonances, tetrad chemical shift sequence for	302t
assignment of resonances in C-13 NMR spectrum of low molecular weight polymer from	61f	backbone methine carbon resonances, triad chemical shift for	301t
and 4-bromo-2,6-dichlorophenoxide, C-13 NMR spectra of copolymers derived from	59f	C-13 NMR spectra from a series of	293
C-13 NMR spectra of polymers derived from	56f	and configurational sensitivity	293
Triflates (esters of trifluoromethane sulfonic acid)	241-245	side-chain carbon resonances, triad chemical shift sequence for	301t
Trifluoromethane sulfonic acid, esters of (triflates)	241-245	VJGM model (Valeur, Jarry, Geny, and Monnerie)	273-284
Two-level decoupling	182-183		
Two-line C-13 NMR	90		
Tyrosine in enkephalin, C-13 chemical shift of C- β of	162f		
		W	
U		Weighted harmonic average correlation time (τ_h^*)	277
Undeuterated allyl compounds	90	Weighted inverse first moment	277
Unprotonated carbons	82f		
Upfield movement of the C-13 chemical shifts	92	Z	
		Zero frequency line broadening component, residual	205
V		Ziegler catalyst system	105
Valeur, Jarry, Geny, and Monnerie (VJGM) model	273-284	Ziegler-Natta catalysts	222